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# Metal-Infused Polyphenol-enriched Phyto-fabricated Nanoparticles: an In-depth Review of their Potent Prebiotic Properties

G. Thirumala Reddy<sup>1</sup>, Sri Lakshmi Aluri<sup>2</sup> and A. R. Shashikala<sup>3</sup>\*

#### **Abstract**

In light of its potential health benefits, including as improved gut health and management of the gut microbiota, herbal extracts have witnessed a major increase in demand as prebiotics in recent years. Many studies have demonstrated that polyphenol-rich herbal extracts can work as prebiotics by encouraging the growth of beneficial gut flora. Unfortunately, the limited solubility and stability of these herbal extracts, as well as their susceptibility to breakdown in the gastrointestinal system, can limited their bioavailability and efficiency. To address these limitations, the integration of metal nanoparticles has emerged as a promising strategy for the efficient delivery of herbal extracts, affording heightened bioavailability and precise targeting of the gut microbiota. In this paper, we present current breakthrough in metal infused nanoparticle-based assessment of herbal extracts as efficient prebiotics, with an emphasis on formulation, characterization, and biological activity. The integration of metal nanoparticles into polyphenol-rich herbal extracts represents a cutting-edge approach to enhancing the prebiotic properties of these compounds. Metal-infused polyphenol-enriched phyto-fabricated nanoparticles hold great promise for improving gut health and modulating the gut microbiota, offering innovative solutions to address the limitations associated with traditional polyphenol delivery. Furthermore, extracts of polyphenol-rich medicinal herbs such as rosehip flower, hibiscus flower, mango bark, bamboo stem, green tea, pomegranate, and dhataki flower have been investigated to assess the viability of employing these phyto generated metal infused nanoparticles as a potential prebiotic.

Keywords: Bioavailability, Herbal Extract, Polyphenol, Prebiotics, Probiotics, Nanoparticles

#### 1.0 Introduction

The gut microbiota is essential for human health and has been related to a variety of disorders including inflammatory bowel disease, obesity, and diabetes<sup>1</sup>. Prebiotics are no digestible dietary substances that specifically stimulate the formation and activity of healthy gut bacteria<sup>2,3</sup>. Due to their capacity to alter the gut flora, herbal extracts, which include a range of bioactive compounds, have been studied as possible prebiotics<sup>4</sup>.

However, the limited solubility and stability of these herbal extracts, together with their susceptibility to digestive system breakdown, might limit their bioavailability and efficiency<sup>5</sup>. Nanoparticles have been suggested as an efficient method of delivering herbal extracts to overcome these difficulties, enabling increased bioavailability and tailored administration to the gut flora<sup>6,7</sup>.

This comprehensive review will provide a broad overview of how Phyto-fabricated nanoparticles made from herbal extracts high in polyphenols

<sup>&</sup>lt;sup>1</sup>Department of Chemistry, Presidency University, Bangalore - 560064, Karnataka, India

<sup>&</sup>lt;sup>2</sup>Manager R and D, Prakruti Products Private Limited, Bangalore - 560079, Karnataka, India

<sup>&</sup>lt;sup>3</sup>Department of Chemistry, School of Engineering, Presidency University, Bangalore - 560064, Karnataka, India; aarudirs@gmail.com

<sup>\*</sup>Author for correspondence

may be used as possible prebiotics to support gut health.

# 2.0 Gut Micro Biota and Its **Importance**

The human intestinal system has a complicated and diverse microbial ecology that is crucial to human health. According to estimations from Ley et al. and Qin et al., our gut contains 100 times more genes and up to 1000 distinct bacterial species than the human genome<sup>8,9</sup>. Because of its considerable impact on human welfare, including host metabolism, physiology, nutrition, and immunological function, this community is sometimes referred to as our hidden metabolic organ<sup>10</sup>. It is well recognized that the intestinal Micro biota of healthy people offers a variety of health advantages such as pathogen defence, nourishment, homeostasis, and immunological regulation<sup>11</sup>.

According to conventional wisdom, infants' intestines are either sterile or contain only a very small number of bacteria after birth12. But, after delivery, the infant's gastrointestinal tract quickly becomes colonized. Many factors, such as the mode of birth, the kind of feeding, or the use of antibiotics, prebiotics, or probiotics, can have a significant impact on the composition of the infant's gut<sup>13</sup>. There is emerging evidence that the gut microbiota has a significant impact on gut health and sickness. Modulating the gut microbiota as a therapeutic strategy to treat chronic illness should thus be researched. Prebiotics and probiotic supplements can modify gut flora to improve host health<sup>10</sup>.

# 3.0 Prebiotics and their **Advantages**

Prebiotics are dietary ingredients that, in addition to boosting the immune system, promote the proliferation of probiotic bacteria in the human stomach<sup>14</sup>. The relationship between prebiotics and human health has drawn more attention in recent years. Prebiotics are a type of nutrition that the gut bacteria digest and use to nourish the intestinal flora. When they are broken down, they release short-chain fatty acids into the bloodstream, affecting not just the digestive system but also other organs<sup>15</sup>.

Prebiotics are non-digestible food components that benefit the host by stimulating one or more types of bacteria in the colon to grow and operate in order to promote better human health<sup>16</sup>. According to a study conducted by Tzounis et al., flavanols have been demonstrated to encourage the formation of lactic acid bacteria in both in vitro and in vivo screening<sup>17</sup>. Prebiotics give gut bacteria energy, enabling them to alter their makeup and activity. The ability of different bacterial species to ferment various prebiotics varies, and the chain length of the prebiotic also plays a role. Although certain by-products of prebiotic fermentation may be hostile, others can act as substrates for other bacteria. Prebiotics can also impact the gut environment by lowering pH levels, which can encourage Firmicutes to produce butyrate and alter the Micro biota in the gut<sup>15</sup>.

## 4.0 Herbal Extracts as a Prebiotic

Herbal extracts have been studied as potential prebiotics, or chemicals that promote the growth and activity of beneficial bacteria in the stomach. Prebiotics are a type of dietary fibre that is resistant to digestion and is preferentially digested by the gut Microbiota, creating Short-chain Fatty Acids (SCFAs) that nourish the cells lining the colon and have other health benefits.

