



## Safety of UV-selected *Helicoverpa armigera* nucleopolyhedrovirus to non-target beneficial organisms

S. JEYARANI, P. KARUPPUCHAMY, N. SATHIAH and S. MANIMEGALAI

Department of Agricultural Entomology, Centre for Plant Protection Studies,  
Tamil Nadu Agricultural University, Coimbatore – 641003, Tamil Nadu, India.

E-mail: jeyajawahar@sify.com

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**ABSTRACT:** The safety of the UV-selected Coimbatore isolate of *Helicoverpa armigera* nucleopolyhedrovirus (*HaNPV*-UVT-CBE1) was tested against the non-target organisms viz., *Trichogramma chilonis*, *Chrysoperla carnea*, honey bee species and *Bombyx mori* to find out whether the cyclical exposure of *HaNPV* to ultra-violet (UV) radiation could cause adverse effect on the above organisms. The results showed that the *HaNPV*-UVT-CBE1 had no adverse effect on the growth parameters, survival and production parameters of the organisms tested in comparison with the UV shielded *HaNPV* (*HaNPV*-UVS-CBE1). The cyclical exposure of *HaNPV* to UV radiation for the selection of UV tolerant strain did not have any deleterious effect on the safety of the virus to the non-target organisms.

**KEY WORDS:** *Apis* sp., *Bombyx mori*, *Chrysoperla carnea*, *Helicoverpa armigera*, nucleopolyhedrovirus, safety, *Trichogramma chilonis*

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### INTRODUCTION

American bollworm, *Helicoverpa armigera* (Hubner) is a serious pest of legumes, cotton and vegetables in the Indian subcontinent and South East Asia (King, 1994). Changing scenario in the management of *H. armigera* embodies the nuclear polyhedrovirus (*HaNPV*) as an important biocontrol agent. However, the use of baculoviruses as biological control agents is hampered by their susceptibility to inactivation by ultraviolet (UV) light. Hence artificial selection for UV tolerance was attempted in baculoviruses viz., *Heliothis zea* (Boddie), *Heliothis virescens* (Fabricius) NPV (Bullock *et al.*, 1970) and *Cydia pomonella* (L.) granulovirus (*CpGV*) (Brassel and Benz, 1979; Krieg *et al.*, 1980). Selection for UV tolerance was also

attempted in *H. armigera* NPV. The UV selected *HaNPV* strain was proved to have increased persistence with higher C/B ratio under field conditions on cotton (1: 3.25) and chickpea (1: 1.82) in comparison with unselected *HaNPV* (Jeyarani, 2004). Utilization of baculoviruses as biopesticides requires proof that they are safe to non-target species, though they are isolated from naturally infected diseased insects. Several nucleopolyhedroviruses (NPVs) have been tested against beneficial organisms and natural enemies of insect pests (Maheshbabu, 1990; Heinz *et al.*, 1995; Geetha, 1997; Parthasarathy, 2002). Though it was reported that both prokaryotic and eukaryotic organisms including baculoviruses have evolved pathways to repair UV induced damage (Petrik *et al.*, 2003), the effect of UV selection of NPVs on the

safety of non-target organisms was not reported so far. It is also important to verify the safety of any organisms that are subjected to genetic improvement. Hence, keeping this in view, the present investigations were made on the safety of UV-selected/tolerant *HaNPV* (*HaNPV-UVT-CBE1*) to non-target organisms.

## MATERIALS AND METHODS

### Insect cultures

The parasitoids and predators, used in this study were obtained from the Biocontrol laboratory, honeybees from an Apiary and silkworms from Department of Sericulture, Tamil Nadu Agricultural University (TNAU), Coimbatore. The insects, *viz.*, *Trichogramma chilonis*, *Chrysoperla carnea* and *Bombyx mori* were maintained at a constant temperature of  $25.0 \pm 2.0^\circ\text{C}$ , honey bee species were maintained in an artificial hive and were utilized for the experiments at appropriate stages.

### Virus inoculum

The Coimbatore isolate-I of *H. armigera* nucleopolyhedrovirus (*HaNPV-CBE1*) maintained at the Biocontrol laboratory, TNAU was used in the present study. The inoculum was propagated in the early fifth instar *H. armigera* larvae by diet surface treatment method (Sathiah, 2001) and the concentration was standardized using Neuber Haemocytometer (Weber, England) (Evans and Shapiro, 1997). The *HaNPV-CBE1* was selected for UV tolerance by seven cyclical exposure to UV-B at a cumulative energy level of 11340 KJ/m<sup>2</sup> and at an irradiance level of 300 watts/m<sup>2</sup> in a weathering system (Sun Test CPS<sup>+</sup>, Atlas GmbH Germany that generated UV-B from xenon arc lamp source). The *HaNPV-UVT-CBE1* strain selected for UV tolerance was compared with the UV shielded strain (*HaNPV-UVS-CBE1*, not selected for UV tolerance) for their safety to the non-target organisms.

