

Menthol Attenuates Cholinergic Dysfunction and Neurotransmitter Imbalance in Experimental Diabetes

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Abstract

One of the most predominant enduring consequences of Diabetes Mellitus (DM) is Diabetic Encephalopathy (DE), which has neither a reliable treatment nor an effective preventive strategy. Cognitive dysfunction is the primary problem allied with DE. The current inquiry aims to determine the potency of menthol in reducing the risk of brain complications induced by Streptozotocin (STZ) in diabetic rats. A single STZ intraperitoneal injection (40 mg/kg body weight) was employed to induce DM in Sprague-Dawley male rats and animals were held without treatment for 30 days to develop DE. The Morris water maze test, followed by the supplementation of menthol and metformin for 60 days at 50 and 100 mg/kg body weight dosages, verified the cognitive deficit in diabetic rats. After 60 days of therapy, rats were sacrificed to obtain blood and brain tissues for biochemical investigation. Oral delivery of menthol enhanced cognitive function in DE rats. Furthermore, menthol markedly reduced fasting blood sugar, glycosylated Hemoglobin (HbA1c), and elevated plasma insulin levels. In the brain, menthol increases neurotransmitter levels and choline acetyltransferase activity while decreasing AChE activity. Menthol also downregulated the expressions of monoamine oxidase A and B. Thus, the study indicates that menthol was effective in attenuating the neurodegenerative alterations in DE rats. It had a therapeutic potential and could be effectively utilized as a dietary supplement for regulating complications associated with encephalopathy.

Keywords: Cholinergic Dysfunction, Diabetic Encephalopathy (DE), Menthol, Neurotransmitters, Streptozotocin

1. Introduction

DM is characterized by hyperglycemia caused by defects in insulin secretion, insulin action, or both. A wide range of long-term problems are connected to chronic hyperglycemia. One such long-term complication of DM is DE, hallmarked by alterations in nervous system anatomy and cognitive function¹. The pathophysiology of DE has been linked to several factors, including diminished secretion or action of insulin, imbalance in glucose homeostasis, raised plasma leptin levels, elevated glucocorticoid levels, neuronal inflammation, impairment in nerve impulse transmission, oxidative stress, apoptosis, and mitochondrial malfunction as well

as variations in the morphology and functions of the hippocampus².

The cerebral cortex and hippocampus, specialized brain areas that control learning and memory, exhibit highly pronounced cholinergic neuroactivities³. Acetylcholine (ACh), the crucial neurotransmitter in the cholinergic system, is produced by choline acetyltransferase (ChAT) and cleaved by acetylcholinesterase (AChE) and mediates a significant function in learning and memory⁴. High levels of AChE are present in the brain, nerves, and RBCs. The cholinergic function is significantly influenced by the constant presence of acetylcholine, either through prolonged synthesis or decreased acetylcholinesterase activity; nevertheless, an upsurge in AChE could impact the degradation of neuronal processes⁵.

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Evidence suggests that neurotransmitter systems are a potential early pathophysiological factor in cognitive impairment⁶ and that many have been used to diagnose degenerative encephalopathy⁷. Acetylcholine and monoamine neurotransmitters are crucial for a variety of physiological processes. An upsurge or drop in the quantities of neurotransmitters has been connected to several central nervous system ailments. The progression of various diseases can be accelerated by balancing the levels of these neurotransmitters. Neurotransmitters like acetylcholine and monoamines are regulated by monoamine oxidase and cholinesterase enzymes⁸.

Monoamine oxidases are flavoenzymes facilitating the oxidative deamination of neurotransmitters, viz, serotonin, norepinephrine, tyramine, dopamine, and certain other amines⁹. A set of central nervous system diseases, including memory loss, cognitive decline, and mental disorders like depression, can be treated symptomatically with monoamine oxidase and cholinesterase inhibitors to prevent further development of the condition⁸. Due to their fewer adverse effects and better efficacy, medicinal herbs and natural phytochemicals are increasingly used to treat diabetes. Terpenoids are a broad class of naturally occurring secondary metabolites having a variety of pharmacological effects. Menthol, a naturally occurring dietary monoterpene found in mint having the chemical formula $C_{10}H_{20}O$, is a principal component of peppermint oil, an herbal remedy adopted in conventional medicine. Figure 1 shows the chemical structure of menthol.

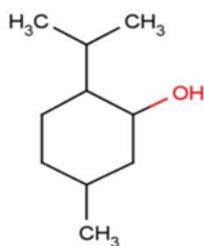


Figure 1. Chemical structure of menthol.

Various studies have demonstrated that menthol has antibacterial¹⁰, anticancer¹¹, antimicrobial¹² and anti-inflammatory properties¹³. In our study, we aimed to figure out if menthol ameliorates cognitive deficits caused by hyperglycemia-induced cholinergic dysfunction and neurotransmitter imbalances in experimental diabetes induced by streptozotocin.

2. Materials and Methods

2.1 Chemicals

The analytical grade reagents employed in the current investigation were purchased from Sisco Research Laboratories (Mumbai, India) and Sigma–Aldrich (St. Louis, Missouri, United States).

2.2 Experimental Animals

Male Sprague Dawley rats weighing 150-250 g were employed in the investigation. The animals kept in propylene cages under standard experimental conditions (25°C, 12 h light/dark cycles) were provided water and a standard laboratory pellet diet. The experimental procedure obtained approval from the Institutional Animal Ethics Committee [IAEC 3-KU-04/2018-19-BCH-SM (44)].

2.3 Induction of Diabetes

STZ (40 mg/kg body weight) is soluble in 0.1 M citrate buffer with pH 4.5 was administered intraperitoneally into rats to develop diabetes. To prevent drug-induced hypoglycemia, a 5% glucose solution was provided to the animals overnight. When the blood glucose level exceeded 250 mg/dL on the third day following the STZ injection, the rats were deemed diabetic.