Polyphenols have aromatic rings with one or more hydroxyl groups in their chemical structure, which can range from a simple phenolic molecule to a complicated high-molecular mass polymer. Polyphenols are secondary plant metabolites<sup>18</sup>. These chemicals are more likely to interact with intestinal bacteria due to their extensive metabolism in the large intestine and restricted bioavailability. In reality, there is a bidirectional interaction between the gut microbiota and polyphenols, with bacteria influencing phenolic compound activity. By regulating their metabolism and absorption, this interaction can change polyphenols into metabolites that may have a variety of physiological impacts on the host<sup>19</sup>.

## 5.0 Herbal Extracts High in Polyphenols as a Possible **Prebiotic**

Polyphenols are naturally occurring chemicals found in many fruits, vegetables, and herbs. They've been connected to a variety of health benefits, including antiinflammatory, antioxidant, and prebiotic properties.

Several studies have found that polyphenol-rich herbal extracts including green tea, grape seed, and olive leaf can work as prebiotics by promoting the growth of beneficial gut bacteria like Bifidobacterium and Lactobacillus<sup>20</sup>.

According to Alves-Santos et al., applying a concentration of polyphenol enriched extracts from medicinal herbs can promote probiotic development while inhibiting pathogen antimicrobial action<sup>19</sup>. The precise mechanism by which polyphenols increase SCFA production is unknown. It is thought that an increase in anaerobic microbes, particularly butyrate-producing ones like Lactobacillus, Lachnospiraceae, and Ruminococcaceae, might encourage a spike in SCFA21. Another notion is that the polyphenols included in decaffeinated green and black tea decrease the activity of the enzymes-amylase and -glucosidase in saliva and the small intestine. As a result, certain carbs may linger in the large intestine and serve as a substrate for SCFA synthesis<sup>22</sup>. SCFA are the primary mediators linking disease, metabolism, intestinal microbiota, and dietary support<sup>23</sup>. Butyrate study has mostly focused on the existence of butyrate-producing bacteria, as well as butyrate itself, which is thought to have favourable effects on human health. This implies that it could be used as an indicator of prebiotic influence<sup>24</sup>.

Because of their antioxidant, anti-inflammatory, and prebiotic qualities, polyphenol-rich medicinal herbal extracts such as rosehip flower, hibiscus flower, mango bark, bamboo stem, green tea leaves, pomegranate rind, and dhataki flower have been found to provide a variety of health advantages. Recently, researchers have been exploring the possibility of using polyphenol enriched herbal extracts as potent prebiotics.

The possible prebiotic effects of rose hip extract on the gut microbiota have been investigated. The fruit of the rose plant, or rose hips, is a good source of vitamin C, polyphenols, and other bioactive substances that may be beneficial for your health<sup>25</sup>. Rose hip extract may promote the growth of beneficial bacteria like Bifidobacteria and Lactobacilli while inhibiting the growth of harmful bacteria like E. coli and Clostridium perfringens, according to numerous in vitro and animal studies. Potential prebiotic effects on the gut microbiota of rose hip extract have been investigated. Rose hips, which are the fruit of the rose plant, are a great source of vitamin C, polyphenols, and other bioactive substances that may be beneficial for your health<sup>25</sup>. According to a number of in vitro and animal studies, rose hip extract may promote the growth of beneficial bacteria like Bifidobacteria and Lactobacilli while inhibiting the growth of harmful bacteria like *E. coli* and *Clostridium perfringens*.

Hibiscus flowers are high in bioactive substances like phenolic acids, flavonoids, anthocyanins, and polysaccharides. Several components have been investigated for possible health advantages, including prebiotic effects<sup>26</sup>. Hibiscus polysaccharides, which are found in the blooms of the plant, have been proven to have prebiotic characteristics by favourably promoting the growth of beneficial bacteria like Lactobacillus and Bifidobacterium. In addition to their prebiotic effects, hibiscus flowers also have anti-inflammatory, antioxidant, and hypoglycaemic effects<sup>27</sup>. These properties make hibiscus flowers a potential functional food ingredient for promoting gut health and overall wellbeing.

On the prebiotic benefits of mango bark, there is little study. The potential health advantages of numerous bioactive compounds found in mango bark, including tannins, flavonoids, and phenolic acids, have been researched28. While some research has suggested that certain components in mango bark, such as tannins, may have prebiotic effects, more research is needed to confirm these findings.

Bamboo stem may be a viable source of prebiotic fibre, according to the minimal study that has been done on its prebiotic benefits<sup>29</sup>. Bamboo stems have polysaccharides that cannot be broken down by human enzymes and, as a result, can specifically promote the development and activity of good bacteria in the gut Bamboo stem extract, according to Azmi et al., can specifically encourage the growth of Bifidobacterium and Lactobacillus, which are known to have probiotic qualities<sup>30</sup>. Additionally, research on bamboo stem extract has revealed that it possesses anti-inflammatory and antioxidant qualities, which may aid in enhancing intestinal health and general wellbeing. To improve intestinal health, adding bamboo stems or bamboo shoot extract to your diet may be a helpful

Green tea leaves have been examined for their possible prebiotic effects on the gut flora. The catechins in green tea, especially epigallocatechin gallate (EGCG), have been proven to have prebiotic characteristics by specifically promoting the growth of beneficial bacteria in the stomach. Alves et al., claim that the probiotic bacteria Lactobacillus and Bifidobacterium can grow more readily when EGCG is present<sup>19</sup>. Green tea extract has also been seen to lessen the amount of potentially harmful bacteria in the gut, including Clostridium and Fusobacterium. Green tea leaves also contain a lot of fibre, which can encourage the development and activity of good bacteria in the stomach. Short-Chain Fatty Acids (SCFAs), which have been linked to a variety of health benefits, can be produced by gut bacteria using the fibre in green tea leaves as a substrate<sup>31</sup>. The polyphenols in decaffeinated green and black tea, which have been demonstrated to lessen the activity of the enzymes-amylase and -glucosidase in saliva and the small intestine, could be the basis of an alternative idea. Because of this, some carbs may persist in the large intestine and serve as a substrate for SCFA synthesis22.

