



Enhancing Agricultural Sustainability through Microbial-Mediated Abiotic Stress Tolerance

Pankaj Singh¹, Fareha Rayeen², Ranjan Singh³, Neelam Pathak², Rudra Pratap Singh⁴,
Vidyanand Tiwari⁵, Manikant Tripathi^{1*} and Pradeep Kumar Singh^{2*}

¹Biotechnology Program, Dr Rammanohar Lohia Avadh University, Ayodhya - 224001, Uttar Pradesh, India

²Department of Biochemistry, Dr Rammanohar Lohia Avadh University, Ayodhya - 224001, Uttar Pradesh, India; pkbt99@gmail.com

³Department of Microbiology, Dr Rammanohar Lohia Avadh University, Ayodhya - 224001, Uttar Pradesh, India

⁴Department of Environmental Sciences, Dr Rammanohar Lohia Avadh University, Ayodhya - 224001, Uttar Pradesh, India

⁵Institute of Food Processing and Technology, University of Lucknow, Lucknow - 226007, Uttar Pradesh, India

Abstract

Global environmental problems lead to plants life extremely stressful. Plants are exposed to more prevalent incidences of abiotic stresses like salinity, drought, high temperature, etc. The most significant factors that reduce agricultural productivity are abiotic stresses. Plants are part of ecosystem entities, and the future of sustainable agriculture will be based on the exploitation of the potential of plant-associated microbial communities. Microorganisms produce significant amounts of metabolites that help plants to cope with these stresses. Plants interactions with microorganisms create a diverse ecosystem in which both partners occasionally share a cooperative relationship. This review emphasizes the plant-microbe interactions and provides a roadmap that how microorganisms such as Arbuscular Mycorrhizal Fungi, Plant Growth Promoting Rhizobacteria and endophytes are used to mitigate the negative effects of various stresses to improve crop productivity. This review also elaborates molecular and biochemical mechanisms in plants and microbes to tolerate abiotic stress. Furthermore, the most recent developments in the study of plant-microbe intermodulation with a novel approach will allow us to use a multifaceted tool “biostimulants” against abiotic stress. The important challenges of commercializing biostimulants for improving crop yield under several plant growth environmental constraints are also included in this review. As a result, the purpose of this review is to illustrate the effects of different abiotic stressors on plants, as well as the role of beneficial plant microbes in helping to overcome the negative impact of abiotic stresses.

Keywords: Abiotic stress, Biostimulants, Microbe, Mycorrhiza, PGPR, Sustainable Agriculture

1. Introduction

The most important threat to modern civilization is climate change. Worldwide, as the food demand is increasing global warming is becoming more severe. As climate change accelerates, there is a significant rise in Earth's temperature. This rise has adverse effects on crop

yields and cultivable land worldwide^{1,2}. Abiotic stresses act in synergy with biotic stresses to minimize the crop yield. Plant-microbe relations are critical components of our biosphere as they ensure agricultural sustainability. Plants are associated with a huge number of microbes including mutualists to pathogens. Positive interaction is demonstrated by mutualistic and symbiotic interactions

*Author for correspondence

with beneficial microbes while negative interaction is demonstrated by interactions with pathogenic microbes^{3,4}. Plant symbiotic microbes have been isolated from plants cultivated in both natural and harsh environmental conditions. Plant-microbial populations from extreme conditions provide hints for understanding how microbes and plants survive in extreme conditions. Beneficial microbe-plant interaction promotes plant development, crop production, and soil fertility. Endophytes (microbes that live within plant tissues without harming the plant), Plant Growth Promoting Rhizobacteria (PGPR) (microbes that colonize in the rhizosphere), and Arbuscular Mycorrhizal Fungi (AMF) can all cause changes in the host plant⁵⁻⁸. Currently, it is well known that certain potent microbial isolates plant microbial diversity, known as Plant Growth Promoting (PGP) microorganisms that improve plant fitness protect against harmful organisms, and help to maintain soil health⁹. Microbes, for example, are utilized to develop a powerful, low cost and eco-friendly tool to minimize the negative effects of extreme environmental conditions^{10,11}. Plant Growth Promoting Rhizobacteria (PGPRs) and Plant Growth-Promoting Fungi (PGPFs) are the two different microbial populations that help to remove abiotic stress¹². Bacteria in the rhizosphere typically secrete plant hormones that repress the abiotic stresses¹³. Furthermore, there is a growing interest in biostimulants to mitigate the negative effects of climate change on agriculture.

This review provides existing knowledge based on plant reactions to abiotic stresses and signalling actions.

2. Major Abiotic Stressors in Plants

Plants are constantly subjected to environmental challenges that affect their growth and yield, including both biotic (pests and viruses) and abiotic¹⁴. There are several types of abiotic stresses including salinity, drought, heavy metals, and temperature (Figure 1) that decrease crop production^{15,16}. Stress leads to changes in various physiological, biochemical, and molecular processes^{17,18}.

2.1 Salinity

Currently, saline land is rapidly increasing for a variety of reasons, including the melting of glaciers, heat stress-mediated accumulating of salt in soils¹⁹, and vigorous use of chemical fertilizers^{20,21}. These processes are predominant in coastal areas, where coastal erosion into groundwater increases soil salinization²². Furthermore, the overuse of pesticides and chemical fertilizers takes part in soil salinity, reducing both the diversity of soil microbes and plant growth and productivity^{23,24}. Several salts are required by the plants for their growth and development but, they can be toxic if consumed in high concentrations²⁵. In salt-prone soils, a suitable amount of NaCl enhances

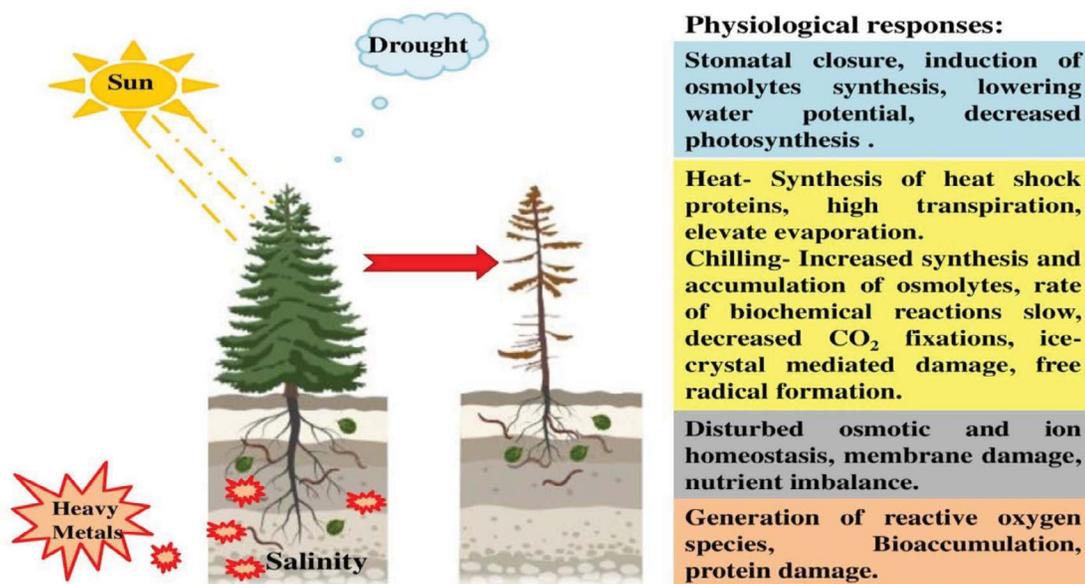


Figure 1. Various abiotic stresses and their physiological responses by the plants.

plant growth, whereas at higher concentrations inhibits seed germination and development^{26,27}. In the context of agricultural yield, moderate salinity can reduce crop yield by 50-80 per cent depending on the plant species^{28,29}, posing a serious threat to food security. It begins with stress detection by sensors, in which molecules or structures change form or lose function, triggering a signaling cascade (Figure 2) that causes a response³⁰. These sensors detect reversible physical changes (for example, changes in membrane fluidity and protein shape, as well as partial separation or melting of DNA and RNA strands), resulting in differential transcription control and stress-sensitive gene regulation¹⁷. The initial location of stress sensing is represented by the cell surface or cell membrane, and this generates changes in the cytosolic calcium (Ca^{2+}) level. Ca^{2+} is a secondary stress messenger that transmits stress signals from cell surface/membranes to effector proteins, activating other messengers/sensors like Calcineurin B-Like proteins (CBLs), Calmodulin (CaMs), Calmodulin-Like proteins (CMLs), and Calcium-Dependent Protein Kinases (CDPKs/CPKs)¹⁷.

Salinity affects plants by damaging the cell through disruption of membrane and by inhibiting the plant's physiological processes such as photosynthesis, osmoregulation, respiration, and transpiration resulting in necrosis or chlorosis^{31,32}. The disruption of ROS homeostasis, resulting in an overabundance of singlet oxygen, superoxide anion radical, hydrogen peroxide, and hydroxyl radical, is a biological response³³. Plants can cope with oxidative stress by employing a scavenging system that includes both enzymes and a non-enzymatic antioxidant including low molecular weight compounds like amino acids, phenolic compounds, Glutathione (GSH), ascorbic acid, carotenoids and α -tocopherol³⁴.