2.4 Experimental Design

Five groups of six rats each were used for the study.

Group I: Normal control (N)

Group II: Normal rats administered with Menthol (N+M)

Group III: Diabetic Encephalopathic control rats (DE)

Group IV: Diabetic Encephalopathic rats administered with Menthol (DE+ M)

Group V: Diabetic Encephalopathic rats administered with Metformin (DE+ Met)

The animals in Groups II and IV were intragastrically supplemented with menthol (50 mg/kg body weight) dissolved in corn oil for 60 days. The dosage and method of administering menthol were chosen in accordance with the outcomes of the prior investigation conducted by Muruganathan *et al*¹⁴. Metformin, 100 mg/kg body weight, was supplemented in Group V. After 60 days of therapy, animals fasted for 12 hrs were sacrificed, and blood and brain were taken for various analyses.

2.5 Morris Water Maze Test (MWM Test)

The MWM test was employed to gauge the cognitive abilities of diabetic rats¹⁵. The experiment was performed in a spherical, water-filled pool maintained at a temperature of 26 ± 2 °C. Four equal quadrants were marked (I, II, III and IV) in the pool. An escape platform (10 cm diameter) was set up permanently in one of the quadrants, 1 cm below the water surface. For the experiment, rats were placed into the water from four quadrants to identify a hidden platform to escape. For four days, the rats were exposed to three trials per stage. Escaping latency (time taken to arrive escape platform) was calculated. The average latency time for all three trials was determined. A substantial decrease in latency time was considered successful learning.

2.6 Biochemical Studies

2.6.1 Serum Glucose and Hepatic Toxicity Markers

The blood glucose, Aspartate Transaminase (AST), Alanine Transaminase (ALT) and Alkaline Phosphatase (ALP) were assessed by kits obtained from Agappe Diagnostics Pvt. Ltd, India.

2.6.2 Glycated Hemoglobin (HbA1c) and Plasma Insulin by ELISA

Glycated hemoglobin was measured by means of an HbA1c kit from Beacon Diagnostics Pvt Ltd., and plasma insulin using an ELISA kit from DRG Diagnostics, Marburg, Germany.

2.6.3 Acetylcholinesterase and Choline Acetyltransferase (ChAT) Activity

AChE activity and ChAT activity were determined

according to the protocol of Ellman *et al.*,¹⁶ and Liu *et al.*,¹⁷ respectively.

2.6.4 Neurotransmitters

The brain Norepinephrine (NE) was assayed by the protocol of Weil-Malherbe and Bigelow¹⁸, Dopamine (DA) by the technique of Atack¹⁹ and serotonin (5-HT) by means of the procedure of Curzon and Green²⁰.

2.6.5 Determination of Protein

The protein concentration was measured according to Lowry *et al.*²¹.

2.6.6 RNA Isolation and Reverse Transcriptase Polymerase Chain Reaction

TRI Reagent was adopted for the isolation of total genomic RNA from the brain (Sigma-Aldrich, St. Louis, MO, USA)²². cDNA synthesis and PCR reactions were carried out in a thermocycler (Eppendorf 5332) in accordance with the manufacturer's directions employing a kit acquired from Fermentas, Vilnius, Lithuania. The primer sequences utilized for the investigation are displayed in Table 1. The PCR products were electrophoresed on 1.5% agarose gels, and the gels were then scanned densitometrically using Bio-Rad Gel Doc to visualize the products. The mRNA level was quantified using the quantity one imaging software (Bio-Rad). In the same sample, the relative expression was compared to the expression of β actin.

2.6.7 Statistical Analysis

SPSS/PC+, Version 17 (SPSS Inc. Chicago, IL, USA), the statistical package, was employed for statistical analysis. Comparison among the groups was accomplished by one-way Analysis of Variance (ANOVA) and, subsequently,

Table 1. PCR primer sequences

Gene	Forward primer (5'-3')	Reverse primer (3'-5')
β actin	CCCATCTATGAGGGTTACGC	TTTAATGTCACGCACGATTTTC
MAO A	AAGACACGCTCAGGAATGGG	ATGGCTACGTACATGGCTGG
MAO B	TTTGCCAGCCAGAACCAGAA	AGCTTGTGTTCCAGTCACCC

Duncan's posthoc multiple comparison tests. The results were denoted as mean value \pm SE ($n = 6$). ' p ' values less than 0.05 were considered significant.

3. Results

3.1 Morris Water Maze Test

Spatial learning and memory in laboratory rats were investigated by adopting the MWM test. After thirty days following STZ induction, animals in untreated diabetic groups displayed greater escape latencies during training sessions than the healthy control rats (Figure 2A). It demonstrated cognitive impairment. Menthol and metformin were then administered to the diabetic rats for 60 days. In four successive trials, the normal healthy groups performed better on the MWM test. Throughout training sessions, the escape latencies of STZ-induced rats were longer than those of the normal rats. In comparison to diabetic controls, menthol, and metformin supplementation significantly ($p < 0.05$) diminished mean escape delay time (Figure 2B). These findings demonstrated that the diabetic group treated with menthol and metformin significantly improved their spatial learning abilities compared to the diabetic encephalopathic group.

3.2 Fasting Blood Glucose, HbA1c and Plasma Insulin Levels

Blood glucose and HbA1c levels were significantly higher in the DE control groups than in normal rats, but plasma

insulin levels were lower. Administration of menthol and metformin reduced blood glucose and HbA1c and augmented plasma insulin levels in DE rats (Figure 3). A significant effect was noticed for blood glucose and plasma insulin in menthol supplemented group than metformin ($p < 0.05$). Nevertheless, for HbA1c, the results were comparable. In normal and normal rats treated with menthol, there was no substantial change in blood glucose, HbA1c and plasma insulin. These results indicate that menthol offers protection to the pancreas in diabetic rats and enhances insulin release from pancreatic β cells.