Punicalagin, a polyphenol found in high concentrations in the rind of pomegranates, has been studied for its potential prebiotic effects on gut flora. Prebiotics are indigestible food ingredients that specifically promote the growth and activity of beneficial bacteria in the digestive tract. Punicalagin from pomegranate rind, according to Li et al., can limit the growth of harmful bacteria like Escherichia coli and Clostridium perfringens while promoting the development of good bacteria in the gut like Bifidobacterium and Lactobacillus<sup>32</sup>. Punicalagin has also been discovered to boost the production of Short-Chain Fatty Acids (SCFAs) in the gut, which have been associated to a number of health advantages, such as improved glucose management, reduced inflammation, and improved gut barrier function. Pomegranate peel has been reported to have prebiotic effects in addition to antiinflammatory, antioxidant, and antibacterial qualities, which may contribute to its potential health benefits.<sup>33</sup>

Ayurveda has long prized the Dhataki flower, also known as Woodfordia fruticosa, as a medicinal plant because of the variety of health benefits it offers. The dried blooms of the Dhataki tree have been used as an astringent to treat ulcers, wounds, and diarrhoea in southern Asia<sup>34,35</sup>. Dhataki flower may affect the gut microbiome prebiotically, according to recent studies. Flavonoids, phenolic acids, and tannins are just a few of the bioactive substances found in dhataki flowers, and studies have shown that they have antioxidant, antiinflammatory, and antibacterial activities<sup>36-38</sup>. In addition, it has been discovered that certain of these substances, particularly the tannins, have prebiotic effects on the

gut microbiome by specifically promoting the growth of good bacteria like Bifidobacterium and Lactobacillus<sup>39,40</sup>. According to a study by Das et al., a Dhataki flower extract greatly enhanced the development of Bifidobacterium and Lactobacillus in vitro while inhibiting the growth of harmful bacteria like Escherichia coli and Staphylococcus aureus<sup>34</sup>. According to the experts, the high concentration of tannins and other polyphenols in Dhataki flower is what causes its prebiotic benefits.

As a potential source of prebiotics, the current review largely focuses on botanical extracts rich in polyphenols. While these findings are promising, there is no conclusive evidence to support the prebiotic effects of aforementioned medicinal herbs. However, in detailed research is required to confirm their prebiotic effects and its potential benefits for human health.

## 6.0 Limitation of using Herbal Extracts as a Prebiotic

Prebiotics have been used with herbal extracts to promote the development of good gut flora. Despite numerous potential health advantages, there are a number of drawbacks to using plant extracts as prebiotics.

#### 6.1 Lack of Standardization

One of the major limitations of herbal extracts as prebiotics is the lack of standardization. Different extraction methods can yield different chemical compositions, leading to inconsistent results in terms of their prebiotic effects<sup>41,42</sup>.

#### **6.2 Variable Composition**

Herbal extracts may include a wide range of substances, some of which may be poisonous or hazardous to gut microbes. Additionally, different plant species, environmental circumstances, and processing techniques might affect the content of herbal extracts<sup>43,44</sup>.

#### 6.3 Limited Research

The prebiotic effects of certain plant extracts, notably in humans, have not gotten much attention. Despite some promising findings, more research is needed to determine the long-term efficacy and safety of using plant extracts as prebiotics<sup>45</sup>.

#### **6.4 Regulatory Issues**

The regulation of herbal extracts can be a challenge, particularly in terms of ensuring quality and safety. In many cases, herbal extracts are marketed as dietary supplements rather than drugs, which means they are subject to less rigorous testing and oversight<sup>46</sup>.

#### 6.5 Interactions with Medications

Herbal extracts can interact with certain medications, potentially causing harmful side effects. This can be particularly problematic for individuals who are taking multiple medications or who have underlying health conditions<sup>47</sup>.

## 6.6 Limited Drug Delivery

Herbal extracts have been used for centuries as traditional medicines. However, the use of herbal extracts in modern medicine has some limitations. One of the limitations is reduced drug delivery. The low solubility and permeability of herbal extracts, according to Kesarwani et al., contribute to their low bioavailability<sup>48</sup>. This means that when herbal extracts are administered orally, they may not be absorbed efficiently into the bloodstream, leading to reduced drug delivery.

To address this issue, researchers have created revolutionary medication delivery devices based on nanotechnology. In a review article, it is discussed how nanotechnology-based medication delivery systems might increase the bioavailability and effectiveness of herbal remedies. According to Bonifacio and co-worker's, the solubility and permeability of herbal extracts can be increased by these methods, which will result in enhanced medication delivery<sup>49</sup>.

Although there have been improvements, using herbal medicines still raises safety questions. Many studies talk about problems with toxicity that come up when using herbal medicines<sup>45,50</sup>. Additionally, pharmaceutical companies have reduced their focus on natural products due to several drawbacks such as inconsistent quality and quantity of active compounds in natural products<sup>51,52</sup>.

Although plant extracts have been used for centuries as traditional medicines, their application in modern medicine is constrained by problems including decreased drug delivery brought on by poor solubility and permeability. Researchers developed brand-new drug delivery systems based on nanotechnology to overcome this issue. However, there are still issues with safety that need to be resolved when using herbal medicines.

# 7.0 Nanoparticle-based Delivery **Systems**

Nanoparticle-based delivery techniques have been developed to boost the bioavailability and bioactivity of herbal extracts enriched with polyphenols. By increasing phytochemical bioactivity and bioavailability, these delivery systems work as drug carriers that can overcome some of the disadvantages that herbal medicines have<sup>53</sup>. Nanotechnology is a potential method for boosting the efficacy of natural products<sup>54</sup>.

Different types of nanoparticles, including polymeric nanoparticles (PNPs), nanocapsules, and nanospheres, have been used in drug delivery systems<sup>55</sup>. Prunus avium L. cherry extracts that are rich in polyphenols have been enclosed in PLGA (Poly Lactic-co-glycolic Acid), a biodegradable polymer<sup>56</sup>. Solid Lipid Nanoparticles (SLN) and Nanostructured Lipid Carriers (NLC) are other nanoparticle kinds that have been used in the creation of herbal medicines.53

A promising method for administering plant extracts that have been enhanced with polyphenols is nanoparticlebased delivery systems. These techniques can increase the bioavailability, site-specificity, and controlled release of nanoparticle medicines. To deliver natural substances like polyphenols, polymeric nanoparticles like nanospheres and nanocapsules are ideal drug delivery vehicles<sup>57</sup>.

Phyto-fabricated nanoparticles have several advantages as prebiotics over traditional prebiotic compounds<sup>58</sup>. They are biocompatible, biodegradable, and can be easily synthesized from natural sources without the need for complex chemical reactions<sup>59</sup>. In addition to their prebiotic properties, phyto-fabricated nanoparticles have potential applications in the fields of drug delivery, wound healing, and cancer therapy due to their biocompatibility and ability to target specific cells and tissues60,61.