2.2 Drought

Climate change induces water scarcity and causes an agricultural threat, limiting crop productivity and thus food security. Alizadeh *et al.*,³⁵ and Lesk *et al.*,³⁶ estimated that physical dryness and ultra-high temperatures minimised worldwide cereal production by 9-10 % over the last few years. Like salt stress, drought also affects crop growth and productivity. Changes in rhizosphere physicochemical and biological properties due to drought stress hurt soil microbes and crop yield³⁷. Temperature increases above optimum cause membrane disruption, protein denaturation, DNA damage and the accumulation

of Reactive Oxygen Species (ROS), resulting in oxidative stress and ultimately plant cell death^{38,39}. Stomatal closure is the first response of plants to control water loss which disrupts respiration and photosynthetic activity⁴⁰. The stomatal closure leads to an increase in solar radiation causing a reactive oxygen species to burst provoked by water deficit, disrupting the rate of electron production⁴¹. Plants can use phytohormones to magnify the early stress signals during stress exposure. These phytohormone-related signaling events may either initiate new signaling pathways including early signals or induce new signaling pathways with diverse components^{42,43}.

2.3 Heavy Metals

Heavy metals accumulate in soil due to industrial and agricultural activities. Because of their higher density, heavy metals are lethal to plants at low concentrations⁴⁴. The composition and nature of the bedrock determine the heavy metal content in the soil. Many heavy metals (As, Cu, Cd, Cr, Pb, Hg, Ni and Zn) are now hazardous and hurt human health worldwide⁴⁵. Plants have developed a wide array of metabolic, physiologic, and genetic defence mechanisms to deal with heavy metal toxicity. The primary goal of these mechanisms is to limit the metal uptake from soil to stop heavy metal entrance into plant roots^{46,47}. Low molecular weight organic acids, such as those found in root exudates, may act as chelating agents, limiting heavy metal entry into plants⁴⁸. Furthermore, heavy metals activate detoxification and antioxidant defence mechanisms in plant tissues⁴⁹.

2.4 Temperature

In plants, temperature-driven stress is of three types: High, chilling and freezing. Global climate change affects current and future mean temperatures, as well as the risk of extreme weather events. Heat and cold are physical stresses that affect plant growth and productivity by directly influencing molecular and supramolecular structures⁵⁰. One of the most serious results of heat and cold stress is an increase in ROS production, which causes oxidative stress^{51,52}, causing damage to biomembranes, proteins, pigments and nucleic acids causing impairment of plant growth and development⁵³. Heat and cold stress also affect chlorophyll biosynthesis and photosynthesis because both have a large impact on chloroplast metabolism and structure. Heat shock, for example, disrupts the thylakoid membrane and supports grana

stacking and swelling⁵⁴, whereas low temperature causes the development of a huge thylakoid protein complex⁵⁵. Furthermore, heat and cold stresses can diminish plant water absorption which leads to dehydration⁵⁶. Plant-associated bacteria, such as PGPR, may be able to improve these responses by allowing plants more time to adapt to heat and cold stresses.

Plants also activate their response to heat stress through enzyme biosynthesis and osmolyte accumulation. Furthermore, the synthesis of Heat Shock Proteins (HSP-20, HSP-60, HSP-70, HSP-90, and HSP-100) as well as ROS scavenging enzymes allows plants to survive during brief periods of heat stress. During heat and cold stress, different signal transduction molecules are involved in stress-responsive gene activation⁵⁷. Together with transcription factors, these molecules activate stress-responsive genes. Once the stress-responsive genes are activated, they aid in the detoxification of ROS as well as the reactivation of essential enzymes and structural proteins⁵⁸.

3. Plant-Microbe Interaction Under Abiotic Stress

Several microbes are found in the rhizosphere region of plants, on leaf surfaces and other plant parts. Collectively, these microbial populations are considered plant microbiomes. These plant-linked microorganisms have a beneficial result on the plant they support plant growth and development. These microbes help the plants by increasing nutrient acquirement, granting resistance to pathogens, and increasing tolerance towards abiotic stresses including drought, heat soil salinity etc. The function and composition of the plant microbiome are regulated by environmental factors⁵⁹.

3.1 Beneficial Microbes

About ecosystem practices, plant-microbe interactions are crucial as the plant root system contains several microbial populations⁶⁰. Microbes around the roots form the niche where the microbial populations thrive

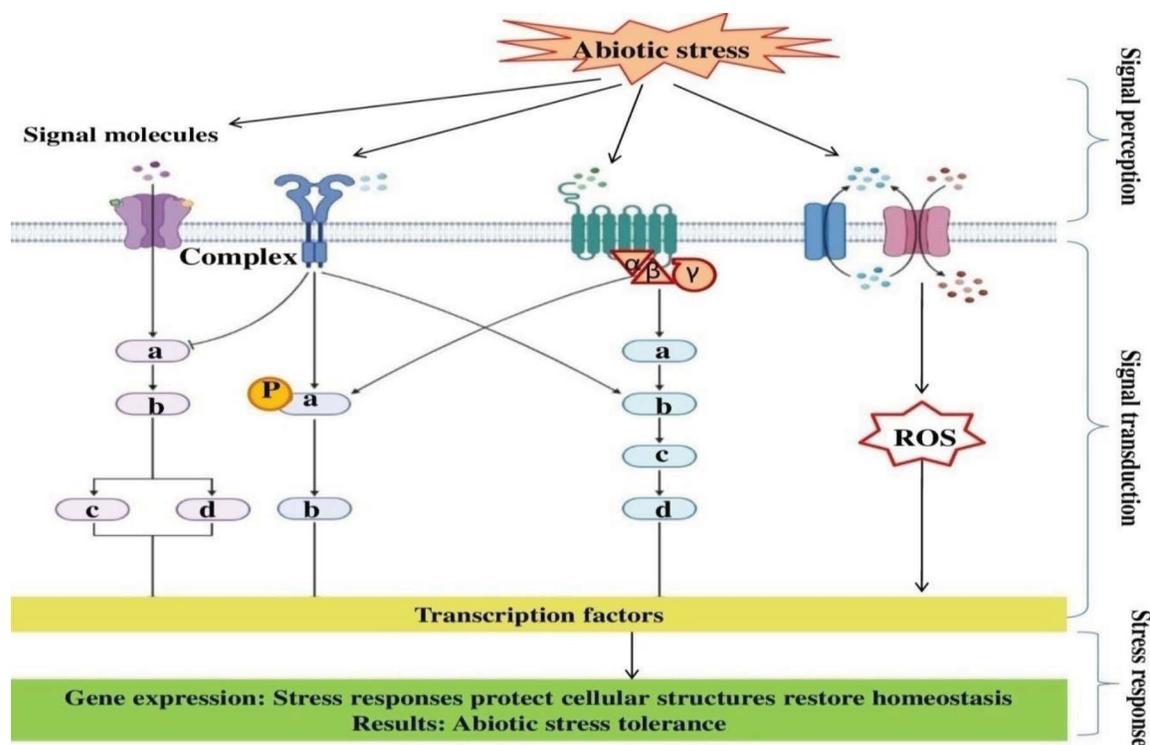


Figure 2. General abiotic stress signalling pathways in plants, starting from signal perception to stress responses. ROS-PKs (ROS-modulated protein kinase), PPs (Protein Phosphatases), MAPKs (Mitogen-Activated Protein Kinase), and CDPKs (Ca⁺-Dependent Protein Kinase) are represented as a, b, c, d.

whereas microbes are present on the leaves for example Plant Growth Promoting Microbes (PGPM) promote the nutritional condition, growth, and wellness of plants⁶¹.

PGPM is a helpful microbe that includes Arbuscular Mycorrhizal Fungi (AMF), PGPB (Plant Growth-Promoting Bacteria) and Rhizobia (PGPR)^{62,63}. Drought state, inoculation of both AMF and PGPB was used to speed up water deficit tolerance by enhancing the Glutathione Peroxidase (GPX) and Ascorbate Peroxidase (APX) accumulation in plants. The dual inoculation has proved its beneficial effect on plant metabolism^{64,65}. PGPM is supposed to offer a crucial role in controlling the genetic machinery which controls root-shoot formation during germination of seed. These microbial populations may colonize the area near the root and help to withstand the plants during various abiotic stresses like drought, salinity and ultra-high temperatures^{66,67}.

PGPR includes various groups of soil bacteria, for example, *Bacillus*, *Azobacter*, *Alcaligenes*, *Arthrobacter*, *Azospirillum*, *Bradyrhizobium*, *Burkholderia*, *Enterobacter*, *Flavobacterium*, *Klebsiella*, *Mesorhizobium*, *Pseudomonas*, *Streptomyces*, *Variovorax*, *Rhodococcus* and *Serratia* etc., that are an important part of soil-plant systems and thus affect the plant growth and development and yield. PGPR supports plant growth directly and indirectly by releasing plant hormones or other bioactive compounds, changing internal levels of plant hormones, increasing nutrient uptake, and reducing the harmful effects of pathogens on plants. PGPR are grouped into two main categories: (a) symbiotic rhizobacteria that exist in the interior of the cell (intracellular PGPR like nodule forming bacteria) and (b) free-living rhizobacteria, that are present outside of the plant cells (extracellular PGPR like *Azotobacter*)⁶⁸.

Endophytes reside inside healthy plants without causing any negative action on the host plant. Several fungal endophytes supports plant growth even with environmental limitations⁶⁹. They play a key role by providing the host plant with increased phosphorus, nitrogen, iron etc. by which the host plant defends itself from environmental stresses^{70,71}.

Similarly, AMF also assists the host plants in defeating many environmental stresses such as acidity, pathogens, desiccation and heavy metal toxicity by improved photosynthesis, nutrient uptake and gaseous exchange^{72,73}. AMF are mainly used as biofertilizers and plants make a symbiotic connection with AMF particularly under water deficit conditions by osmotic regulation^{74,75}. AMF

association can increase nutrient withdrawal by plants and thus increase the rate of photosynthesis and biomass accumulation^{76,77}.