3.3 Liver Toxicity Markers

Several markers, including AST, ALT and ALP, are principal parameters representing the health of the fundamental liver function, and variations in these parameters can reveal the degree of hepatocytic damage²³. As shown in Table 2, the liver enzymes AST, ALT, and ALP were elevated in DE rats. In comparison to diabetic control, menthol, and metformin supplementation significantly ($p < 0.05$) diminished AST, ALT and ALP activities (Table 2). Between the N and N+M groups, there were no significant alterations in the activities of hepatic toxicity markers. A superior effect was observed for menthol than metformin. Therefore, menthol promotes protection by averting the outflow of hepatic enzymes into the bloodstream without any toxic effects.

Values are represented as mean \pm SE ($n = 6$). 'a' depicts values that differed significantly from N, DE is contrasted to DE + M and DE + Met ('b' depicts values that varied significantly from DE and DE + M is a contrasted with

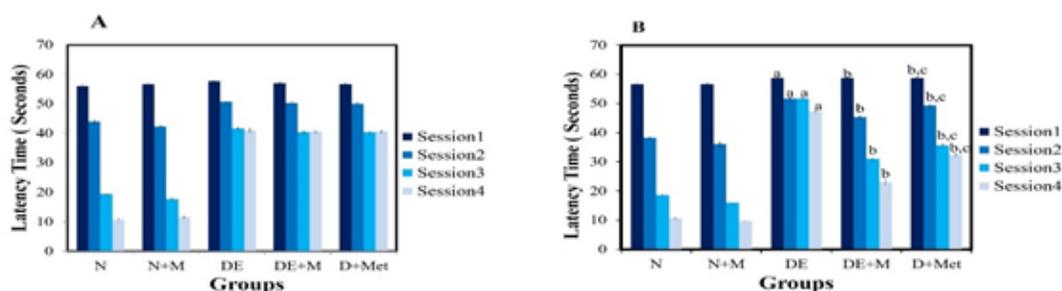


Figure 2. Learning and memory. (A) MWM test before treatment, (B) MWM test after treatment. Values are represented as mean \pm SE ($n = 6$). 'a' depicts values that differed significantly from N, DE is contrasted to DE+M and DE+Met ('b' depicts values that differed significantly from DE and DE+M is a contrasted with DE+met ('c' depicts values that differed significantly from DE+M. Significance accepted at $p < 0.05$.

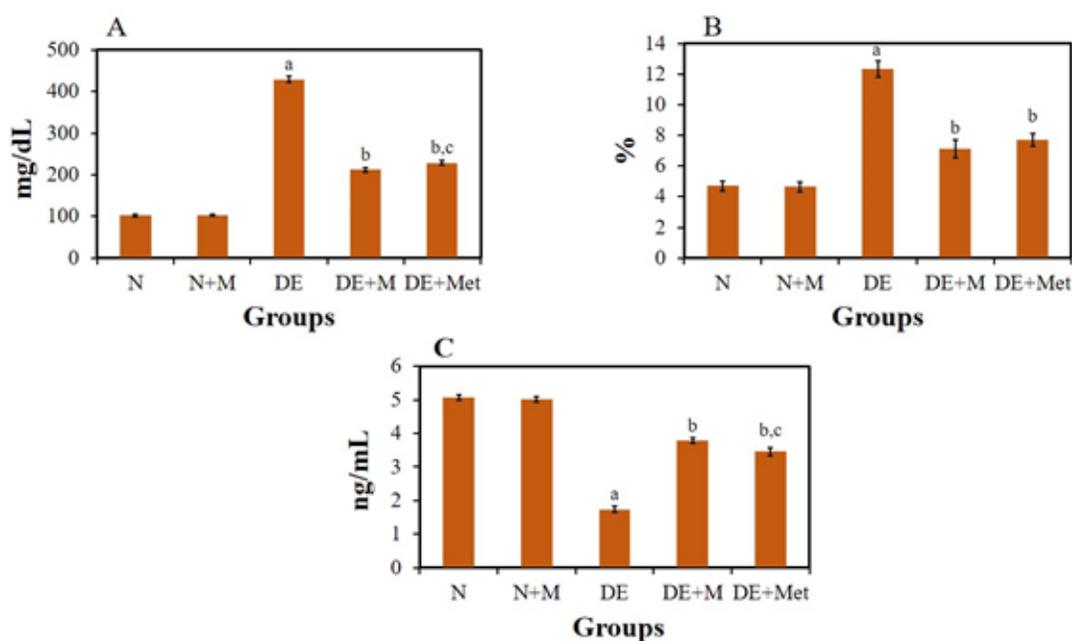


Figure 3. (A) Serum glucose, (B) Glycated hemoglobin, (C) Plasma insulin. Values are represented as mean \pm SE (n = 6). 'a' depicts values that differed significantly from N, DE is contrasted to DE + M and DE + Met 'b' depicts values that differed significantly from DE and DE + M is a contrasted with DE + met 'c' depicts values that differed significantly from DE + M. Significance accepted at $p < 0.05$.

Table 2. Liver toxicity markers

Group	SGPT (U/L)	SGOT (U/L)	ALP(U/L)
N	43.05 \pm 2.67	54.09 \pm 1.75	8.53 \pm .28
N+M	43.04 \pm 2.05	54.68 \pm 3.63	8.40 \pm .35
DE	83.76 \pm 4.61 ^a	102.96 \pm 1.75 ^a	19.71 \pm .42 ^a
DE+M	53.51 \pm 2.97 ^b	70.97 \pm 2.67 ^b	10.36 \pm .42 ^b
DE+Met	62.12 \pm 4.58 ^{b,c}	76.78 \pm 1.75 ^{b,c}	12.29 \pm .58 ^{b,c}

DE + Met ('c' depicts values that varied significantly from DE+M. Significance accepted at $p < 0.05$.