### Safety studies

The safety of *HaNPV-UVT-CBE1* strain was tested against *T. chilonis*, *C. carnea*, *Apis mellifera*, *A. cerana indica*, *Trigona iridipennis* and *B. mori*.

The treatments for the safety tests included *HaNPV-UVT-CBE1* @  $2 \times 10^7$  POB ml<sup>-1</sup>, *HaNPV-UVS-CBE1* @  $2 \times 10^7$  POB ml<sup>-1</sup>, endosulfan 35EC @ 0.07% and control with water spray. Against silkworm, the insecticide treatment was not included as it is a known fact that they are highly sensitive to chemicals (Kariappa and Narasimhanna, 1981; Bhosale *et al.*, 1988).

### *T. chilonis*

Newly emerged *T. chilonis* adults were anaesthetized using etherized carbon dioxide for 15 seconds (Tiwari and Khan, 2004) and were released @ 50 per treatment on the treated egg cards in polybags. Parasitization was allowed for two days after which the egg cards were transferred to fresh polybags. Observations on per cent parasitism, parasitoid emergence, adult longevity and total life cycle were recorded. The parasitoids emerged in the respective treatments in the first generation were counted by anaesthetizing with CO<sub>2</sub> and utilized for second generation studies and the procedure repeated.

### *C. carnea*

*Chrysoperla carnea* first instar grubs emerging from previously treated *C. carnea* eggs were released @ 1: 100 (Grub: Eggs) per treatment to the UVT-CBE1, UVS-CBE I and endosulfan treated *H. armigera* eggs. Each treatment had 50 grubs in five replicates. The grubs were confined in test tubes covered with muslin cloth and secured tightly with a rubber band. The grubs were daily fed with treated one day old *H. armigera* eggs till pupation. After pupation, they were separated and transferred to plastic containers (20cm ht, and 8cm dia) for adult emergence. The adults were allowed in plastic jars and fed with a mixture of honey, protein hydrolysate, fructose, yeast and (1: 1: 1: 1: 1). Observations on hatching, larval duration, larval mortality, pupal period and adult longevity were recorded.

### Honey bees

The safety of *HaNPV-UVT-CBE1* and UVS-CBE I was tested with three honey bee species *viz.*,

*A. cerana indica*, *A. mellifera* and *T. iridipennis*. One day old worker bees were used @ 30 per treatment in five replicates. The bees were caged (30x30x30 cm) and were fed with 50 per cent sucrose solution containing respective viruses @  $2 \times 10^7$  POB ml<sup>-1</sup>. Similarly, endosulfan @ 0.07% was mixed in 50 per cent sucrose solution (50%) and fed for 24h. Afterwards, 50 per cent sucrose alone was provided till the bees died. In control, bees were fed with sucrose solution alone. The mortality of bees was observed daily until all the bees died. The bees were observed for the presence of viral particles in the tissues.

### B. mori

Third instar larvae of hybrid race, PM x NB<sub>4</sub> D<sub>2</sub> were fed with chopped mulberry leaves treated with *HaNPV* (UVT and UVS) @  $2 \times 10^7$  POB ml<sup>-1</sup> for 24h. Subsequently, fresh untreated leaf bits were provided at 12 h interval. Each treatment had 20 larvae replicated five times. A check without virus was maintained by feeding the larvae with leaves dipped in distilled water. Observations on the larval weight, mortality, cocoon weight, pupal period, shell weight and adult emergence were recorded.

### Statistical analysis

Data were analyzed for significance using analysis of variance (ANOVA) and the means were separated using Duncan multiple range test (Duncan, 1966).

## RESULTS AND DISCUSSION

### Safety to *T. chilonis*

Exposure of *T. chilonis* to UVT-*HaNPV* by egg treatment did not show any deleterious effect on parasitism and parasitoid emergence, however, the per cent parasitism, adult emergence was significantly lower in endosulfan treated eggs in the first generation (Table 1). Similar trend was also observed in the second generation. The parasitoid had a total life cycle of 9.2 and 9 days in UVS-CBE I treatment and 9 and 9.4 days in UVT-CBE I treatment and the adult longevity was 3.2 and 3 days in UVS-CBE I and was 3 and 3.2 days in UVT-CBE I both in the first and second generation, respectively which were on par with control. Safety of *HaNPV* to *T. chilonis* has been reported by Muthiah and Rabindra (1991) and Boomathi *et al.* (2005).