3.2 Tolerance Mechanism by Microbes to Abiotic Stresses

Microbial association with the plant is the key adaptation, required for plant survival under extreme conditions. Microbial-mediated resistance against abiotic stresses is known as Induced Systemic Tolerance (IST). The microbiome supports vegetation to overcome such stress by utilizing its inherent metabolic properties⁷⁸. It was found that the crucial rhizospheric inhabitants that help in the removal of many plant-related abiotic stresses are from genera *Azotobacter*⁷⁹, *Azospirillum*⁸⁰, *Bacillus*, *Enterobacter*, *Rhizobium*, *Pantoea*⁸¹, *Burkholderia* and *Trichoderma*⁸², *Methylobacterium*⁸³ and the group Cyanobacteria⁸⁴. PGPRs employed both direct and indirect modes of action for plant growth and development under stress conditions. In the direct mode of action, PGPRs facilitate N₂-fixation and the production of plant regulators and organic catalysts in plants. The indirect mode of action involves antibiotics production, siderophores production and enzyme release⁸⁵.

3.2.1 Direct Mechanism of Tolerance

Nitrogen Fixation

The plant growth and yield directly rely on the presence of important nutritional elements such as nitrogen, phosphorus iron etc. N₂-fixing microorganisms are grouped into symbiotic and non-symbiotic nitrogen-fixing bacteria. Symbiotic nitrogen-fixing bacteria include leguminous (pulses) and non-leguminous plants such as *Rhizobia* and *Frankia* etc. The non-nitrogen fixing bacterium includes cyanobacteria like *Azotobacter*, *Azocarus* and *Nostoc*⁸⁶. The symbiotic association leads to the formation of root nodules in which N₂-fixation occurs efficiently⁸⁷.

Phosphate Solubilization

Plants usually face a scarcity of phosphorous under stress conditions. Both organic and inorganic form of phosphorous is naturally present in the soil⁸⁸. The deficiency of phosphorous occurs in plants because it can only be absorbed in its monobasic and dibasic ionic form⁸⁶. Phosphate-solubilizing bacteria supply the

phosphorous in the form of biofertilizers to enhance plant growth and yield. Phosphate solubilizer includes *Azotobacter*, *Burkholderia*, *Enterobacter*, *Microbacterium*, *Bacillus*, *Flavobacterium*, *Rhizobium*, *Erwinia* and *Serratia*⁸⁹.

Siderophore Production

Typically, iron is present in ferric form (Fe^{3+}) in soil. PGPRs make it soluble by the secretion of siderophores, which promotes the chelation of ferric iron (Fe^{3+}). Microbial siderophores are metal chelating agent, which ensures the iron presence in the rhizosphere of plants⁹⁰.

Phytohormone Production

Several microbes help in the biosynthesis of phytohormone auxin. Several microbes isolated from many crop plants show the potential to synthesize auxin as a secondary metabolite⁹¹. Auxin mediates an important role in the communication between rhizobacteria and plants⁹².

3.2.2 Indirect Mechanisms of Tolerance

The eco-friendly way to control plant diseases is the appliance of microorganisms. Mostly PGPRs biocontrol activity controls the onset of systemic tolerance, nutrient accessibility, and the liberation of antifungal compounds. It was reported that many rhizobacteria generate antifungal molecules or compounds like hydrogen cyanide, 2,4-diacetyl phloroglucinol, viscosinamide, pyoluteorin and pyrrolnitrin. Rhizobacteria act together with plant roots and provide resistance against pathogenic microorganisms by Induced Systemic Resistance (ISR)⁹³. The symbiotic relation of AMFs promotes the growth of plants and water availability under abiotic stress conditions⁹⁴. Similarly, endophytes help in N_2 fixation and produce plant hormones and nutrient uptake for better plant growth. During the initial phase of endophyte colonization, the bacterial cells produce exo-polysaccharides for attachment to the root surface and guard the bacterial cells against oxidative damage⁹⁵. The common mycorrhizal networks involved in the phosphorus transport and nitrogen to plants and thus improve plant growth during tense environmental conditions⁹⁶. The endophytes that promote superior growth of plants are *Bacillus pumilus* (Ps19), *Bacillus subtilis* (Ps8), *Bacillus licheniformis* (Ps14), *Lysinibacillus fusiformis* (Ps7), and *Pseudomonas putida* (Ps30), which produce plant phytohormones for example

Indole Acetic Acid (IAA), Gibberellic Acid (GA3), Zeatin and Abscisic Acid (ABA)⁹⁷.

4. Microbe-Mediated Mitigation of Abiotic Stresses

Diagne *et al.*,⁶² Liu *et al.*,⁶³ and Sangiorgio *et al.*,⁹⁸ defined that beneficial microorganisms include PGPB, AMF and rhizobia found in rhizospheres or free-living soils, or the interiors of plant tissues. Over the last few years, PGPM has been widely employed in numerous regions of the world for sustainable agriculture to limit the usage of chemical pesticides and fertilizers⁹⁹⁻¹⁰¹. It is evident that beneficial microbes, including Plant Growth-Promoting Bacteria (PGPB), rhizobia, and fungi, play a promising role in sustainable agriculture and increasing plant tolerance to abiotic stresses¹⁰² (Figure 3).

4.1 Rhizobacterial Based Mitigation

Plants have adapted in several ways to protect themselves in stressful environments and stimulate their growth and development^{103,104}. One of the most peculiar adaptations for the survival of plants in a stressed environment is a microbial relationship with the plant. The microbiome aids plants in mitigating abiotic stress by utilizing metabolic and genetic mechanisms⁷⁸. The application of useful microbes to increase tolerance for abiotic stress in plants is cheaper and more feasible¹⁰⁵⁻¹⁰⁷. Microbes on the roots create a niche for microbe populations to thrive, whereas microbes on the leaves, particularly PGPB, boost plant nutritional status, development, growth and fitness⁶¹. These soil microbes conduct abiotic stress regulation by several mechanisms simultaneously improving crop water relation and improving ion balance pathways¹⁰⁵. It was found that the number of rhizospheric microbes that are used for mitigation of abiotic stresses in plants related to the genera *Pseudomonas*¹⁰⁵, *Azotobacter*⁷⁹, *Azospirillum*⁸⁰ and the group cyanobacteria⁸⁴. PGPR helps to mitigate the negative impact of abiotic stress by the production of phytohormones, antioxidants and degradation of 1-Aminocyclopropane-1-Carboxylate (ACC) by bacterial ACC deaminase^{108,109}. Plants inoculated with PGPR expressing the enzyme ACC deaminase can help to minimize abiotic stress by controlling ethylene¹¹⁰. Microbes can boost the production of low molecular

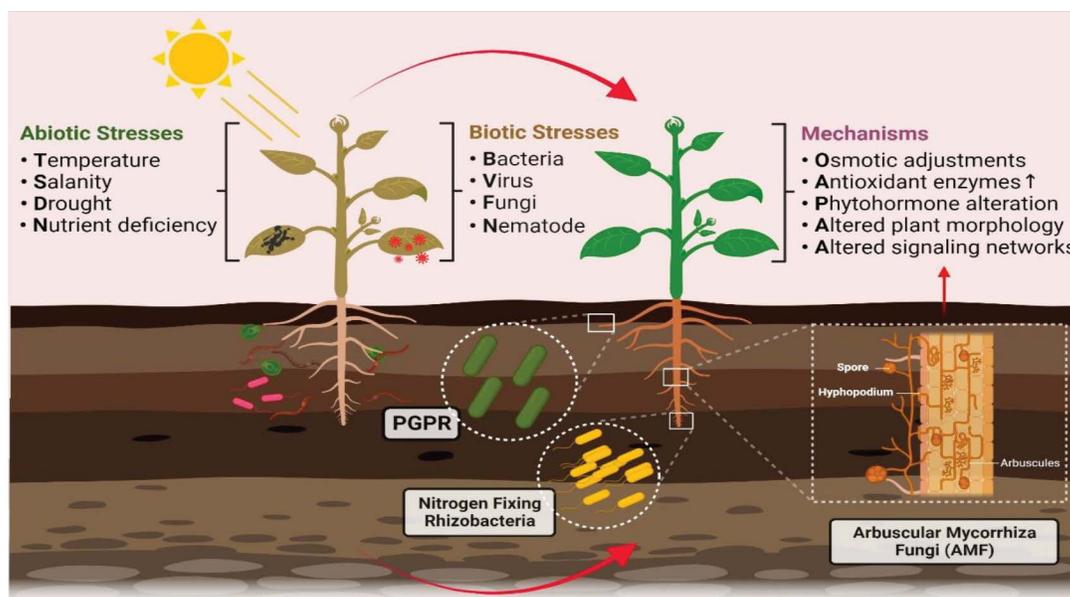


Figure 3. Mechanism of Plant Growth Promoting Rhizobacteria (PGPR) and *Arbuscular Mycorrhiza Fungi* (AMF) against abiotic stress tolerance in plants. Figure 3 is reprinted from Kamran *et al.*,¹⁰² and is an open-access article (Copyright © 2022 by authors) distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license.

weight osmoprotectives, N_2 -fixation, organic acids and mineral phosphate solubilization to face abiotic stress¹¹¹. Microbes speed up heavy metal tolerance by transporting them across the plasma membrane¹¹².

4.2 Mycorrhizae Based Mitigation

Arbuscular Mycorrhizal Fungi (AMF) is a symbiotic fungus that can also play a role in plant development and health¹¹³. More than 70% of vascular plants form symbiotic associations with AMF, specifically during dryness for osmotic adjustment and increased antioxidant enzyme activity^{74,75}. Crop yield is normally affected by drought and AMFs assist plants in retaining growth, increasing productivity and yield¹¹⁴. AMFs help in drought tolerance by ensuring continuous water intake to plants¹¹⁵.