3.4 Cholinergic Function

ChAT and AchE activities are depicted in Figure 4. Compared to the normal control rats, DE animals exhibited a noticeable reduction in ChAT activity and an upsurge in AchE activity. In STZ-induced DE rats, ChAT activity was considerably ($p < 0.05$) elevated, and AchE activity was lowered following menthol or metformin

therapy. Both menthol and metformin produced comparable effects.

3.5 Neurotransmitters

Monoaminergic neurotransmitter systems are crucial for neuronal function and behaviour. They were significantly affected by the disruption of glucose homeostasis. In our study, monoaminergic neurotransmitters such as dopamine, serotonin, and norepinephrine declined in DE rats more than in normal groups. However, menthol

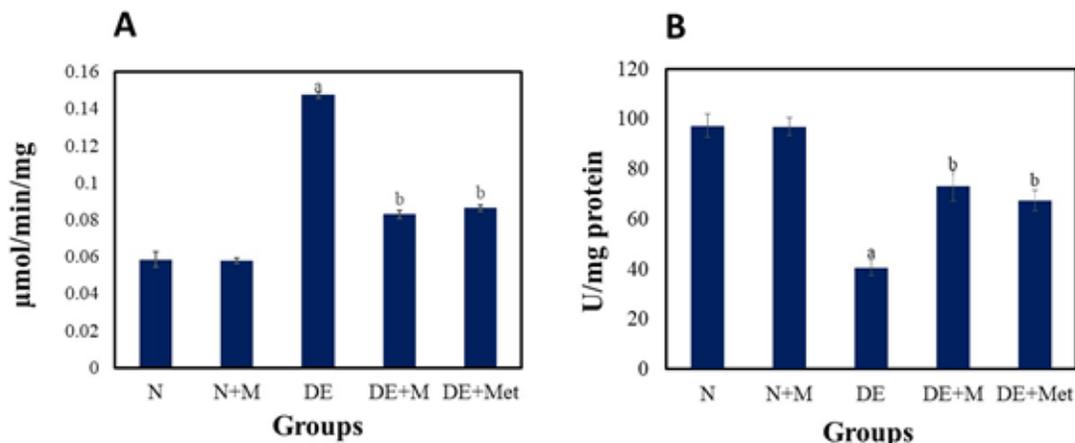


Figure 4. (A) Acetylcholinesterase, (B) Choline acetyl transferase. Values are represented as mean ± SE (n = 6). ‘a’ depicts values that differed significantly from N, DE is contrasted to DE+M and DE+ Met (‘b’ depicts values that varied significantly from DE). Significance accepted at $p < 0.05$.

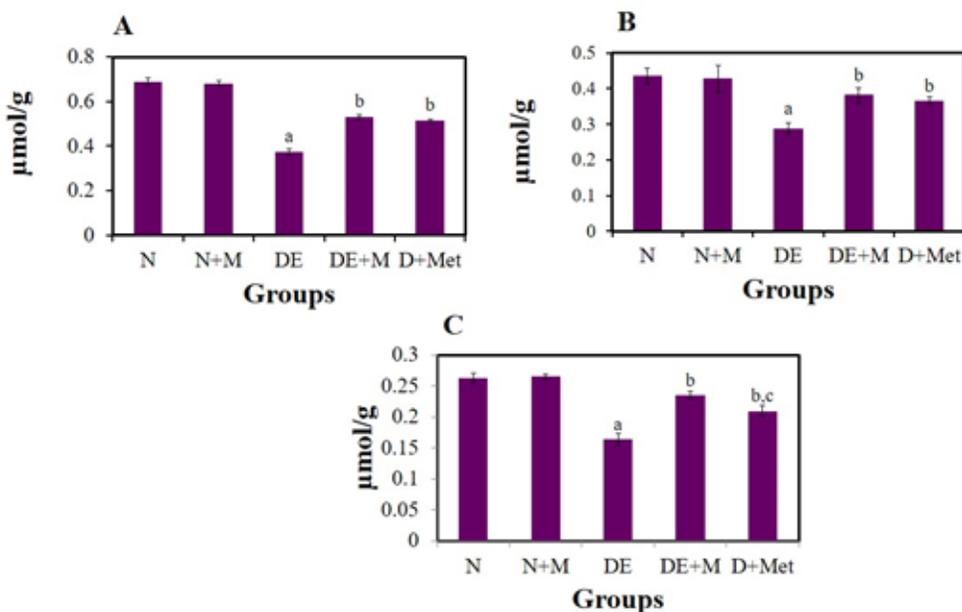


Figure 5. (A) Dopamine, (B) Serotonin, (C) Norepinephrine. Values are represented as mean ± SE (n = 6). ‘a’ depicts values that differed significantly from N, DE is contrasted to DE + M and DE + Met (‘b’ depicts values that differed significantly from DE, and DE + M is a contrasted with DE + Met (‘c’ depicts values that differed significantly from DE + M). Significance accepted at $p < 0.05$.

and metformin treatment markedly raised the level of these neurotransmitters (Figure 5). Both menthol and metformin displayed comparable effects except for norepinephrine, where menthol displayed a superior effect.

3.6 mRNA Expressions of MAO A and MAO B

In the present work, DE rats exhibited upregulated MAO A and MAO B expressions compared to normal rats. However, MAO A and B expressions were markedly reduced after menthol and metformin administration.