**Table 1. Safety of *HaNPV* - UVT - CBE1 to *T. chilonis* by egg treatment method**

a. Generation I

Treatment	Parasitization *(%)	Parasitoid emergence * (%)	Period (days) <sup>5</sup>	
			Adult longevity	Total life cycle
UVS - CBE1	81.20 ± 1.14 <sup>a</sup>	91.60 ± 0.93 <sup>a</sup>	3.20 ± 0.37	9.20 ± 0.20
UVT - CBE1	80.40 ± 1.21 <sup>a</sup>	91.60 ± 0.68 <sup>a</sup>	3.00 ± 0.32	9.00 ± 0.32
Endosulfan 35EC	77.60 ± 1.08 <sup>b</sup>	89.20 ± 0.86 <sup>b</sup>	2.60 ± 0.40	8.80 ± 0.37
Control	81.60 ± 1.28 <sup>a</sup>	93.20 ± 0.89 <sup>a</sup>	3.20 ± 0.20	9.40 ± 0.24

<sup>5</sup> Differences between the means are not significant (P = 0.05) by DMRT; \*in a column means followed by similar letters are not significantly different (P = 0.05) by DMRT

a. Generation II

Treatment	Parasitization *(%)	Parasitoid emergence * (%)	Period (days) <sup>5</sup>	
			Adult longevity	Total life cycle
UVS - CBE1	80.80 ± 0.58 <sup>a</sup>	90.80 ± 0.58 <sup>a</sup>	3.00 ± 0.45	9.00 ± 0.32
UVT - CBE1	80.40 ± 1.21 <sup>a</sup>	91.20 ± 0.71 <sup>a</sup>	3.20 ± 0.20	9.40 ± 0.24
Endosulfan 35EC	78.00 ± 0.71 <sup>b</sup>	88.40 ± 0.93 <sup>b</sup>	3.00 ± 0.32	9.00 ± 0.32
Control	82.00 ± 0.84 <sup>a</sup>	91.60 ± 0.68 <sup>a</sup>	3.20 ± 0.20	9.20 ± 0.20

<sup>5</sup> Differences between the means are not significant (P = 0.05) by DMRT; \*in a column means followed by similar letters are not significantly different (P = 0.05) by DMRT

**Table 2. Safety of *Ha*NPV - UVT – CBE I to *C. carnea***

Parameter	Treatment			
	UVS – CBE11	UVT – CBE11	Endosulfan	Control
Hatchability (%) <sup>s</sup>	92.80±0.86	93.20±0.66	91.20±0.55	94.00±0.32
Larval period (days) <sup>s</sup>	10.40±0.24	10.40±0.24	9.80±0.20	10.20±0.20
Pupation (%) *	81.20±0.68 <sup>a</sup>	82.00±0.71 <sup>a</sup>	79.20±0.58 <sup>b</sup>	81.20±0.58 <sup>a</sup>
Pupal period (days) <sup>s</sup>	10.40±0.24	10.20±0.20	10.80±0.20	10.20±0.20
Adult emergence (%) *	89.60±0.37 <sup>ab</sup>	90.40±0.68 <sup>a</sup>	88.40±0.40 <sup>b</sup>	90.40±0.51 <sup>a</sup>
Adult longevity (days) <sup>s</sup>	37.00±0.45	36.40±0.81	35.60±0.60	37.40±0.40
Total life cycle (days) *	59.80±0.37 <sup>a</sup>	60.00±0.55 <sup>a</sup>	58.20±0.20 <sup>b</sup>	59.80±0.37 <sup>a</sup>
Larval mortality (%) *	8.80±0.37 <sup>a</sup>	9.20±0.32 <sup>a</sup>	10.40±0.20 <sup>b</sup>	8.40±0.40 <sup>a</sup>

<sup>s</sup> Differences between the means are not significant (P = 0.05) by DMRT; \* in a column means followed by similar letters are not significantly different (P = 0.05) by DMRT