AMF has the potential to thrive in salty environments. AMF can improve nutrient intake, assimilate carbohydrates, and reduce Cl^- and Na^+ ions in plants. AMFs can also increase stomatal conductance and reduce oxidative damage in plants exposed to salt stress¹¹¹. Al-Karaki *et al.*,¹⁰⁵ observed that when a tomato plant was inoculated with fungi *Funneliformis mosseae* under salty conditions, plant biomass increased. When wheat plants are infected with AMFs under salt conditions, oxidative damage is dramatically decreased¹¹⁶.

5. Plant Biostimulants and their Role in Abiotic Stresses

Due to uncontrolled anthropogenic activities, plants are now facing several abiotic stresses that cause harmful effects on plant growth and thus reduce plant productivity¹¹⁷. These stresses may influence the biochemical as well as physiological processes of plants which make plants more prone to damage to pests and pathogens¹¹⁸. Currently, abiotic stresses are a major risk for food safety. Under abiotic stresses, the plant produces a variety of secondary metabolites for molecular, cellular, and physiological changes to produce resistance against abiotic stress. Diverse phytochemicals or agrochemicals are being used traditionally to mitigate adverse environmental conditions and their effects¹¹⁹. Biostimulants are the products obtained from plants and/or microbes and proven their role in enhancing resistance to many abiotic stresses and supporting various physiological processes for nutrient uptake, plant quality traits and translocation in plants. Biostimulants are non-nutrient entities, and they mediate the uptake of nutrients and have a beneficial role in stress resistance or plant growth promotion¹²⁰. Currently, one of the most important and eco-friendly methods is to use biostimulants to counteract abiotic stress which have

been proposed as agronomic tools. Various raw materials from algae extracts, plant hormones (auxins, gibberellins and cytokinins), humic acids and PGPB have been used as biostimulant compositions¹²¹.

5.1 Plant Hormone as Biostimulant

Plant hormones auxins, gibberellins and cytokinins directly affect the life of plants. Auxin is the key regulator of apical dominance, cell differentiation, cell division, flowering, senescence, and abscission whereas cytokinins mainly regulate cell division, vascular development, apical dominance, and nutrient mobilization¹²². Gibberellic acid regulates the seed germination process, promotes the breakdown of seed dormancy, induction of hydrolytic enzymes α -amylase and protease and stem elongation and leaf expansion¹²³. Experimental data showed that when a combination of Indole-Butyric Acid (IBA) cytokinin and gibberellic acid is used as a biostimulant on the seed of *Gossypium hirsutum* L. plant causes an increase in the seedling emergence percentage, leaf area, height, as well as the growth of seedlings¹²⁴. Indole acetic acid and gibberellic acid are well-studied bacterial and fungal signalling molecules that are produced during plant-microbe interactions as Microbial Plant Biostimulants (MPB) to boost plant growth and tolerance to abiotic stresses^{125,126}. It has been reported that IAA improves root development in wheat with the administration of MPB *Azospirillum brasilense*¹²⁷.

5.2 Algal Extract as Biostimulant

Algal extracts as biostimulants are promising preparations to apply as plant growth-promoting factors and have beneficial effects against abiotic stresses. It considerably improved the total chlorophyll content and antioxidant compound in plants¹²⁸. Ghaderiardakani *et al.*,¹²⁹ reported that the application of algal extracts on Kentucky bluegrass (*Poa pratensis* L.) showed more salinity stress tolerance from saline soil. de Vasconcelos *et al.*,¹³⁰ reported that when a leaf of *Glycine max* (L.) is exposed to algal extract causes higher seed yield. Currently, more than 47 companies are working on algal formulations in producing and marketing for agricultural use in which brown algae (*Ascophyllum nodosum*) and red algae (*Lithothamnium calcareum*), scientists are using various algal extract formulations^{131,132}. Seaweed extracts from *A. nodosum* have been used for enhancing drought tolerance in ornamental plants (*Spiraea nipponica* and

Pittosporum eugenoides) and results showed that plants treated with *A. nodosum* extract have higher phenolic content and improved physiology under mild drought stress conditions¹³³.

5.3 Plant Parts as Biostimulants

The application of natural bioactive compounds as plant biostimulants has a profound impact on plant physiology. They trigger metabolic pathways of plant and leads to diverse expression of plant genes that are engaged in plant defense¹³⁴. It has been reported that leaf extract of *Moringa oleifera* is used as biostimulants under normal and salty conditions for plant growth. Mohamed De *et al.*¹³⁵ demonstrated that the biostimulants derived from ascorbate and *Moringa oleifera* leaf extract were shown to improve salt stress in pea plants by increasing antioxidant enzymes.

5.4 Microbes as Biostimulants

Currently, microbes as plant biostimulants are used for plant growth under stress conditions. Some microorganisms that show association with plants and increase abiotic stress tolerance have been identified and reported as *Rhizobium*, *Azospirillum*, *Bradyrhizobium*, *Azotobacter*, *Pseudomonas*, and *Bacillus*¹³⁶. Members of these genera developed tolerant mechanisms by changing cell wall composition, forming protective biofilm and accumulating high concentrations of solutes which increases water-holding capacity. Inoculation of maize and wheat with the bacterium *Azotobacter* leads to increased biomass, nitrogen content and grain yield under salt stress¹³⁷. Additionally, Bradacova *et al.*¹³⁸ revealed that zinc and manganese-containing seaweed extract applied to maize crops as biostimulants showed improved cold resistance by improved ROS scavenging systems.

There are some categories of biostimulants which may be food and industrial waste-derived extracts, manures, composts and vermicompost extracts¹³⁹. Agro-industrial by-product-derived biostimulants were also reported to be effective in improving plant productivity, and secondary metabolites synthesis which supports several plant physiological responses. Juarez-Maldonado *et al.*¹⁴⁰ reported that nanoparticles and nanomaterials are considered a new source of biostimulants. It has been found that nanoparticles and nanomaterials positively interact with plant surfaces and modulate the transportation of ions and metabolites which increases

the plant's tolerance against abiotic stresses. Raliya *et al.*,¹⁴¹ reported that the application of zinc oxide nanoparticles as a biostimulant on tomatoes increased chlorophyll and total soluble protein content as well as plant height.

6. Conclusion and Future Directions

Abiotic stresses lead to economic and social difficulties for the global population. Changes in environmental scenarios have a lethal impact on plants, resulting in reduced growth and yields. PGPB is an excellent alternative to chemical fertilizers as they offer affordability, sustainability, and long-term effectiveness in increasing plant tolerance to many abiotic stresses. Future research is used to promote sustainable agriculture by using PGPBs that can offer plant resistance towards a variety of environmental stresses. For the effective use of beneficial microbes, researchers must conduct field trials and communicate results to farmers regarding the benefits of bacteria on plant growth, soil fertility and crop yield. Nano-encapsulation, a newly designed technology is ready for field testing to improve plant tolerance. This technology has the potential to save PGPRs from environmental disturbances, increase their distribution, and help in the regulation of microbial release in the soil. Furthermore, an investigation is required to search whether the “plant-fungal-bacterial” interactions can have cumulative effects on plants. Future research should also account for the ecological fear of the large-scale use of PGPRs. Thus, it should be wrapped up that PGPRs, by various mechanisms, tolerate abiotic stresses and provide a better environment for sustainable agriculture. There is also an important role of governments and the private sector in the promotion of PGPB, PGPR, biostimulants and AMF-formulated organic fertilizers for sustainable agriculture.

7. References

- Mohanty P, Singh PK, Chakraborty D, Mishra S, Pattnaik R. Insight into the role of PGPR in sustainable agriculture and environment. *Front Sustain Food Syst.* 2021; 5. <https://doi.org/10.3389/fsufs.2021.667150>
- Shah A, *et al.* PGPR in agriculture: A sustainable approach to increasing climate change resilience. *Front Sustain Food Syst.* 2021; 5. <https://doi.org/10.3389/fsufs.2021.667546>
- Rodriguez PA, Rothballer M, Chowdhury SP, Nussbaumer T, Gutjahr C, Falter-Braun P. Systems biology of plant-microbiome interactions. *Mol Plant.* 2019; 12(6):804–21. <https://doi.org/10.1016/j.molp.2019.05.006> PMID:31128275
- Malgioglio G, *et al.* Plant-microbe interaction in sustainable agriculture: The factors that may influence the efficacy of PGPM application. *Sustainability.* 2022; 14(4):2253. <https://doi.org/10.3390/su14042253>
- Umsha S, Singh PK, Singh RP. Microbial biotechnology and sustainable agriculture, in *biotechnology for sustainable agriculture* edited by Singh RL, and Mondal S. Elsevier. (Woodhead Publishing). 2017; 185-205. <https://doi.org/10.1016/B978-0-12-812160-3.00006-4>
- Lal MK, *et al.* Mechanistic concept of physiological, biochemical, and molecular responses of the potato crop to heat and drought stress. *Plants.* 2022; 11(21):2857. <https://doi.org/10.3390/plants11212857> PMID:36365310 PMCid: PMC9654185
- Khan N, Ali S, Shahid MA, Mustafa A, Sayyed RZ, Cura JA. Insights into the interactions among roots, rhizosphere, and rhizobacteria for improving plant growth and tolerance to abiotic stresses: A review. *Cells.* 2021; 10(6):1551. <https://doi.org/10.3390/cells10061551> PMID:34205352 PMCid: PMC8234610
- Verma KK, *et al.* The interactive role of silicon and plant-rhizobacteria mitigating abiotic stresses: A new approach for sustainable agriculture and climate change. *Plants.* 2020; 9 (9):1055. <https://doi.org/10.3390/plants9091055> PMID:32824916 PMCid: PMC7569970
- Kumar M, *et al.* The synergistic effect of *Pseudomonas putida* and *Bacillus amyloliquefaciens* ameliorates drought stress in chickpeas (*Cicer arietinum* L.). *Plant Signal Behav.* 2016; 11(1):e1071004. <https://doi.org/10.1080/15592324.2015.1071004> PMID:26362119 PMCid: PMC4871671
- Jalal A, *et al.* Regulatory mechanisms of plant growth-promoting rhizobacteria and plant nutrition against abiotic stresses in Brassicaceae family. *Life.* 2023; 13(1). <https://doi.org/10.3390/life13010211> PMID:36676160 PMCid: PMC9860783
- Ait-El-Mokhtar M, Ben Laouane R, Anli M, Boutasknit A, Wahbi S, Meddich A. Use of mycorrhizal fungi in improving tolerance of the date palm (*Phoenix dactylifera* L.) seedlings to salt stress. *Sci Hortic. (Amsterdam).* 2019; 253:429–38. <https://doi.org/10.1016/j.scienta.2019.04.066>
- Evelin H, Devi TS, Gupta S, Kapoor R. Mitigation of salinity stress in plants by arbuscular mycorrhizal symbiosis: Current understanding and new challenges. *Front Plant Sci.* 2019; (10). <https://doi.org/10.3389/fpls.2019.00470> PMID:31031793 PMCid: PMC6473083
- Porcel R, Aroca R, Azcon R, Ruiz-Lozano JM. Regulation of cation transporter genes by the arbuscular mycorrhizal