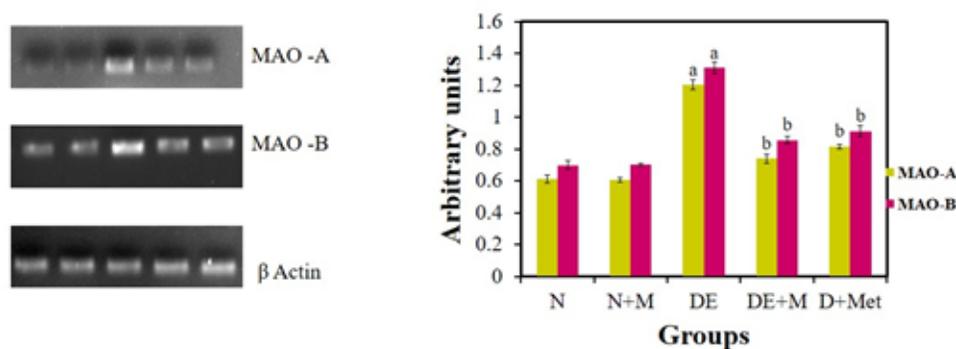


Figure 6. mRNA expressions of monoamine oxidase A and B. Values are represented as mean \pm SE (n = 6). 'a' depicts values that differed significantly from N, DE is contrasted to DE + M and DE + Met ('b' depicts values that differed significantly from DE). Significance accepted at $p < 0.05$.

In DE rats treated with menthol and metformin, there was no discernible difference between the expressions of MAO A and MAO B (Figure 6).

4. Discussion

DM is a metabolic condition marked by persistent hyperglycemia resulting from poor insulin production, insulin action, or both. One of the most alarming microvascular complications of DM is diabetic encephalopathy²⁴. It is distinguished by cognitive impairment caused by neurochemical and structural alterations.

Treatment of CNS complications by modern medicine has several shortcomings because of their strong side effects and higher chances of reoccurrence. As a result, it is essential to look for safer and more effective drugs, and phytochemicals may be preferable. Secondary plant metabolites with various structural and functional properties have recently received much attention as potential therapeutic leads for the therapy and avoidance of dementia caused by diabetes. In the current study, the neuroprotective effect of dietary monoterpene menthol in STZ-induced diabetic rats was determined.

MWM is a popular tool for evaluating spatial learning and memory²⁵. Increased escape latency values in DE rats in the current study revealed that diabetes considerably reduced learning and memory function, which is in agreement with the outcomes of earlier investigations^{26,27}. However, the escape latency values of DE rats fed with

menthol and metformin were diminished significantly compared with the DE rats. These findings suggested that menthol administration improves learning and memory in DE rats. The ameliorative potential of menthol might be attributable to an improvement in acetylcholine in the brain²⁸.

The hallmark of T2D is chronic hyperglycemia, caused by insulin resistance and beta cell malfunction and linked to long-term consequences such as retinopathy, nephropathy, and micro- and macrovascular disorders. Blood sugar, HbA1c, and insulin are the three leading glycemic indicators of diabetes²⁹. In the present investigation, blood glucose and HbA1c levels were significantly raised, and plasma insulin levels declined in DE groups than in normal rats administered with menthol. Menthol and metformin supplementation decreased blood glucose and HbA1c levels. Furthermore, they also elevated plasma insulin levels (Figure 3). Menthol showed a superior effect on both blood glucose and plasma insulin. Nevertheless, for HbA1c, the results were comparable for menthol and metformin. These outcomes align with previous studies by Muruganathan *et al.*, which revealed that menthol supplementation stimulated insulin release from the rejuvenated pancreatic β -cells, which consequently raises glucose consumption by the tissues¹⁴.

The cholinergic system is crucial in promoting neuronal plasticity and is essential for neuronal function in memory, learning, and cognition³⁰. Cognitive processes are greatly influenced by normal cholinergic

function, maintained by the constant presence of acetylcholine, either by sustained synthesis or reduced acetylcholinesterase activity³¹. The two distinct indicators of cholinergic neurons in the hippocampus and cerebral cortex, AChE, and ChAT, are crucial for controlling the cholinergic pathway. The outcomes of the present investigation demonstrated an upsurge in AChE and a decline in ChAT activity in the diabetic encephalopathic group, representing inadequate cholinergic action because of the depletion of acetylcholine in the brain. Rats in the menthol-supplemented group exhibited diminished AChE activity and augmented ChAT activity. Similar results were seen in the metformin-treated group as well. Our results agree with the previous study by Mao *et al.*, where huperzine A ameliorates cognitive deficits by augmented ChAT activity and decreased AChE activity in STZ-induced diabetic rats³². The observed reduction in AChE activity and an upsurge in ChAT activity could avert the quick degradation of acetylcholine, thereby increasing its availability for nerve impulse transmission. It is crucial to inhibit AChE to maintain memory function as its improved activity leads to the development of amyloid plaques, another mechanism causing cognitive decline^{33,34}.

Neurotransmitters are chemical molecules that may convey and amplify impulses and are essential for transferring information throughout the neurological system³⁵. One of the central neurotransmitters in the mammalian nervous system, dopamine, enhances learning, locomotion, sexual fulfilment, motivation, attention, and arousal³⁶. Additionally, it plays a role in sustaining homeostasis and acts as a precursor for norepinephrine and epinephrine³⁷. Serotonin regulates brain function, behaviour, vasoconstriction, peristalsis, gastrointestinal secretion, sleep and waking states, and respiration³⁸. Norepinephrine is recognized to play a role in waking-state arousal and alertness, emotion regulation, sensory signal recognition, memory, learning, and attention. DA, 5-HT, and NE are monoaminergic neurotransmitters that are crucial for neuronal behaviour and function. Neuropsychiatric and neurodegenerative illnesses, often known as monoamine neurotransmitter disorders, are caused by the dysregulation of these systems³⁹. The current study observed that diabetic rats had lower amounts of monoaminergic neurotransmitters such as DA, 5-HT, and NE than normal control rats. These outcomes align with the previous study by Ezzeldin

*et al.*⁴⁰. On the other hand, menthol and metformin treatment significantly improved the level of DA, 5-HT, and NE and thus played an essential role in solving many neuropathologies associated with DE. Our findings agree with those of Wang *et al.*, who found that the modulation of the 5-hydroxytryptaminergic, GABAergic, and dopaminergic systems by the antidepressant-like actions of menthol results in the maintenance of neurotransmitters⁴¹.