**Table 3. Safety of *Ha*NPV - UVT – CBE I to different species of honey bees**

Species	Treatment	Per cent Honey bee mortality (days after feeding)				Mean number of days survived *
		5 *	10	15	20	
<i>Apis cerana indica</i>	UVS – CBE1	43.33 <sup>a</sup>	66.67	96.67	100.00	11.00 ± 0.41 <sup>a</sup>
	UVT – CBE1	46.67 <sup>b</sup>	66.67	93.33	100.00	11.50 ± 0.29 <sup>a</sup>
	Endosulfan	100.00 <sup>c</sup>	-	-	-	0.75 ± 0.25 <sup>b</sup>
	Control	43.33 <sup>a</sup>	63.33	96.67	100.00	11.75 ± 0.48 <sup>a</sup>
<i>Apis mellifera</i>	UVS – CBE1	20.00 <sup>a</sup>	50.00	86.67	100.00	16.75 ± 0.48 <sup>a</sup>
	UVT – CBE1	23.33 <sup>b</sup>	46.67	83.33	96.67	16.25 ± 0.25 <sup>a</sup>
	Endosulfan	100.00 <sup>c</sup>	-	-	-	1.25 ± 0.25 <sup>b</sup>
	Control	20.00 <sup>a</sup>	46.67	86.67	96.67	17.25 ± 0.25 <sup>a</sup>
<i>Trigona iridipennis</i>	UVS – CBE1	53.33 <sup>a</sup>	93.33	100.00	-	8.50 ± 0.29 <sup>a</sup>
	UVT – CBE1	56.67 <sup>b</sup>	93.33	100.00	-	8.75 ± 0.25 <sup>a</sup>
	Endosulfan	100.00 <sup>c</sup>	-	-	-	0.50 ± 0.29 <sup>b</sup>
	Control	56.67 <sup>a</sup>	96.67	100.00	-	9.00 ± 0.41 <sup>a</sup>

\* In a column means followed by similar letters are not significantly different (P = 0.05) by DMRT

### Safety to *C. carnea*

*Chrysoperla carnea* exposed to *Ha*NPV-UVT-CBE1 isolate showed no adverse effects on per cent hatchability, larval period, per cent pupation, pupal period, adult emergence, adult longevity, total life cycle and larval mortality in comparison with UVS-CBE1 and control. Endosulfan was found to be toxic compared to NPV and control. Larval mortality of 8.8 and 9.2 per cent in UVS-CBE1 and UVT-CBE1 indicated that larval mortality in the virus treatment

was due to natural causes only and was on par with control (8.4%). Application of UVS and UVT-CBE1 had no adverse effect on *C. carnea* with reference to pupation (81.2 and 82%), adult emergence (89.6 and 90.4%) and total life cycle (59.8 and 60 days), respectively (Table 2). Safety of baculoviruses to *C. carnea* has been reported by many workers (Heinz *et al.*, 1995; Subramanian, 2003) and the present investigation also proved the safety of UV selected *Ha*NPV. This shows that

**Table 4. Safety of *HaNPV* - UVT – CBE I to *B. mori***

Parameter	Treatment*		
	UVS – CBE11	UVT – CBE11	Control
Larval weight (g) at instar III	0.414±0.005	0.412±0.005	0.427±0.013
Larval weight (g) at instar IV	1.502±0.025	1.492±0.031	1.553±0.026
Larval weight (g) at instar V	3.224±0.056	3.185±0.044	3.273±0.021
Larval mortality (%)	12.143±1.010	11.428±0.922	10.714±0.714
Pupation (%)	82.143±1.056	81.714±0.565	83.571±0.481
Pupal period (days)	10.571±0.297	10.857±0.261	10.714±0.184
Adult emergence (%)	90.000±0.488	90.143±0.459	91.286±0.421
Cocoon weight (g)	1.683±0.020	1.666±0.021	1.715±0.012
Shell weight (g)	0.361±0.007	0.357±0.010	0.358±0.008

the *HaNPV* on UV exposure had not undergone any modification that could cause interference with the entomophages activity. However exposure to endosulfan treated eggs caused significant variation in the biological parameters like egg hatchability, larval period, pupal period and adult longevity of *C. carnea*.

#### Safety to honey bees

Results indicated no observable difference in the behaviour of caged bees during this test. Longevity of *A. cerana indica* was 11 and 11.5 days in UVS-CBE1 and UVT-CBE1, respectively, and was on par with control (11.75 days), while it was 16.75, 16.25 and 17.25 days in *A. mellifera* and 8.5, 8.75 and 9 days in *T. iridipennis* for UVS-CBE1, UVT-CBE1 and control, respectively (Table 3). Microscopical examination of the tissue smears revealed no evidence of infection or other pathological manifestations in the bees, perhaps because of cage effect mortalities were also noted in control. However, honeybees fed with endosulfan (0.07%) through sucrose solution caused 100 per cent mortality within five days. The safety of viruses to honey bees was reported earlier by Dhaduti and Mathad (1980), Santharam *et al.* (1982) and Parthasarathy (2002).