- symbiosis in rice plants subjected to salinity suggests improved salt tolerance due to reduced Na (+) root-to-shoot distribution. *Mycorrhiza*. 2016; 26(7):673–84. <https://doi.org/10.1007/s00572-016-0704-5> PMID:27113587
14. Sandrini M, Nerva L, Sillo F, Balestrini R, Chitarra W, Zampieri E. Abiotic stress and below-ground microbiome: the potential of omics approaches. *Int J Mol Sci*. 2022; 23(3):1091. <https://doi.org/10.3390/ijms23031091> PMID:35163015 PMCID: PMC8835006
 15. Umar OB, *et al*. Stresses in plants: biotic and abiotic. In current trends in wheat research. *IntechOpen*, 2022. <https://doi.org/10.5772/intechopen.100501>
 16. Zhang H, Zhu J, Gong Z, Zhu JK. Abiotic stress responses in plants. *Nat Rev Genet*. 2022; 23(2):104–19. <https://doi.org/10.1038/s41576-021-00413-0> PMID:34561623
 17. Lohani N, Jain D, Singh MB, Bhalla PL. Engineering multiple abiotic stress tolerance in canola, *Brassica napus*. *Front Plant Sci*. 2020; 11. <https://doi.org/10.3389/fpls.2020.00003> PMID:32161602 PMCID: PMC7052498
 18. Nephali L, *et al*. Biostimulants for plant growth and mitigation of abiotic stresses: A metabolomics perspective. *Metabolites*. 2020; 10(12):505. <https://doi.org/10.3390/metabo10120505> PMID:33321781 PMCID: PMC7764227
 19. The Core Writing Team IPCC, Climate Change 2014: Synthesis Report. Contribution of working Groups I, II and III to the fifth assessment report of the Intergovernmental Panel on Climate Change. IPCC; 2015.
 20. Manning DAC. Mineral sources of potassium for plant nutrition. A review. *Agron Sustain Dev*. 2010; 30(2):281–94. <https://doi.org/10.1051/agro/2009023>
 21. Buvaneshwari S, *et al*. Potash fertilizer promotes incipient salinization in groundwater-irrigated semi-arid agriculture. *Sci Rep*. 2020; 10(1). <https://doi.org/10.1038/s41598-020-60365-z> PMID:32111896 PMCID: PMC7048856
 22. Balasuriya A. Coastal area management: Biodiversity and ecological sustainability in Sri Lankan perspective. *Biodivers Clim Chang Adapt Trop Islands*. 2018; 701–24. <https://doi.org/10.1016/B978-0-12-813064-3.00025-9>
 23. Banerjee S, *et al*. Agricultural intensification reduces microbial network complexity and the abundance of keystone taxa in roots. *ISME J*. 2019; 13(7):1722–36. <https://doi.org/10.1038/s41396-019-0383-2> PMID:30850707 PMCID: PMC6591126
 24. Singh PK, Singh P, Singh RP, Singh RL. Transgenesis in plants: Principle and methods. *plant genomics for sustainable agriculture*. Singapore: Springer; 2022. https://doi.org/10.1007/978-981-16-6974-3_3
 25. Nejat N, Mantri N. Plant immune system: Crosstalk between responses to biotic and abiotic stresses the missing link in understanding plant defence. *Curr Issues Mol Biol*. 2017; 23:1–16. <https://doi.org/10.21775/cimb.023.001> PMID:28154243
 26. Shabala S, Wu H, Bose J. Salt stress sensing and early signalling events in plant roots: Current knowledge and hypothesis. *Plant Sci*. 2015; 241:109–19. <https://doi.org/10.1016/j.plantsci.2015.10.003> PMID:26706063
 27. Gupta A, Rai S, Bano A, Khanam A, Sharma S, Pathak N. Comparative evaluation of different salt-tolerant plant growth-promoting bacterial isolates in mitigating the induced adverse effect of salinity in *Pisum sativum*. *Biointerface Res Appl Chem*. 2021; 11(5):13141–54. <https://doi.org/10.33263/BRIAC115.1314113154>
 28. Zorb C, Geilfus CM, Dietz KJ. Salinity and crop yield. *Plant Biol (Stuttg)*. 2019; 21(1):31–8. <https://doi.org/10.1111/plb.12884> PMID:30059606
 29. Morcillo RJL, Manzanera M. The effects of plant-associated bacterial exo-polysaccharides on plant abiotic stress tolerance. *Metabolites*. 2021; 11(6): <https://doi.org/10.3390/metabo11060337> PMID:34074032 PMCID: PMC8225083
 30. Orlando M, Molla G, Castellani P, Pirillo V, Torretta V, Ferronato N. Microbial enzyme biotechnology to reach plastic waste circularity: Current status, problems and perspectives. *Int J Mol Sci*. 2023; 24(4). <https://doi.org/10.3390/ijms24043877> PMID:36835289 PMCID: PMC9967032
 31. Rattan A, Kapoor D, Kapoor AN, Sharma A. Involvement of brassinosteroids in plant response to salt stress. *Brassinosteroids Plant Dev Biol Stress Toler*. 2022; 237–53. <https://doi.org/10.1016/B978-0-12-813227-2.00003-5>
 32. Perri S, Entekhabi D, Molini A. Plant osmoregulation as an emergent water-saving adaptation. *Water Resour Res*. 2018; 54(4):2781–98. <https://doi.org/10.1002/2017WR022319>
 33. Wani KI, Naeem M, Castroverde CDM, Kalaji HM, Albaqami M, Aftab T. Molecular mechanisms of Nitric Oxide (NO) signalling and Reactive Oxygen Species (ROS) homeostasis during abiotic stresses in plants. *Int J Mol Sci*. 2021; 22(17). <https://doi.org/10.3390/ijms22179656> PMID:34502565 PMCID: PMC8432174
 34. Choudhury FK, Rivero RM, Blumwald E, Mittler R. Reactive oxygen species, abiotic stress and stress combination. *Plant J*. 2017; 90(5):856–67. <https://doi.org/10.1111/tpj.13299> PMID:27801967
 35. Alizadeh MR, Adamowski J, Nikoo MR, AghaKouchak A, Dennison P, Sadegh M. A century of observations reveals the increasing likelihood of continental-scale compound dry-hot extremes. *Sci Adv*. 2020; 6(39). <https://doi.org/10.1126/sciadv.aaz4571> PMID:32967839 PMCID: PMC7531886
 36. Lesk C, Rowhani P, Ramankutty N. Influence of extreme weather disasters on global crop production. *Nature*. 2016; 529(7584):84–7. <https://doi.org/10.1038/nature16467> PMID:26738594
 37. Ma Y, Dias MC, Freitas H. Drought and salinity stress responses and microbe-induced tolerance in plants.