Monoamine oxidases play a pivotal function in the brain through the metabolic control of monoamine neurotransmitters⁴². They catalyze the oxidative deamination of epinephrine, norepinephrine, and dopamine and produce hydrogen peroxide (H₂O₂)⁴³. Combined with metal ions, H₂O₂ will produce Reactive Oxygen Species (ROS) in the brain of AD rats⁴⁴. Increased ROS-mediated oxidative stress brought on by A β accumulation and deposition will eventually cause neurons to die⁴⁵. In our study, mRNA expressions of monoamine oxidases (A and B) were significantly upregulated in the diabetic encephalopathic group. This was consistent with the study conducted by Adefegha *et al.*⁴⁶. This finding could suggest that DA, 5-HT, and NE were destroyed in DE rats' brains. However, compared to DE rats, menthol and metformin administration in DE rats significantly downregulated the expression of MAO A and B. This downregulated MAO-A and B expression suggested that menthol had MAO inhibitory effects and may confer neuroprotection. In addition, our results concur with those of Emory and Mizrahi, who found that inhibiting MAO reduced dopamine deficiency in diabetic patients and depression⁴⁷. The findings of the current investigation suggest that menthol could improve brain cholinergic function and regulate the level of neurotransmitters in experimental diabetic rats.

5. Conclusion

To date, cognitive deterioration unquestionably represents a novel and developing long-term DM consequence, resulting in poor glycemic control and a lack of diabetes self-management. Scientists, researchers, and pharmaceutical companies are increasingly searching for bioactive compounds to discover and develop targeted novel anti-diabetic drugs. Compared to conventional diabetes medications, these compounds may be less likely to cause side effects. The present

study revealed that menthol treatment ameliorates high glucose-induced cognitive impairment and attenuates cholinergic dysfunction and neurotransmitter imbalance in experimental diabetes. Thus, menthol seems to be a promising compound to prevent the complications associated with diabetic encephalopathy.

6. Acknowledgments

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7. References

- Dong M, Ren M, Li C, Zhang X, Yang C, Zhao L, *et al.* Analysis of metabolic alterations related to pathogenic process of diabetic encephalopathy rats. *Front Cell Neurosci.* 2019; 12:1-17. <https://doi.org/10.3389/fncel.2018.00527>
- Han W-N, Hölscher C, Yuan L, Yang W, Wang X-H, Wu M-N, *et al.* Liraglutide protects against amyloid- β protein-induced impairment of spatial learning and memory in rats. *Neurobiol Aging.* 2013; 4(2):576-88. <https://doi.org/10.1016/j.neurobiolaging.2012.04.009>
- Mesulam M-M, Guillozet A, Shaw P, *et al.* Acetylcholinesterase knockouts establish central cholinergic pathways and can use butyrylcholinesterase to hydrolyze acetylcholine. *Neuroscience.* 2002; 110(4):627-39. [https://doi.org/10.1016/S0306-4522\(01\)00613-3](https://doi.org/10.1016/S0306-4522(01)00613-3)
- Hegazy AM, Azeem AA, Shahy EM, El-Sayed EM. Comparative study of cholinergic and oxidative stress biomarkers in brains of diabetic and hypercholesterolemic rats. *Hum Exp Toxicol.* 2016; 35(3):251-8. <https://doi.org/10.1177/0960327115583361>
- Tiwari V, Kuhad A, Chopra K. Suppression of neuro-inflammatory signaling cascade by tocotrienol can prevent chronic alcohol-induced cognitive dysfunction in rats. *Behav Brain Res.* 2009; 203(2):296-303. <https://doi.org/10.1016/j.bbr.2009.05.016>
- Prakash A, Kalra J, Mani V, Ramasamy K, Majeed ABA. Pharmacological approaches for Alzheimer's disease: Neurotransmitter as drug targets. *Expert Rev Neurother.* 2015; 15(1):53-71. <https://doi.org/10.1586/14737175.2015.988709>
- Levenga J, Krishnamurthy P, Rajamohamedsait H, Wong H, Franke TF, Cain P, *et al.* Tau pathology induces loss of GABAergic interneurons leading to altered synaptic plasticity and behavioral impairments. *Acta Neuropathol Commun.* 2013; 1:34. <https://doi.org/10.1186/2051-5960-1-34>
- Aksoz BE, Aksoz E. Vital role of monoamine oxidases and cholinesterases in central nervous system drug research: A sharp dissection of the pathophysiology. *Comb Chem High Throughput Screen.* 2020; 23(9):877-86. <https://doi.org/10.2174/1386207323666200220115154>
- Manzoor S, Hoda N. A comprehensive review of monoamine oxidase inhibitors as Anti-Alzheimer's disease agents: A review. *Eur J Med Chem.* 2020; 206. <https://doi.org/10.1016/j.ejmech.2020.112787>
- Hajlaoui H, Snoussi M, Jannet HBEN, Mighri Z, Bakhrouf A. Comparison of chemical composition and antimicrobial activities of *Mentha longifolia* L. ssp. *longifolia* essential oil from two Tunisian localities (Gabes and Sidi Bouzid). *Ann Microbiol.* 2008; 58(3):513-20. <https://doi.org/10.1007/BF03175551>
- Kim S, Lee S, Piccolo SR, Karuppayil SM. Menthol induces cell-cycle arrest in PC-3 cells by down-regulating G2/M genes, including polo-like kinase 1. *Biochem Biophys Res Commun.* 2012; 422(3):436-41. <https://doi.org/10.1016/j.bbrc.2012.05.010>
- Raut JS, Shinde RB, Chauhan NM, Karuppayil SM. Terpenoids of plant origin inhibit morphogenesis, adhesion, and biofilm formation by *Candida albicans*. *Biofouling.* 2013; 29(1):87-96. <https://doi.org/10.1080/08927014.2012.749398>
- Baylac S, Racine P. Inhibition of 5-lipoxygenase by essential oils and other natural fragrant extracts. *Int J Aromather.* 2003; 13(2-3):138-42. [https://doi.org/10.1016/S0962-4562\(03\)00083-3](https://doi.org/10.1016/S0962-4562(03)00083-3)
- Muruganathan U, Srinivasan S, Vinothkumar V. Antidiabetogenic efficiency of menthol, improves glucose homeostasis and attenuates pancreatic β -cell apoptosis in streptozotocin-nicotinamide induced experimental rats through ameliorating glucose metabolic enzymes. *Biomed Pharmacother.* 2017; 92:229-39. <https://doi.org/10.1016/j.biopha.2017.05.068>
- Morris R. Developments of a water-maze procedure for studying spatial learning in the rat. *J Neurol Meth.* 1984; 11:47-60. [https://doi.org/10.1016/0165-0270\(84\)90007-4](https://doi.org/10.1016/0165-0270(84)90007-4)
- Ellman GL, Courtney KD, Andres V, Featherstone RM. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem Pharmacol.*