It has been shown that nanoparticle-based delivery technologies improve the biological activity of herbal extracts as prebiotics<sup>62</sup>. For instance, by specifically encouraging the growth of advantageous gut bacteria, the encapsulating of ginger extract in chitosan nanoparticles improved its stability and increased its prebiotic effect<sup>63</sup>. Similar to this, resveratrol's solubility and stability

were increased through its encapsulation in solid lipid nanoparticles, enabling targeted administration to the colon and regulation of the gut microbiota<sup>64</sup>.

Nanoparticle-based delivery methods have emerged as a viable technique for increasing the bioavailability and efficacy of herbal extracts as prebiotics<sup>5</sup>. Nanoparticles can improve the stability and solubility of herbal extracts and enable targeted administration to the gut bacteria<sup>65</sup>. Further study is required to optimize the formulation and characterisation of nanoparticles for herbal extract administration, as well as to assess their safety and efficacy in vivo. Overall, the adoption of nanoparticlebased delivery methods has the potential to increase the potency of herbal extracts as prebiotics for improving gut health and avoiding illness.

Nanoparticles have been suggested as a successful delivery technique to get around the drawbacks of herbal extract as a prebiotic. By boosting their solubility and stability, nanoparticles can prevent the gastrointestinal tract from degrading herbal extracts and increase their bioavailability66. Additionally, nanoparticles can be created to target particular regions of the gastrointestinal system, enabling localised distribution of the herbal extracts to the gut Micro<sup>67,68</sup>.

## 8.0 Preparation of Nanoparticles or Encapsulation of Extracts with Prebiotic Potential

The preparation of nanoparticles or encapsulation of extracts with prebiotic potential involves several steps, including selection of appropriate materials, formulation optimization, and characterization of the final product<sup>69</sup>. Here is an elaboration on each of these steps:

#### 8.1 Selection of Materials

The choice of suitable materials is the first step in creating nanoparticles or encapsulating extracts. The material selection is based on the required characteristics of the finished product, including solubility, stability, and biocompatibility. Lipids, polymers, and metals are often employed materials in the creation of nanoparticles<sup>70,71</sup>. For instance, polymers like chitosan can be used to create polymeric nanoparticles, whereas lipids like phospholipids can be utilised to create liposomes<sup>72,73</sup>.

Additionally, metallic nanoparticles can be created using metals like gold<sup>74</sup>.

#### 8.2 Formulation Optimization

The next step is the optimization of the formulation, which involves determining the optimal ratio of materials and the appropriate conditions for nanoparticle formation. The formulation can be optimized using various methods, such as statistical experimental design, response surface methodology, or artificial neural75. Particle size, surface charge, drug loading, and drug release kinetics are among the formulation variables that are frequently optimised. Based on desirable product characteristics including bioavailability, stability, and targeted distribution, formulation parameters are chosen<sup>76</sup>.

#### 8.3 Characterization

The finished product is characterised once the formulation has been optimised to make sure it adheres to the required standards<sup>77</sup>. Measurements of the nanoparticles' or extracts' physical and chemical characteristics, such as their size, shape, surface charge, drug loading, and drug release kinetics, are done during characterization<sup>78</sup>. Dynamic light scattering, transmission electron microscopy, and Fourier-transform infrared spectroscopy are examples of typical characterisation methods. These methods can help guarantee that the nanoparticles or encapsulated extracts fulfil the required standards by providing information on the physical and chemical characteristics of each<sup>79,80</sup>. The prebiotic action and safety of the encapsulated extracts can also be assessed in vitro and in vivo<sup>81,82</sup>.

In conclusion, there are various phases involved in creating nanoparticles or encapsulating extracts with prebiotic potential, including choosing the right components, adjusting the formulation, and characterising the finished product. These procedures are essential for ensuring the manufacturing of prebioticpotential nanoparticles or encapsulated extracts that are secure, efficient, and high-quality83.

## 9.0 Conclusion

The potential of phyto-fabricated nanoparticles as a prebiotic has been addressed in this review. For

polyphenol-enriched herbal extracts, nanoparticle-based delivery systems have been investigated, and nanostrategies have significantly increased the utilisation of polyphenols. High-molecular-weight phenolic compounds bioavailability has been shown to be improved by encapsulation, and polyphenols' bioavailability-related drug delivery systems have also been extensively used. The review highlights how these nanoparticles have the potential to improve gut health by encouraging the growth of helpful bacteria and reducing the activity of harmful bacteria. Additionally, it has been discovered that phytofabricated nanoparticles contain anti-inflammatory and antioxidant capabilities that may help to further improve general health. In conclusion, phyto-fabricated nanoparticles have a promising future as prebiotics because they can increase the stability and bioavailability of herbal extracts high in polyphenols. However, more investigation is required to completely comprehend the mechanisms underlying these effects and to establish the ideal dosages for secure and efficient use. Overall, this review suggests that phyto-fabricated nanoparticles have great potential as a prebiotic supplement for improving gut health and overall well-being.

## 10.0 References

- 1. Guarner F, Malagelada JR. Gut flora in health and disease. The lancet. 2003 Feb 8; 361(9356):512-9. https://doi.org/10.1016/S0140-6736(03)12489-0 PMid:12583961
- 2. Collins S, Reid G. Distant site effects of ingested prebiotics. Nutrients. 2016 Aug 26; 8(9):523. https://doi.org/10.3390/ nu8090523 PMid:27571098 PMCid:PMC5037510
- 3. Louis, P., Flint, H. J., Michel, C. How to manipulate the Micro biota: prebiotics. Microbiota of the Human Body: Implications in Health and Disease. 2016; pp. 119-42. https://doi.org/10.1007/978-3-319-31248-4\_9 PMid:27161355
- 4. Anhê FF, Varin TV, Le Barz M, Desjardins Y, Levy E, Roy D, Marette A. Gut microbiota dysbiosis in obesitylinked metabolic diseases and prebiotic potential of polyphenol-rich extracts. Curr Obes Rep. 2015 Dec; 4:389-400. https://doi.org/10.1007/s13679-015-0172-9 PMid:26343880
- 5. Zhao Q, Luan X, Zheng M, Tian XH, Zhao J, Zhang WD, Ma BL. Synergistic mechanisms of constituents in herbal extracts during intestinal absorption:

- **Focus** natural occurring nanoparticles. on Pharmaceutics. 2020 Feb 3; 12(2):128. https://doi. org/10.3390/pharmaceutics12020128 PMid:32028739 PMCid:PMC7076514
- 6. Gunasekaran T, Haile T, Nigusse T, Dhanaraju MD. Nanotechnology: an effective tool for enhancing bioavailability and bioactivity of phytomedicine. Asian Pac J Trop Biomed. 2014 May 1; 4:S1-7. https://doi. org/10.12980/APJTB.4.2014C980 PMid:25183064 PMCid:PMC4025268
- 7. Chenthamara D, Subramaniam S, Ramakrishnan SG, Krishnaswamy S, Essa MM, Lin FH, Qoronfleh MW. Therapeutic efficacy of nanoparticles and routes of administration. Biomater Res. 2019 Dec; 23(1):1-29. https://doi.org/10.1186/s40824-019-0166-x PMid:31832232 PMCid:PMC6869321
- 8. Ley RE, Peterson DA, Gordon JI. Ecological and evolutionary forces shaping microbial diversity in the human intestine. Cell. 2006 Feb 24; 124(4):837-48. https:// doi.org/10.1016/j.cell.2006.02.017 PMid:16497592
- 9. Qin J, Li R, Raes J, Arumugam M, Burgdorf KS, Manichanh C, Nielsen T, Pons N, Levenez F, Yamada T, Mende DR. A human gut microbial gene catalogue established by metagenomic sequencing. Nature. 2010 Mar 4; 464(7285):59-65. https://doi.org/10.1038/nature08821 PMid:20203603 PMCid:PMC3779803
- 10. Guinane CM, Cotter PD. Role of the gut microbiota in health and chronic gastrointestinal disease: understanding a hidden metabolic organ. Therap Adv Gastroenterol. 2013 Jul; 6(4):295-308. https:// doi.org/10.1177/1756283X13482996 PMid:23814609 PMCid:PMC3667473
- 11. O'Hara AM, Shanahan F. The gut flora as a forgotten organ. EMBO Reports. 2006 Jul; 7(7):688-93. https:// PMid:16819463 doi.org/10.1038/sj.embor.7400731 PMCid:PMC1500832
- 12. Jiménez E, Marín ML, Martín R, Odriozola JM, Olivares M, Xaus J, Fernández L, Rodríguez JM. Is meconium from healthy new borns actually sterile? Res Microbiol. 2008 Apr 1; 159(3):187-93. https://doi.org/10.1016/j. resmic.2007.12.007 PMid:18281199
- 13. Fouhy F, Ross RP, Fitzgerald GF, Stanton C, Cotter PD. Composition of the early intestinal microbiota: knowledge, knowledge gaps and the use of highthroughput sequencing to address these gaps. Gut Microbes. 2012 May 1; 3(3):203-20. https:// doi.org/10.4161/gmic.20169 PMid:22572829 PMCid:PMC3427213

- 14. Ahlawat S, Asha, Sharma KK. Gut-organ axis: a microbial outreach and networking. Letters in applied microbiology. Blackwell Publishing Ltd. 2021 Jun 1; 72(6):636-68. https://doi.org/10.1111/lam.13333 PMid:32472555
- 15. Davani-Davari D, Negahdaripour M, Karimzadeh I, Seifan M, Mohkam M, Masoumi SJ, Berenjian A, Ghasemi Y. Prebiotics: definition, types, sources, mechanisms, and clinical applications. Foods. 2019 Mar 9; 8(3):92. https://doi.org/10.3390/foods8030092 PMid:30857316 PMCid:PMC6463098
- 16. Gibson GR, Roberfroid MB. Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. J Nutr. 1995 Jun 1; 125(6):1401-12. https:// doi.org/10.1093/jn/125.6.1401 PMid:7782892
- 17. Tzounis X, Rodriguez-Mateos A, Vulevic J, Gibson GR, Kwik-Uribe C, Spencer JP. Prebiotic evaluation of cocoa-derived flavanols in healthy humans by using a randomized, controlled, double-blind, crossover intervention study. Am J Clin Nutr. 2011 Jan 1; 93(1):62-72. https://doi.org/10.3945/ajcn.110.000075 PMid:21068351
- 18. Tsao R. Chemistry and biochemistry of dietary polyphenols. Nutrients. 2010 Dec 10; 2(12):1231-46. https://doi.org/10.3390/nu2121231 PMid:22254006 PMCid:PMC3257627
- 19. Alves-Santos AM, Sugizaki CS, Lima GC, Naves MM. Prebiotic effect of dietary polyphenols: A systematic review. J Funct Foods. 2020 Nov 1; 74: 104169. https:// doi.org/10.1016/j.jff.2020.104169
- 20. Moorthy M, Sundralingam U, Palanisamy UD. Polyphenols as prebiotics in the management of high-fat diet-induced obesity: A systematic review of animal studies. Foods. 2021 Feb 2; 10(2):299. https:// doi.org/10.3390/foods10020299 PMid:33540692 PMCid:PMC7913110
- 21. Li J, Wu T, Li N, Wang X, Chen G, Lyu X. Bilberry anthocyanin extract promotes intestinal barrier function and inhibits digestive enzyme activity by regulating the gut microbiota in aging rats. Food Funct. 2019; 10(1):333-43. https://doi.org/10.1039/C8FO01962B PMid:30575836
- 22. Henning SM, Yang J, Hsu M, Lee RP, Grojean EM, Ly A, Tseng CH, Heber D, Li Z. Decaffeinated green and black tea polyphenols decrease weight gain and alter microbiome populations and function in diet-induced obese mice. Eur. J. Nutr. 2018 Dec; 57(8):2759-69. https:// doi.org/10.1007/s00394-017-1542-8 PMid:28965248 PMCid:PMC7367598