- Front Plant Sci. 2020; 11: 591911. <https://doi.org/10.3389/fpls.2020.591911> PMID:33281852 PMCID: PMC7691295
38. Hartmann M, Six J. Soil structure and microbiome functions in agroecosystems. *Nat Rev Earth Environ.* 2022; 4(1):4–18. <https://doi.org/10.1038/s43017-022-00366-w>
 39. Shekhawat K, Almeida-Trapp M, Garcia-Ramirez GX, Hirt H. Beat the heat: Plant- and microbe-mediated strategies for crop thermotolerance. *Trends Plant Sci.* 2022; 27(8): 802–13. <https://doi.org/10.1016/j.tplants.2022.02.008> PMID:35331665
 40. Agurla S, Gahir S, Munemasa S, Murata Y, Raghavendra AS. Mechanism of stomatal closure in plants exposed to drought and cold stress. *Adv Exp Med Biol.* 2018; 1081:215–32. https://doi.org/10.1007/978-981-13-1244-1_12 PMID:30288712
 41. Liu H, et al. Signaling transduction of ABA, ROS, and Ca²⁺ in plant stomatal closure in response to drought. *Int J Mol Sci.* 2022; 23. <https://doi.org/10.3390/ijms232314824> PMID:36499153 PMCID: PMC9736234
 42. Lephatsi MM, Meyer V, Piater LA, Dubery IA, Tugizimana F. Plant responses to abiotic stresses and rhizobacterial biostimulants: metabolomics and epigenetics perspectives. *Metabolites.* 2021; 11. <https://doi.org/10.3390/metabo11070457> PMID:34357351 PMCID: PMC8305699
 43. Chen K, Li GJ, Bressan RA, Song CP, Zhu JK, Zhao Y. Abscisic acid dynamics, signalling, and functions in plants. *J Integr Plant Biol.* 2020; 62(1):25–54. <https://doi.org/10.1111/jipb.12899> PMID:31850654
 44. Mavrodi DV, et al. Long-term irrigation affects the dynamics and activity of the wheat rhizosphere microbiome. *Front Plant Sci.* 2018; (9):1–15. <https://doi.org/10.3389/fpls.2018.00345> PMID:29619036 PMCID: PMC5871930
 45. Alengebawy A, Abdelkhalek ST, Qureshi SR, Wang MQ. Heavy metals and pesticide toxicity in agricultural soil and plants: Ecological risks and human health implications. *Toxics.* 2021; 9(3):42. <https://doi.org/10.3390/toxics9030042> PMID:33668829 PMCID: PMC7996329
 46. Varma S, Ekta, Jangra M. Heavy metals stress and defence strategies in plants: An overview. *J Pharmacogn Phytochem.* 2021; 10(1):608-14.
 47. Goyal, D, et al. Effect of heavy metals on plant growth: An overview. In: Naeem M, Ansari A, Gill S, editors. *Contaminants in Agriculture.* Springer, Cham; 2020. https://doi.org/10.1007/978-3-030-41552-5_4
 48. Montiel-Rozas MM, Madejon E, Madejon P. Effect of heavy metals and organic matter on root exudates (low molecular weight organic acids) of herbaceous species: An assessment in sand and soil conditions under different levels of contamination. *Environ Pollut.* 2016; 216:273–81. <https://doi.org/10.1016/j.envpol.2016.05.080> PMID:27267743
 49. Riyazuddin R, et al. A comprehensive review on the heavy metal toxicity and sequestration in plants. *Biomolecules.* 2021; 12(1):43. <https://doi.org/10.3390/biom12010043> PMID:35053191 PMCID: PMC8774178
 50. Suzuki N. Temperature stress and responses in plants. *Int J Mol Sci.* 2019; 20(8). <https://doi.org/10.3390/ijms20082001> PMID:31022827 PMCID: PMC6514902
 51. Zhang H, Zhu J, Gong Z, Zhu JK. Abiotic stress responses in plants. *Nat Rev Genet.* 2022; 23(2):104–19. <https://doi.org/10.1038/s41576-021-00413-0> PMID:34561623
 52. Ritonga FN, Chen S. Physiological and molecular mechanism involved in cold stress tolerance in plants. *Plants.* 2020; 9(5):560. <https://doi.org/10.3390/plants9050560> PMID:32353940 PMCID: PMC7284489
 53. Fahad S, et al. Crop production under drought and heat stress: Plant responses and management options. *Front. Plant Sci.* 2017; 8. <https://doi.org/10.3389/fpls.2017.01147> PMID:28706531 PMCID: PMC5489704
 54. Zhu JK. Abiotic stress signalling and responses in plants. *Cell.* 2016; 167(2):313–24. <https://doi.org/10.1016/j.cell.2016.08.029> PMID:27716505 PMCID: PMC5104190
 55. Bhattacharya A. Effect of low-temperature stress on photosynthesis and allied traits: A review. *Physiol Process. Plants Under Low Temp Stress.* 2022; 199–297. https://doi.org/10.1007/978-981-16-9037-2_3
 56. Nievola CC, Carvalho CP, Carvalho V, Rodrigues E. Rapid responses of plants to temperature changes. *Temperature.* 2017; 4(4):371–405. <https://doi.org/10.1080/23328940.2017.1377812> PMID:29435478 PMCID: PMC5800372
 57. Mittler R, Zandalinas SI, Fichman Y, Van BF. Reactive oxygen species signalling in plant stress responses. *Nat Rev Mol Cell Biol.* 2022; 23(10):663–79. <https://doi.org/10.1038/s41580-022-00499-2> PMID:35760900
 58. Haider S, et al. Molecular mechanisms of plant tolerance to heat stress: Current landscape and future perspectives. *Plant Cell Rep.* 2021; 40(12):2247–71. <https://doi.org/10.1007/s00299-021-02696-3> PMID: 33890138
 59. Xun W, Shao J, Shen Q, Zhang R. Rhizosphere microbiome: Functional compensatory assembly for plant fitness. *Computational and Structural Biotechnology Journal.* 2021; 19:5487–93. <https://doi.org/10.1016/j.csbj.2021.09.035> PMID:34712394 PMCID: PMC8515068
 60. Forni C, Duca D, Glick BR. Mechanisms of plant response to salt and drought stress and their alteration by rhizobacteria. *Plant Soil.* 2017; 410(1–2):335–56. <https://doi.org/10.1007/s11104-016-3007-x>
 61. Pascale A, Proietti S, Pantelides IS, Stringlis IA. Modulation of the root microbiome by plant molecules: the basis for targeted disease suppression and plant growth promotion. *Front Plant Sci.* 2020; 10. <https://doi.org/10.3389/fpls.2019.01741> PMID:32038698 PMCID: PMC6992662
 62. Sangiorgio D, Cellini A, Donati I, Pastore C, Onofrietti C, Spinelli F. Facing climate change: Application of microbial biostimulants to mitigate stress in horticultural crops.

- Agronomy. 2020; 10(6):794. <https://doi.org/10.3390/agronomy10060794>
63. Diagne N, Ngom M, Djighaly PI, Fall D, Hocher V, Svistoonoff S. Roles of arbuscular mycorrhizal fungi on plant growth and performance: Importance in biotic and abiotic stressed regulation. *Diversity*. 2020; 12(10):370. <https://doi.org/10.3390/d12100370>
 64. Vafadar F, Amooghaie R, Otroshy M. Effects of plant-growth-promoting rhizobacteria and arbuscular mycorrhizal fungus on plant growth, stevioside, NPK, and chlorophyll content of *Stevia rebaudiana*. *J Plant Interact*. 2014; 9(1):128–36. <https://doi.org/10.1080/17429145.2013.779035>
 65. Kumawat KC, *et al.* Dual microbial inoculation, a game changer? – Bacterial biostimulants with multifunctional growth-promoting traits to mitigate salinity stress in spring mungbean. *Front Microbiol*. 2021; 11:600576. <https://doi.org/10.3389/fmicb.2020.600576> PMID:33584566 PMCID: PMC7874087
 66. Ali S, Khan N. Delineation of mechanistic approaches employed by plant growth promoting microorganisms for improving drought stress tolerance in plants. *Microbiol Res*. 2021; 249:126771. <https://doi.org/10.1016/j.micres.2021.126771> PMID:33930840
 67. Saxena R, Kumar M, Tomar RS. Plant–rhizobacteria interactions to induce biotic and abiotic stress tolerance in plants. 2021; 1–18. https://doi.org/10.1007/978-981-16-3364-5_1
 68. Singh RP, Singh PK, Gupta R, Singh RL. *Biotechnological tools to enhance sustainable production, in biotechnology for sustainable agriculture* edited by Singh RL, and Mondal S. Elsevier. (Woodhead Publishing); 2018. p. 19-66. <https://doi.org/10.1016/B978-0-12-812160-3.00002-7>
 69. Morelli M, Bahar O, Papadopoulou KK, Hopkins DL, Obradovic A. Editorial: Role of endophytes in plant health and defence against pathogens. *Front. Plant Sci*. 2020; 11: 577603. <https://doi.org/10.3389/fpls.2020.01312> PMID:32983202 PMCID: PMC7479191
 70. Bacon CW, White JF. Functions, mechanisms and regulation of endophytic and epiphytic microbial communities of plants. *Symbiosis*. 2016; 68(1–3):87–98. <https://doi.org/10.1007/s13199-015-0350-2>
 71. White JF, *et al.* Disease protection and allelopathic interactions of seed-transmitted endophytic pseudomonads of invasive reed grass (*Phragmites australis*). *Plant Soil*. 2018; 422(1–2):195–208. <https://doi.org/10.1007/s11104-016-3169-6>
 72. Sun Z, Song J, Xin X, Xie X, Zhao B. Arbuscular mycorrhizal fungal 14-3-3 proteins are involved in arbuscular formation and responses to abiotic stresses during AM symbiosis. *Front Microbiol*. 2018; 9. <https://doi.org/10.3389/fmicb.2018.00091> PMID:29556216 PMCID: PMC5844941
 73. Riaz M, *et al.* Arbuscular mycorrhizal fungi-induced mitigation of heavy metal phytotoxicity in metal contaminated soils: A critical review. *J Hazard Mater*. 2021; 402:123919. <https://doi.org/10.1016/j.jhazmat.2020.123919> PMID:33254825
 74. Wu QS, Zou YN. Arbuscular mycorrhizal fungi and tolerance of drought stress in plants. In *Arbuscular mycorrhizas and stress tolerance of plants*, Singapore: Springer Singapore; 2017. p. 25–41. https://doi.org/10.1007/978-981-10-4115-0_2
 75. Kohl L, Lukasiewicz CE, van der Heijden MGA. Establishment and effectiveness of inoculated arbuscular mycorrhizal fungi in agricultural soils. *Plant Cell Environ*. 2016; 39(1):136–46. <https://doi.org/10.1111/pce.12600> PMID:26147222
 76. Chen S, *et al.* Combined inoculation with multiple arbuscular mycorrhizal fungi improves growth, nutrient uptake and photosynthesis in cucumber seedlings. *Front Microbiol*. 2017; 8. <https://doi.org/10.3389/fmicb.2017.02516> PMID: 29312217 PMCID: PMC5742139
 77. Li, Z. Wu N, Meng S, Wu F, Liu T. Arbuscular Mycorrhizal Fungi (AMF) enhance the tolerance of *Euonymus maackii* Rupr. at a moderate level of salinity. *PLoS One*. 2022; 15(4):e0231497. <https://doi.org/10.1371/journal.pone.0231497> PMID:32287291 PMCID: PMC7156074
 78. Vurukonda SSKP, Vardharajula S, Shrivastava M, Ali SkZ. Enhancement of drought stress tolerance in crops by plant growth promoting rhizobacteria. *Microbiol Res*. 2016; 184:13–24. <https://doi.org/10.1016/j.micres.2015.12.003> PMID:26856449
 79. Jha Y, *et al.* Bacterial-induced expression of RAB18 protein in *Oryza sativa* salinity stress and insights into molecular interaction with GTP ligand. *J Mol Recognit*. 2014; 27(9): 521–7. <https://doi.org/10.1002/jmr.2371> PMID:25042706
 80. Singh S. A review on possible elicitor molecules of cyanobacteria: Their role in improving plant growth and providing tolerance against biotic or abiotic stress. *J Appl Microbiol*. 2014; 117(5):1221–44. <https://doi.org/10.1111/jam.12612> PMID:25069397
 81. Marulanda A, Azcon R, Chaumont F, Ruiz-Lozano JM, Aroca R. Regulation of plasma membrane aquaporins by inoculation with a *Bacillus megaterium* strain in maize (*Zea mays* L.) plants under unstressed and salt-stressed conditions. *Planta*. 2010; 232(2):533–43. <https://doi.org/10.1007/s00425-010-1196-8> PMID:20499084
 82. Del-Amor FM, Cuadra-Crespo P. Plant growth-promoting bacteria as a tool to improve salinity tolerance in sweet pepper. *Funct Plant Biol*. 2012; 39(1):82–90. <https://doi.org/10.1071/FP11173> PMID:32480762
 83. Barka EA, Nowak J, Clement C. Enhancement of chilling resistance of inoculated grapevine plantlets with a plant growth promoting *Rhizobacterium*, *Burkholderia phytofirmans* Strain PsJN. *Appl Environ Microbiol*. 2006;