- 1961; 7(2):88-95. [https://doi.org/10.1016/0006-2952\(61\)90145-9](https://doi.org/10.1016/0006-2952(61)90145-9)
17. Liu J, Feng L, Zhang M, Ma D-y, Wang S-y, Gu J, *et al.* Neuroprotective effect of Liuwei Dihuang decoction on cognition deficits of diabetic encephalopathy in streptozotocin-induced diabetic rat. *J Ethnopharmacol.* 2013; 150(1):371-81. <https://doi.org/10.1016/j.jep.2013.09.003>
 18. Weil-Malherbe H, Bigelow LB. The fluorometric estimation of epinephrine and norepinephrine: An improved modification of the trihydroxyindole method. *Anal Biochem.* 1968; 22(2):321-34. [https://doi.org/10.1016/0003-2697\(68\)90322-9](https://doi.org/10.1016/0003-2697(68)90322-9)
 19. Atack CV. The determination of dopamine by a modification of the dihydroxyindole fluorimetric assay. *Br J Pharmacol.* 1973; 48(4):699-714. <https://doi.org/10.1111/j.1476-5381.1973.tb08259.x>
 20. Curzon G, Green AR. Rapid method for the determination of 5-hydroxytryptamine and 5-hydroxyindoleacetic acid in small regions of rat brain. *Br J Pharmacol.* 1970; 39(3):653-5. <https://doi.org/10.1111/j.1476-5381.1970.tb10373.x>
 21. Lowry H, Randall RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem.* 1951; 193(1):265-75. [https://doi.org/10.1016/S0021-9258\(19\)52451-6](https://doi.org/10.1016/S0021-9258(19)52451-6)
 22. Skrypina NA, Timofeeva A V, Khaspekov GL, Savochkina LP, Beabealashvilli RS. Total RNA suitable for molecular biology analysis. *J Biotechnol.* 2003; 105(1-2):1-9. [https://doi.org/10.1016/S0168-1656\(03\)00140-8](https://doi.org/10.1016/S0168-1656(03)00140-8)
 23. De Silva NMG, Borges MC, Hingorani AD, Engmann J, Shah T, Zhang X, *et al.* Liver function and risk of type 2 diabetes: Bidirectional mendelian randomization study. *Diabetes.* 2019; 68(8):1681-91. <https://doi.org/10.2337/db18-1048>
 24. Cheng L-Z, Li W, Chen Y-X, Lin Y-J, Miao Y. Autophagy and diabetic encephalopathy: Mechanistic insights and potential therapeutic implications. *Aging Dis.* 2022; 13(2):447-57. <https://doi.org/10.14336/AD.2021.0823>
 25. Othman MZ, Hassan Z, Che Has AT. Morris water maze: A versatile and pertinent tool for assessing spatial learning and memory. *Exp Anim.* 2022; 71(3):264-80. <https://doi.org/10.1538/expanim.21-0120>
 26. Gao M, Kang Y, Zhang L, Li H, Qu C, Luan X, *et al.* Troxerutin attenuates cognitive decline in the hippocampus of male diabetic rats by inhibiting NADPH oxidase and activating the Nrf2/ARE signaling pathway. *Int J Mol Med.* 2020; 46(3):1239-48. <https://doi.org/10.3892/ijmm.2020.4653>
 27. Liu X, Liu M, Mo Y, Peng H, Gong J, Li Z, *et al.* Naringin ameliorates cognitive deficits in streptozotocin-induced diabetic rats. *Iran J Basic Med Sci.* 2016; 19(4):417-22.
 28. Majmudar F, Bhadania M, Joshi H. Menthol causes reduction of Scopolamine induced Glutamatergic neurotoxicity and cognitive deficits. *Asian J Pharm Pharmacol.* 2018; 4(3):256-64. <https://doi.org/10.31024/ajpp.2018.4.3.3>
 29. Shali KS, Soumya NPP, Mondal S, Mini S. Hepatoprotective effect of morin via regulating the oxidative stress and carbohydrate metabolism in STZ induced diabetic rats. *Bioact Compd Heal Dis.* 2022; 5:53. <https://doi.org/10.31989/bchd.v5i3.893>
 30. Hampel H, Mesulam M-M, Cuello AC, Farlow MR, Giacobini E, Grossberg GT, *et al.* The cholinergic system in the pathophysiology and treatment of Alzheimer's disease. *Brain.* 2018; 141(7):1917-33. <https://doi.org/10.1093/brain/awy132>
 31. Rodrigo R, Cauli O, Gomez-Pinedo U, *et al.* Hyperammonemia induces neuroinflammation that contributes to cognitive impairment in rats with hepatic encephalopathy. *Gastroenterology.* 2010; 139(2):675-84. <https://doi.org/10.1053/j.gastro.2010.03.040>
 32. Mao X-Y, Cao D-F, Li X, Yin J-Y, Wang Z-B, Zhang Y, *et al.* Huperzine A ameliorates cognitive deficits in streptozotocin-induced diabetic rats. *Int J Mol Sci.* 2014; 15(5):7667-83. <https://doi.org/10.3390/ijms15057667>
 33. Sandireddy R, Yerra VG, Areti A, Komirishetty P, Kumar A. Neuroinflammation and oxidative stress in diabetic neuropathy: Futuristic strategies based on these targets. *Int J Endocrinol.* 2014; 2014. <https://doi.org/10.1155/2014/674987>
 34. Morán MA, Mufson EJ, Gómez-Ramos P. Colocalization of cholinesterases with beta amyloid protein in aged and Alzheimer's brains. *Acta Neuropathol.* 1993; 85(4):362-9. <https://doi.org/10.1007/BF00334445>
 35. Teleanu RI, Niculescu AG, Roza E, Vladăcenco O, Grumezescu AM, Teleanu DM. Neurotransmitters-key factors in neurological and neurodegenerative disorders of the central nervous system. *Int J Mol Sci.* 2022; 23(11). <https://doi.org/10.3390/ijms23115954>
 36. Swamy BK, Shiprath K, Rakesh G, Ratnam KV, Manjunatha H, Janardan S, *et al.* Simultaneous detection of dopamine, tyrosine and ascorbic acid using NiO/graphene modified graphite electrode. *Biointerface Res Appl Chem.* 2020; 10(3):5599-609. <https://doi.org/10.33263/BRIAC103.599609>