- 23. Ríos-Covián D, Ruas-Madiedo P, Margolles A, Gueimonde M, De Los Reyes-gavilán CG, Salazar N. Intestinal short chain fatty acids and their link with diet and human health. Front. Microbiol. 2016 Feb 17; 7:185. https://doi.org/10.3389/fmicb.2016.00185 PMid:26925050 PMCid:PMC4756104
- 24. Koh A, De Vadder F, Kovatcheva-Datchary P, Bäckhed F. From dietary fiber to host physiology: short-chain fatty acids as key bacterial metabolites. Cell. 2016 2; 165(6):1332-45. https://doi.org/10.1016/j. cell.2016.05.041 PMid:27259147
- 25. Xu J, Jönsson T, Plaza M, Håkansson Å, Antonsson M, Ahrén IL, Turner C, Spégel P, Granfeldt Y. Probiotic fruit beverages with different polyphenol profiles attenuated early insulin response. J Nutr. 2018 Dec; 17:1-0. https:// doi.org/10.1186/s12937-018-0335-0 PMid:29486772 PMCid:PMC5827978
- 26. Diez-Echave P, Vezza T, Rodriguez-Nogales A, Ruiz-Malagón AJ, Hidalgo-Garcia L, Garrido-Mesa J, Molina-Tijeras JA, Romero M, Robles-Vera I, Pimentel-Moral S, Borras-Linares I. The prebiotic properties of Hibiscus sabdariffa extract contribute to the beneficial effects in diet-induced obesity in mice. Food Res Int. 2020 Jan 1; 127:108722. https://doi.org/10.1016/j. foodres.2019.108722 PMid:31882094
- 27. Sanadheera S, Subasinghe D, Solangaarachchi MN, Suraweera M, Suraweera NY, Tharangika N. Hibiscus rosa-sinensis L. (red Hibiscus) Tea, Can It Be Used as A Home-Remedy to Control Diabetes and Hypercholesterolemia?. Biol. Med. Natural Prod. Chem. 2021 Jul 27; 10(1):59-65. https://doi.org/10.14421/ biomedich.2021.101.59-65
- 28. Kim H, Castellon-Chicas MJ, Arbizu S, Talcott ST, Drury NL, Smith S, Mertens-Talcott SU. Mango (Mangifera indica L.) polyphenols: Anti-inflammatory intestinal microbial health benefits, and associated mechanisms of actions. Molecules. 2021 May 6; 26(9):2732. https:// doi.org/10.3390/molecules26092732 PMid:34066494 PMCid:PMC8124428
- 29. An Y, Lu W, Li W, Pan L, Lu M, Cesarino I, Li Z, Zeng W. Dietary fiber in plant cell walls-the healthy carbohydrates. Food Qual. Saf. 2022 Jan 1; 6:fyab037. https://doi.org/10.1093/fqsafe/fyab037
- 30. Azmi AF, Mustafa S, Hashim DM, Manap YA. Prebiotic activity of polysaccharides extracted from Gigantochloa levis (Buluh beting) shoots. Molecules. 2012 Feb 7; 17(2):1635-51. https://doi. org/10.3390/molecules17021635 PMid:22314383 PMCid:PMC6268289

- 31. Pérez-Burillo S, Navajas-Porras B, López-Maldonado A, Hinojosa-Nogueira D, Pastoriza S, Rufián-Henares JÁ. Green tea and its relation to human gut microbiome. Molecules. 2021 Jun 26; 26(13):3907. https://doi. org/10.3390/molecules26133907 PMid:34206736 PMCid:PMC8271705
- 32. Li Z, Summanen PH, Komoriya T, Henning SM, Lee RP, Carlson E, Heber D, Finegold SM. Pomegranate ellagitannins stimulate growth of gut bacteria in vitro: Implications for prebiotic and metabolic effects. Anaerobe. 2015 Aug 1; 34:164-8. https://doi. org/10.1016/j.anaerobe.2015.05.012 PMid:26051169
- 33. Howell AB, D'Souza DH. The pomegranate: effects on bacteria and viruses that influence human health. Evidence-Based Complementary Alternative Medicine. 2013 Oct; 2013:606212. https:// doi.org/10.1155/2013/606212 PMid:23762148 PMCid:PMC3671682
- 34. Das PK, Goswami S, Chinniah A, Panda N, Banerjee S, Sahu NP, Achari B. Woodfordia fruticosa: Traditional uses and recent findings. J Ethnopharmacology. 2007 Mar 21; 110(2):189-99. https://doi.org/10.1016/j. jep.2006.12.029 PMid:17276634
- 35. Sekar S, Vinothkanna A. Polyherbal and submerge fermented medicines of Ayurveda: Convergence of tradition with scientific trends and needs. S Afr J Bot. 2019 Mar 1; 121:410-7. https://doi.org/10.1016/j. sajb.2018.12.009
- 36. Tayab MA, Chowdhury KA, Jabed M, Mohammed Tareq S, Kamal AM, Islam MN, Uddin AK, Hossain MA, Emran TB, Simal-Gandara J. Antioxidant-rich Woodfordia fruticosa leaf extract alleviates depressivelike behaviors and impede hyperglycemia. Plants. 2021 Feb 3; 10(2):287. https://doi.org/10.3390/plants10020287 PMid:33546288 PMCid:PMC7913287
- 37. Thakur S, Kaurav H, Chaudhary G. A Review on Woodfordia fruticosa Kurz (Dhatki): Ayurvedic, Folk and Modern Uses. J. Drug Deliv Ther. 2021 May 15; 11(3):126-31. https://doi.org/10.22270/jddt.v11i3.4839
- 38. Giri S, Dey G, Sahu R, Paul P, Nandi G, Dua TK. Traditional Uses, Phytochemistry and Pharmacological Activities of Woodfordia fruticosa (L) Kurz: A Comprehensive Review. Indian J. Pharm. Sci. 2023 Mar 1; 85(1):1-12. https://doi.org/10.36468/pharmaceutical-sciences.1062
- 39. Rodríguez-Daza MC, Pulido-Mateos EC, Lupien-Meilleur J, Guyonnet D, Desjardins Y, Roy D. Polyphenol-mediated gut microbiota modulation: Toward prebiotics and further. Front Nutr. 2021 Jun