- 72(11):7246–52. <https://doi.org/10.1128/AEM.01047-06> PMID:16980419 PMCID: PMC1636148
84. Dardanelli MS, et al. Effect of *Azospirillum brasilense* inoculated with *Rhizobium* on *Phaseolus vulgaris* flavonoids and Nod factor production under salt stress. *Soil Biol. Biochem.* 2008; 40(11):2713–21. <https://doi.org/10.1016/j.soilbio.2008.06.016>
 85. Etesami H, Maheshwari DK. Use of Plant Growth Promoting Rhizobacteria (PGPRs) with multiple plant growth promoting traits in stress agriculture: Action mechanisms and prospects. *Ecotoxicol Environ Saf.* 2018; 156:225–46. <https://doi.org/10.1016/j.ecoenv.2018.03.013> PMID:29554608
 86. Singh SK, Wu X, Shao C, Zhang H. Microbial enhancement of plant nutrient acquisition. *Stress Biol.* 2022; 2(1):1–14. <https://doi.org/10.1007/s44154-021-00027-w> PMID:37676341 PMCID: PMC10441942
 87. Fujita Y, Uesaka K. Nitrogen fixation in cyanobacteria. *Cyanobacterial Physiol from Fundam to Biotechnol.* 2022; 29–45. <https://doi.org/10.1016/B978-0-323-96106-6.00007-1> PMID:36413373 PMCID: PMC9695160
 88. Bechtaoui N, Rabiou MK, Raklami A, Oufdou K, Hafidi M, Jemo M. Phosphate-dependent regulation of growth and stresses management in plants. *Front Plant Sci.* 2021; 12:679916. <https://doi.org/10.3389/fpls.2021.679916> PMID:34777404 PMCID: PMC8581177
 89. Kalayu G. Phosphate solubilizing microorganisms: Promising approach as biofertilizers. *Int J Agron.* 2019. <https://doi.org/10.1155/2019/4917256>
 90. Ribeiro M, Simoes M. Siderophores: A novel approach to fight antimicrobial resistance. In: Arora D, Sharma C, Jaglan S, Lichtfouse E, editors. *Pharmaceuticals from microbes. Environmental Chemistry for a Sustainable World.* Springer, Cham; 2019. https://doi.org/10.1007/978-3-030-04675-0_5
 91. Mukherjee A, et al. The bioactive potential of phytohormones: A review. *Biotechnol Reports.* 2022; 35:e00748. <https://doi.org/10.1016/j.btre.2022.e00748> PMID:35719852 PMCID: PMC9204661
 92. Upadhyay SK, et al. Root exudates: Mechanistic insight of plant growth promoting rhizobacteria for sustainable crop production. *Front. Microbiol.* 2022; 13:916488. <https://doi.org/10.3389/fmicb.2022.916488> PMID:35910633 PMCID: PMC9329127
 93. Meena M, et al. PGPR-mediated induction of systemic resistance and physiochemical alterations in plants against the pathogens: Current perspectives. *J Basic Microbiol.* 2020; 60(10):828–61. <https://doi.org/10.1002/jobm.202000370> PMID:32815221
 94. Zhang H, Zhao Y, Zhu JK. Thriving under stress: How plants balance growth and the stress response. *Dev Cell.* 2020; 55(5):529–43. <https://doi.org/10.1016/j.devcel.2020.10.012> PMID:33290694
 95. Gouda S, Das G, Sen SK, Shin HS, Patra JK. Endophytes: A treasure house of bioactive compounds of medicinal importance. *Front Microbiol.* 2016; 7:219261. <https://doi.org/10.3389/fmicb.2016.01538> PMID:27746767 PMCID: PMC5041141
 96. Begum N, et al. Role of arbuscular mycorrhizal fungi in plant growth regulation: Implications in abiotic stress tolerance. *Front Plant Sci.* 2019; 10:466052. <https://doi.org/10.3389/fpls.2019.01068> PMID:31608075 PMCID: PMC6761482
 97. Afzal I, Shinwari ZK, Sikandar S, Shahzad S. Plant beneficial endophytic bacteria: Mechanisms, diversity, host range and genetic determinants. *Microbiol Res.* 2019; 221:36–49. <https://doi.org/10.1016/j.micres.2019.02.001> PMID:30825940
 98. Liu H, Brettell LE, Qiu Z, Singh BK. Microbiome-mediated stress resistance in plants. *Trends Plant Sci.* 2020; 25(8):733–43. <https://doi.org/10.1016/j.tplants.2020.03.014> PMID:32345569
 99. Gupta A, Bano A, Rai S, Dubey P, Khan F, Pathak N, Sharma S. Plant Growth Promoting Rhizobacteria (PGPR): A sustainable agriculture to rescue the vegetation from the effect of biotic stress: A review. *Lett Appl Nano Bio Science.* 2021; 10(3):2459–65. <https://doi.org/10.33263/LIANBS103.24592465>
 100. Mahanty T, et al. Biofertilizers: A potential approach for sustainable agriculture development. *Environ Sci Pollut. Res.* 2017; 24(4):3315–35. <https://doi.org/10.1007/s11356-016-8104-0> PMID:27888482
 101. Naik K, Mishra S, Srichandan H, Singh PK, Sarangi PK. Plant growth promoting microbes: Potential link to sustainable agriculture and environment. *Biocatalysis and Agricultural Biotechnology.* Elsevier; 2019. p. 101326. <https://doi.org/10.1016/j.cbab.2019.101326>
 102. Kamran M, Imran QM, Ahmed MB, Falak N, Khatoon A, Yun BW. Endophyte-mediated stress tolerance in plants: A sustainable strategy to enhance resilience and assist crop improvement. *Cells.* 2022; 11:3292. <https://doi.org/10.3390/cells11203292> PMID:36291157 PMCID: PMC9600683
 103. Raza A, et al. Impact of climate change on crops adaptation and strategies to tackle its outcome: A review. *Plants (Basel, Switzerland).* 2019; 8(2). <https://doi.org/10.3390/plants8020034> PMID:30704089 PMCID: PMC6409995
 104. Raza A, Ashraf F, Zou X, Zhang X, Tosif H. Plant adaptation and tolerance to environmental stresses: Mechanisms and perspectives. In *plant ecophysiology and adaptation under climate change: Mechanisms and perspectives I*, Singapore: Springer Singapore. 2020; 117–45. https://doi.org/10.1007/978-981-15-2156-0_5