37. Strandwitz P. Neurotransmitter modulation by the gut microbiota. *Brain Res.* 2018; 1693:128-33. <https://doi.org/10.1016/j.brainres.2018.03.015>
38. Ou Y, Buchanan AM, Witt CE, Hashemi P. Frontiers in electrochemical sensors for neurotransmitter detection: Towards measuring neurotransmitters as chemical diagnostics for brain disorders. *Anal Methods.* 2019; 11(21):2738-55. <https://doi.org/10.1039/C9AY00055K>
39. Al-Sabri MH, Nikpour M, Clemensson LE, Attwood MM, Williams MJ, Rask-Anderson M, *et al.* The regulatory role of AP-2 β in monoaminergic neurotransmitter systems: Insights on its signalling pathway, linked disorders and theragnostic potential. *Cell Biosci.* 2022; 12(1):151. <https://doi.org/10.1186/s13578-022-00891-7>
40. Ezzeldin E, Souror WAH, El-Nahhas T, Soudi ANMM, Shahat AA. Biochemical and neurotransmitters changes associated with tramadol in streptozotocin-induced diabetes in rats. *Biomed Res Int.* 2014; 2014. <https://doi.org/10.1155/2014/238780>
41. Wang W, Jiang Y, Cai E, Li B, Zhao Y, Zhu H, *et al.* L-menthol exhibits antidepressive-like effects mediated by the modification of 5-HTergic, GABAergic and DAergic systems. *Cogn Neurodyn.* 2019; 13(2):191-200. <https://doi.org/10.1007/s11571-018-9513-1>
42. Duncan J, Johnson S, Ou X-M. Monoamine oxidases in major depressive disorder and alcoholism. *Drug Discov Ther.* 2012; 6(3):112-22. <https://doi.org/10.5582/ddt.2012.v6.3.112>
43. Hureau C, Sasaki I, Gras E, Faller P. Two functions, one molecule: A metal-binding and a targeting moiety to combat Alzheimer's disease. *Chembiochem.* 2010; 11(7):950-3. <https://doi.org/10.1002/cbic.201000102>
44. Alper G, Girgin FK, Ozgönül M, Menteş G, Ersöz B. MAO inhibitors and oxidant stress in aging brain tissue. *Eur Neuropsychopharmacol.* 1999; 9(3):247-52. [https://doi.org/10.1016/S0924-977X\(98\)00035-2](https://doi.org/10.1016/S0924-977X(98)00035-2)
45. Marchi S, Giorgi C, Suski JM, Agnoletto C, Bononi A, Bonora M, *et al.* Mitochondria-ros crosstalk in the control of cell death and aging. *J Signal Transduct.* 2012; 2012. <https://doi.org/10.1155/2012/329635>
46. Adefegha SA, Dada FA, Oyeleye SI, Oboh G. Effects of berberine on cholinesterases and monoamine oxidase activities, and antioxidant status in the brain of Streptozotocin (STZ)-induced diabetic rats. *J Basic Clin Physiol Pharmacol.* 2021; 33(4):389-97. <https://doi.org/10.1515/jbcpp-2020-0173>
47. Emory H, Mizrahi N. Monoamine oxidase inhibition in a patient with type 1 diabetes and depression. *J Diabetes Sci Technol.* 2016; 10:1203-4. <https://doi.org/10.1177/1932296816638106>