- 28; 8:689456. https://doi.org/10.3389/fnut.2021.689456 PMid:34268328 PMCid:PMC8276758
- 40. Sallam IE, Abdelwareth A, Attia H, Aziz RK, Homsi MN, von Bergen M, Farag MA. Effect of gut microbiota biotransformation on dietary tannins and human health implications. Microorganisms. 2021; 9:965. https://doi. org/10.3390/microorganisms9050965 PMid:33947064 PMCid:PMC8145700
- 41. Ong ES. Extraction methods and chemical standardization of botanicals and herbal preparations. J Chromatogr B. 2004 Dec 5; 812(1-2):23-33. https://doi.org/10.1016/ S1570-0232(04)00647-6
- 42. Ionescu MI. Are herbal products an alternative to antibiotics? In Bacterial Pathogenesis and Antibacterial Control 2017 Dec 20. IntechOpen. https://doi. org/10.5772/intechopen.72110
- 43. An X, Bao Q, Di S, Zhao Y, Zhao S, Zhang H, Lian F, Tong X. The interaction between the gut microbiota and herbal medicines. Biomed & Pharmacother. 2019 Oct 1; 118:109252. https://doi.org/10.1016/j. biopha.2019.109252 PMid:31545247
- 44. Oliphant K, Allen-Vercoe E. Macronutrient metabolism by the human gut microbiome: major fermentation by-products and their impact on host health. Microbiome. 2019 Dec; 7(1):1-5. https://doi.org/10.1186/s40168-019-0704-8 PMid:31196177 PMCid:PMC6567490
- 45. Ekor M. The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. Front Pharm. 2014 Jan 10; 4:177. https:// doi.org/10.3389/fphar.2013.00177 PMid:24454289 PMCid:PMC3887317
- 46. Thakkar S, Anklam E, Xu A, Ulberth F, Li J, Li B, Hugas M, Sarma N, Crerar S, Swift S, Hakamatsuka T. Regulatory landscape of dietary supplements and herbal medicines from a global perspective. Regul. Toxicol. Pharmacol. 2020 Jul 1; 114:104647. https://doi.org/10.1016/j. yrtph.2020.104647 PMid:32305367
- 47. Hussain S. Patient counseling about herbal-drug interactions. Afr J Tradit Complement Altern Med. 2011; 8(5S):152-163. https://doi.org/10.4314/ajtcam. v8i5S.8 PMid:22754069 PMCid:PMC3252717
- 48. Kesarwani K, Gupta R. Bioavailability enhancers of herbal origin: An overview. Asian Pac. J. Trop. Biomed. 2013 Apr 1; 3(4):253-66. https://doi.org/10.1016/S2221-1691(13)60060-X PMid:23620848
- 49. Bonifacio BV, da Silva PB, Ramos MA, Negri KM, Bauab TM, Chorilli M. Nanotechnology-based drug delivery systems and herbal medicines: a review. Int J

- Nanomedicine. 2014 Dec; 9:1-5. https://doi.org/10.2147/ IJN.S52634 PMid:24363556 PMCid:PMC3862741
- 50. Bateman J, Chapman RD, Simpson D. Possible toxicity of herbal remedies. Scott. Med. J. 1998 Feb; 43(1):7https://doi.org/10.1177/003693309804300104 15. PMid:9533252
- 51. Beutler JA. Natural products as a foundation for drug discovery. Curr Protoc Pharmacol. 2019 Sep; 86(1):e67. https://doi.org/10.1002/cpph.67 PMid:31539923 PMCid:PMC7442317
- 52. Atanasov AG, Zotchev SB, Dirsch VM, Supuran CT. Natural products in drug discovery: Advances and opportunities. Nat. Rev. Drug Discov. 2021 Mar; 20(3):200-16. https://doi.org/10.1038/s41573-020-00114-z PMid:33510482 PMCid:PMC7841765
- 53. Dewi MK, Chaerunisaa AY, Muhaimin M, Joni IM. Improved Activity of Herbal Medicines through Nanotechnology. Nanomaterials. 2022 Nov 18; 12(22):4073. https://doi.org/10.3390/nano12224073 PMid:36432358 PMCid:PMC9695685
- 54. Patra JK, Das G, Fraceto LF, Campos EV, Rodriguez-Torres MD, Acosta-Torres LS, Diaz-Torres LA, Grillo R, Swamy MK, Sharma S, Habtemariam S. Nano based drug delivery systems: recent developments and future prospects. J Nanobiotechnology. 2018 Dec; 16(1):1-33. https://doi.org/10.1186/s12951-018-0392-8 PMid:30231877 PMCid:PMC6145203
- 55. Moradi SZ, Momtaz S, Bayrami Z, Farzaei MH, Abdollahi M. Nanoformulations of herbal extracts in treatment neurodegenerative disorders. Front. Bioeng. Biotechnol. 2020 Apr 7; 8:238. https://doi.org/10.3389/ fbioe.2020.00238 PMid:32318551 PMCid:PMC7154137
- 56. Abdul Rahim R, Jayusman PA, Muhammad N, Ahmad F, Mokhtar N, Naina Mohamed I, Mohamed N, Shuid AN. Recent advances in nanoencapsulation systems using PLGA of bioactive phenolics for protection against chronic diseases. Int. J. Environ. Res. Public Health. 2019 Dec; 16(24):4962. https://doi.org/10.3390/ ijerph16244962 PMid:31817699 PMCid:PMC6950714
- 57. Salehi B, Machin L, Monzote L, Sharifi-Rad J, Ezzat SM, Salem MA, Merghany RM, El Mahdy NM, Kılıç CS, Sytar O, Sharifi-Rad M. Therapeutic potential of quercetin: New insights and perspectives for human health. Acs Omega. 2020 May 14; 5(20):11849-72. https:// doi.org/10.1021/acsomega.0c01818 PMid:32478277 PMCid:PMC7254783
- 58. Das P, Ghosh S, Nayak B. Phyto-fabricated nanoparticles and their anti-biofilm activity: Progress and current