105. Ilangumaran G, Smith DL. Plant growth promoting rhizobacteria in amelioration of salinity stress: A systems biology perspective. *Front Plant Sci.* 2017; 8. <https://doi.org/10.3389/fpls.2017.01768> PMID:29109733 PMCID: PMC5660262
106. Etesami H, Adl, SM. Can interaction between silicon and non-rhizobial bacteria help in improving nodulation and nitrogen fixation in salinity-stressed legumes: A review. *Rhizosphere.* 2020; 15:100229. <https://doi.org/10.1016/j.rhisph.2020.100229>
107. Khan N, Bano A, Shahid MA, Nasim W, Ali MB. Interaction between PGPR and PGR for water conservation and plant growth attributes under drought conditions. *Biologia (Bratisl).* 2018; 73(11):1083–98. <https://doi.org/10.2478/s11756-018-0127-1>
108. Farooq MA, *et al.* Acquiring control: The evolution of ROS-Induced oxidative stress and redox signalling pathways in plant stress responses. *Plant Physiol. Biochem.* 2019; 141:353–69. <https://doi.org/10.1016/j.plaphy.2019.04.039> PMID:31207496
109. Porcel R, Zamarreno A, Garcia-Mina J, Aroca R. Involvement of plant endogenous ABA in *Bacillus megaterium* PGPR activity in tomato plants. *BMC Plant Biol.* 2014; 14(1):36. <https://doi.org/10.1186/1471-2229-14-36> PMID:24460926 PMCID: PMC3903769
110. Duan B, *et al.* 1-Aminocyclopropane-1-carboxylate deaminase-producing plant growth-promoting rhizobacteria improve drought stress tolerance in grapevine (*Vitis vinifera* L.). *Front Plant Sci.* 2021; 12. <https://doi.org/10.3389/fpls.2021.706990> PMID:37388278 PMCID: PMC10305780
111. Omae N, Tsuda K. Plant-microbiota interactions in abiotic stress environments. *Mol Plant-Microbe Interact.* 2022; 35(7):511–26. <https://doi.org/10.1094/MPMI-11-21-0281-FI> PMID:35322689
112. Nanda M, Kumar V, Sharma DK. Multimetal tolerance mechanisms in bacteria: The resistance strategies acquired by bacteria that can be exploited to ‘clean up’ heavy metal contaminants from water. *Aquatic Toxicol.* 2019; 212:1–10. <https://doi.org/10.1016/j.aquatox.2019.04.011> PMID:31022608
113. Sports A, *et al.* A historical perspective on mycorrhizal mutualism emphasizing arbuscular mycorrhizas and their emerging challenges. *Mycorrhiza.* 2021; 31(6):637–53. <https://doi.org/10.1007/s00572-021-01053-2> PMID: 34657204
114. Babu AG, Shea PJ, Sudhakar D, Jung IB, Oh BT. Potential use of *Pseudomonas koreensis* AGB-1 in association with *Miscanthus sinensis* to remediate heavy metal(loid)-contaminated mining site soil. *J Environ Manage.* 2015; 151:160–6. <https://doi.org/10.1016/j.jenvman.2014.12.045> PMID:25575343
115. Shen FT, Yen JH, Liao CS, Chen WC, Chao YT. Screening of rice endophytic biofertilizers with fungicide tolerance and plant growth-promoting characteristics. *Sustainability.* 2019; 11(4):1133. <https://doi.org/10.3390/su11041133>
116. Caverzan A, Casassola A, Brammer SP. Antioxidant responses of wheat plants under stress. *Genet Mol Biol.* 2016; 39(1):1–6. <https://doi.org/10.1590/1678-4685-GMB-2015-0109> PMID:27007891 PMCID: PMC4807390
117. Rai N, Rai SP, Sarma BK. Prospects for abiotic stress tolerance in crops utilizing phyto and bio-stimulants. *Front Sustain Food Syst.* 2021; 5. <https://doi.org/10.3389/fsufs.2021.754853>
118. Maharshi A, Rashid MM, Teli B, Yadav SK, Singh DP, Sarma BK. Salt stress alters the pathogenic behaviour of *Fusarium oxysporum* f. sp. ciceris and contributes to the severity of chickpea wilt incidence. *Physiol Mol Plant Pathol.* 2021; 113:101602. <https://doi.org/10.1016/j.pmpp.2021.101602>
119. Yakhin OI, Lubyantsov AA, Yakhin IA, Brown PH. Biostimulants in plant science: A global perspective. *Front Plant Sci.* 2017; 7. <https://doi.org/10.3389/fpls.2016.02049> PMID:28184225 PMCID: PMC5266735
120. du Jardin P. Plant biostimulants: Definition, concept, main categories and regulation. *Sci Hortic. (Amsterdam).* 2015; 196:3–14. <https://doi.org/10.1016/j.scienta.2015.09.021>
121. de Vasconcelos CFA, Chaves HGL. Biostimulants and their role in improving plant growth under abiotic stresses. In *Biostimulants in Plant Science*. IntechOpen; 2020. <https://doi.org/10.5772/intechopen.88829>
122. Boundless, 30.21. Plant Sensory Systems and Responses- Auxins, Cytokinins, and Gibberellins. *Biology Libre Texts*; 2022. [https://bio.libretexts.org/Bookshelves/Introductory_and_General_Biology/Book%3A_General_Biology_\(Boundless\)/30%3A_Plant_Form_and_Physiology/30.21%3A_Plant_Sensory_Systems_and_Responses_-_Auxins_Cytokinins_and_Gibberellins](https://bio.libretexts.org/Bookshelves/Introductory_and_General_Biology/Book%3A_General_Biology_(Boundless)/30%3A_Plant_Form_and_Physiology/30.21%3A_Plant_Sensory_Systems_and_Responses_-_Auxins_Cytokinins_and_Gibberellins).
123. Shah SH, Islam S, Mohammad F, Siddiqui MH. Gibberellic acid: A versatile regulator of plant growth, development and stress responses. *J Plant Growth Regul.* 2023. <https://doi.org/10.1007/s00344-023-11035-7>
124. Sosnowski J, Król J, Truba M. The effects of indole-3-butyric acid and 6-benzylaminopuryn on Fabaceae plants morphometrics. *J Plant Interact.* 2019; 14(1):603–9. <https://doi.org/10.1080/17429145.2019.1680753>
125. Kang SM, *et al.* Gibberellin-producing *Serratia nematodiphila* PEJ1011 ameliorates low-temperature stress in *Capsicum annuum* L. *Eur J Soil Biol.* 2015; 68:85–93. <https://doi.org/10.1016/j.ejsobi.2015.02.005>

126. Backer R, et al. Plant growth-promoting rhizobacteria: Context, mechanisms of action, and roadmap to commercialization of biostimulants for sustainable agriculture. *Front Plant Sci.* 2018; 9. <https://doi.org/10.3389/fpls.2018.01473> PMID:30405652 PMCID: PMC6206271
127. Saleemi M, Kiani MZ, Sultan T, Khalid A, Mahmood S. Integrated effect of plant growth-promoting rhizobacteria and phosphate-solubilizing microorganisms on the growth of wheat (*Triticum aestivum* L.) under rainfed condition. *Agric Food Security.* 2017; 6(1):46. <https://doi.org/10.1186/s40066-017-0123-7>
128. Gonzalez-Perez BK, Rivas-Castillo AM, Valdez-Calderón A, Gayosso-Morales MA. Microalgae as biostimulants: A new approach in agriculture. *World J Microbiol Biotechnol.* 2022; 38(1):4. <https://doi.org/10.1007/s11274-021-03192-2> PMID:34825262
129. Ghaderiadekani F, Collas E, Damiano DK, Tagg K, Graham NS, Coates JC. Effects of green seaweed extract on Arabidopsis's early development suggest roles for hormone signalling in plant responses to algal fertilizers. *Sci Reports.* 2019; 9:1–13. <https://doi.org/10.1038/s41598-018-38093-2> PMID:30760853 PMCID: PMC6374390
130. de Vasconcelos ACF, Chaves LHG, de Vasconcelos ACF, Chaves LHG. Biostimulants and their role in improving plant growth under abiotic stresses. *Biostimulants Plant Sci.* 2019. <https://doi.org/10.5772/intechopen.88829>
131. Lakshmi PK, Meenakshi S. Micro and macroalgae: A potential biostimulant for abiotic stress management and crop production. *New Futur Dev Microb Biotechnol Bioeng Sustain Agric Microorg as Biostimulants.* 2022; 63–82. <https://doi.org/10.1016/B978-0-323-85163-3.00001-6>
132. Hines S, van der Zwan T, Shiell K, Shotton K, Prithiviraj B. Alkaline extract of the seaweed *Ascophyllum nodosum* stimulates arbuscular mycorrhizal fungi and their endomycorrhization of plant roots. *Sci Reports.* 2021; 11(1):1–12. <https://doi.org/10.1038/s41598-021-93035-9> PMID:34188188 PMCID: PMC8241850
133. Elansary HO, Skalicka-Woźniak K, King IW. Enhancing stress growth traits as well as phytochemical and antioxidant contents of *Spiraea* and *Pittosporum* under seaweed extract treatments. *Plant Physiol Biochem.* 2016; 105:310–20. <https://doi.org/10.1016/j.plaphy.2016.05.024> PMID:27336837
134. Jamiołkowska A. Natural compounds as elicitors of plant resistance against diseases and new biocontrol strategies. *Agronomy.* 2020; 10(2):173. <https://doi.org/10.3390/agronomy10020173>
135. Mohamed De ES, Mohamed Me AR, Salah Elry A. Response of pea plants to natural bio-stimulants under soil salinity stress. *Am J Plant Physiol.* 2016; 12(1):28–37. <https://doi.org/10.3923/ajpp.2017.28.37>
136. Bastias DA, Johnson LJ, Card SD. Symbiotic bacteria of plant-associated fungi: Friends or foes? *Curr Opin Plant Biol.* 2020; 56:1–8. <https://doi.org/10.1016/j.pbi.2019.10.010> PMID:31786411
137. Oliveira DM, et al. Cell wall remodelling under salt stress: Insights into changes in polysaccharides, feruloylation, lignification, and phenolic metabolism in maize. *Plant Cell Environ.* 2020; 43(9):2172–91. <https://doi.org/10.1111/pce.13805> PMID:32441772
138. Bradacova K, et al. Micronutrients (Zn/Mn), seaweed extracts, and plant growth-promoting bacteria as cold-stress protectants in maize. *Chem Biol Technol Agric.* 2016; 3(1):1–10. <https://doi.org/10.1186/s40538-016-0069-1>
139. Xu L, Geelen D. Developing biostimulants from agro-food and industrial by-products. *Front Plant Sci.* 2018; 9. <https://doi.org/10.3389/fpls.2018.01567> PMID:30425724 PMCID: PMC6218572
140. Juarez-Maldonado A, et al. Nanoparticles and nanomaterials as plant biostimulants. *Int J. Mol Sci.* 2019; 20(1):162. <https://doi.org/10.3390/ijms20010162> PMID:30621162 PMCID: PMC6337539
141. Raliya R, Nair R, Chavalmane S, Wang WN, Biswas P. Mechanistic evaluation of translocation and physiological impact of titanium dioxide and zinc oxide nanoparticles on the tomato (*Solanum lycopersicum* L.) plant. *Metallomics.* 2015; 7(12):1584–94. <https://doi.org/10.1039/C5MT00168D> PMID:26463441