- status. Front Nanotechnol. 2021 Oct 25; 3:739286. https://doi.org/10.3389/fnano.2021.739286
- 59. Durazzo A, Nazhand A, Lucarini M, Atanasov AG, Souto EB, Novellino E, Capasso R, Santini A. An updated overview on nanonutraceuticals: Focus on nanoprebiotics and nanoprobiotics. Int. J. Mol. Sci. 2020 Mar 26; 21(7):2285. https://doi.org/10.3390/ ijms21072285 PMid:32225036 PMCid:PMC7177810
- 60. Dutt Y, Pandey RP, Dutt M, Gupta A, Vibhuti A, Raj, VS, Priyadarshini A. Silver Nanoparticles Phytofabricated through Azadirachta indica: Anticancer, Apoptotic, and Wound-Healing Properties. Antibiotics. 2023. 12(1):121. https://doi.org/10.3390/antibiotics12010121 PMid:36671322 PMCid:PMC9855199
- 61. Baharlouei P, Rahman A. Chitin and chitosan: Prospective biomedical applications in drug delivery, cancer treatment, and wound healing. Mari Drugs. 2022 Jul 17; 20(7):460. https://doi.org/10.3390/md20070460 PMid:35877753 PMCid:PMC9319611
- 62. Ashaolu TJ. Emerging applications of nanotechnologies to probiotics and prebiotics. Int J Food Sci Techn. 2021 Aug; 56(8):3719-25. https://doi.org/10.1111/ijfs.15020
- 63. Farmoudeh A, Shokoohi A, Ebrahimnejad P. Preparation and evaluation of the antibacterial effect of chitosan nanoparticles containing ginger extract tailored by central composite design. Adv. Pharm. Bull. 2021 Sep; 11(4):643. https://doi.org/10.34172/apb.2021.073 PMid:34888211 PMCid:PMC8642796
- 64. Goktas Z, Zu Y, Abbasi M, Galyean S, Wu D, Fan Z, Wang S. Recent advances in nanoencapsulation of phytochemicals to combat obesity and its comorbidities. J. Agric. Food Chem. 2020 Jul 7; 68(31):8119-31. https:// doi.org/10.1021/acs.jafc.0c00131 PMid:32633507 PMCid:PMC8507418
- 65. Grgić J, Šelo G, Planinić M, Tišma M, Bucić-Kojić A. Role of the encapsulation in bioavailability of phenolic compounds. Antioxidants. 2020 Sep 26; 9(10):923. https://doi.org/10.3390/antiox9100923 PMid:32993196 PMCid:PMC7601682
- 66. Teja PK, Mithiya J, Kate AS, Bairwa K, Chauthe SK. Herbal nanomedicines: Recent advancements, challenges, opportunities and regulatory overview. Phytomedicine. 2022 Feb 1; 96:153890. https://doi.org/10.1016/j. phymed.2021.153890 PMid:35026510
- 67. Hu CM, Zhang L, Aryal S, Cheung C, Fang RH, Zhang L. Erythrocyte membrane-camouflaged polymeric nanoparticles as a biomimetic delivery platform. Proceedings of the National Academy of Sciences.

- 2011 Jul 5; 108(27):10980-5. https://doi.org/10.1073/ pnas.1106634108 PMid:21690347 PMCid:PMC3131364
- 68. Lu L, Chen G, Qiu Y, Li M, Liu D, Hu D, Gu X, Xiao Z. Nanoparticle-based oral delivery systems for colon targeting: principles and design strategies. Sci Bull. 2016 May 1; 61(9):670-81. https://doi.org/10.1007/s11434-016-1056-4
- 69. Razavi S, Janfaza S, Tasnim N, Gibson DL, Hoorfar M. Microencapsulating polymers for probiotics delivery systems: Preparation, characterization, and applications. Food Hydrocoll. 2021 Nov 1; 120:106882. https://doi. org/10.1016/j.foodhyd.2021.106882
- 70. Keawchaoon L, Yoksan R. Preparation, characterization and in vitro release study of carvacrol-loaded chitosan nanoparticles. Colloids and Surfaces B: Biointerfaces. 2011 May 1; 84(1):163-71. https://doi.org/10.1016/j. colsurfb.2010.12.031 PMid:21296562
- 71. Jaiswal M, Dudhe R, Sharma PK. Nanoemulsion: an advanced mode of drug delivery system. 3 Biotech. 2015 Apr; 5:123-7. https://doi.org/10.1007/s13205-014-0214-0 PMid:28324579 PMCid:PMC4362737
- 72. Pereira P, Carvalho V, Ramos R, Gama M. Chitosan nanoparticles for biomedical applications. Chitosan: manufacture, properties, and usage. Nova Science Publishers, Inc. New York, USA. 2011:321-64.
- 73. Allen TM, Cullis PR. Liposomal drug delivery systems: from concept to clinical applications. Adv. Drug Deliv. Rev. 2013 Jan 1; 65(1):36-48. https://doi.org/10.1016/j. addr.2012.09.037 PMid:23036225
- 74. Dreaden EC, Alkilany AM, Huang X, Murphy CJ, El-Sayed MA. The golden age: gold nanoparticles for biomedicine. Chem Soc Rev. 2012; 41(7):2740-79. https://doi.org/10.1039/C1CS15237H PMid:22109657 PMCid:PMC5876014
- 75. Pathak Y, Thassu D, editors. Drug delivery nanoparticles formulation and characterization. CRC Press; 2016 Apr 19. https://doi.org/10.3109/9781420078053

- 76. Moghimi SM, Hunter AC, Murray JC. Long-circulating and target-specific nanoparticles: theory to practice. Pharmacol Rev. 2001 Jun 1; 53(2):283-318.
- Torchilin VP. Multifunctional, stimuli-sensitive nanoparticulate systems for drug delivery. Nat. Rev. Drug Discov. 2014 Nov; 13(11):813-27. https://doi.org/10.1038/nrd4333 PMid:25287120 PMCid:PMC4489143
- 78. Tan Q, Liu W, Guo C, Zhai G. Preparation and evaluation of quercetin-loaded lecithin-chitosan nanoparticles for topical delivery. Int J Nanomedicine. 2011 Aug 10:1621-30. https://doi.org/10.2147/IJN.S22411 PMid:21904452 PMCid:PMC3160948
- 79. Chopra M, Jain R, Dewangan AK, Varkey S, Mazumder S. Design of curcumin loaded polymeric nanoparticlesoptimization, formulation and characterization. J. Nanosci. Nanotechnol. 2016 Sep 1; 16(9):9432-42. https://doi.org/10.1166/jnn.2016.12363
- 80. Jeong K, Kang CS, Kim Y, Lee YD, Kwon IC, Kim S. Development of highly efficient nanocarrier-mediated delivery approaches for cancer therapy. Cancer Lett. 2016 Apr 28; 374(1):31-43. https://doi.org/10.1016/j. canlet.2016.01.050 PMid:26854717
- 81. Muhamad II, Jusoh YM, Nawi NM, Aziz AA, Padzil AM, Lian HL. Advanced natural food colorant encapsulation methods: Anthocyanin plant pigment. In Natural and Artificial Flavoring Agents and Food Dyes. Academic Press. 2018 Jan 1; pp. 495-526.
- 82. Gao C, Lyu F, Yin Y. Encapsulated metal nanoparticles for catalysis. Chem Rev. 2020 Jun 25; 121(2):834-81. https:// doi.org/10.1021/acs.chemrev.0c00237 PMid:32585087
- 83. Dubey SK, Parab S, Achalla VP, Narwaria A, Sharma S, Jaswanth Gowda BH, Kesharwani P. Microparticulate and nanotechnology mediated drug delivery system for the delivery of herbal extracts. J Biomater Sci, Polym Ed. 2022 Aug 13; 33(12):1531-54. https://doi.org/10.1080/09 205063.2022.2065408 PMid:35404217