#### Safety to *B. mori*

Results on the larval and cocoon parameters revealed that they were not affected by the virus treatments in comparison with control. UVT-CBE1 and UVS-CBE1 recorded the shell weight of 0.357g

and 0.361g, respectively which was on par with control (0.358g) (Table 4). Dhaduti and Mathad (1979) reported that *Mythimna separata* (Walker) NPV was innocuous to *B. mori*. No signs of abnormality in *B. mori* treated with *HaNPV* were reported earlier (Bijjur *et al.*, 1991; Muthiah and Rabindra, 1991). Safety of other baculoviruses were also established previously (Parthasarathy, 2002; Subramanian, 2003).

The findings revealed the relative safety of UV selected *HaNPV* to the beneficial organisms and natural enemies that are often encountered along with the *H. armigera* under field conditions.

## REFERENCES

- Bhosale, S. H., Yadwad, V. B. and Kallapur, V. L. 1988. Residual toxicity of Ekalux EC-25 on the biochemical constituents of the fat body of 5<sup>th</sup> instar, *Bombyx mori*. *Indian Journal of Sericulture*, **27**: 73-77.
- Bijjur, S., Kulkarni, K. A., Lingappa, S., 1991. Safety of *Heliothis armigera* (Hübner) NPV to some non-target beneficial organisms. *Indian Journal of Entomology*, **53**: 475-478.
- Boomathi, N., Sivasubramanian, P., Raguraman, S. and Premalatha, K. 2005. Safety of botanical and microbial pesticide mixtures to *Trichogramma chilonis* Ishii (Hymenoptera: Trichogrammatidae). *Journal of Biological Control*, **19**: 23-29.
- Brassel, J. and Benz, G. 1979. Selection of a strain of the granulosis virus of the codling moth with improved

- resistance against artificial ultraviolet radiation and sunlight. *Journal of Invertebrate Pathology*, **33**: 358–363.
- Bullock, H. R., Hollingsworth, J. P. and Harstack, Jr. A. W. 1970. Virulence of *Heliothis* nuclear polyhedrosis virus exposed to monochromatic ultraviolet irradiation. *Journal of Invertebrate Pathology*, **16**: 419–422.
- Dhaduti, S. G., Mathad S. B., 1979. Effect of NPV of the armyworm *Mythimna (Pseudaletia) separata* on the silkworm *Bombyx mori*. *Experientia*, **35**: 81.
- Dhaduti, S. G., Mathad, S. B., 1980. Effect of nuclear polyhedrosis virus of the armyworm *Mythimna separata* on colonies of *Apis cerana indica*. *Journal of Apicultural Research*, **19**: 77-78.
- Duncan, O. D. 1966. Path analysis: Sociological examples. *American Journal of Sociology*, **72**: 1-6.
- Evans, H. F. and Shapiro, M. 1997. Viruses. In: Lacey, L. (Ed.). *Manual of Techniques in Insect Pathology*, 17-54. Academic Press, London.
- Geetha, N. 1997. Studies on genetic variability and development of improved formulations of nuclear polyhedrosis virus of *Helicoverpa armigera* (Hübner). Ph. D thesis, Tamil Nadu Agricultural University, Coimbatore, India. 138 pp.
- Heinz, K. M., McCutchen, B. F., Hermann, R., Parrella, M. P., Hammock, B. D. 1995. Direct effects of recombinant nuclear polyhedrosis viruses on selected non-target organisms. *Journal of Economic Entomology*, **88**: 259-264.
- Jeyarani, 2004. Studies on the Photostability and Compatibility of the Nucleopolyhedrovirus of the Cotton Bollworm, *Helicoverpa armigera* (Hübner) with Granulovirus and *Pseudomonas fluorescens*. Ph. D thesis, Tamil Nadu Agricultural University, Coimbatore, India. 152 pp.
- Kariappa, B. K. and Narasimhanna, M. N. 1981. Efficacy of insecticides against wingless grasshopper *Neorthacris acuticeps nilgirensis* U. infesting mulberry and the effect of feeding BHC sprayed leaves on silk worm *Bombyx mori* L. *Indian Journal of Sericulture*, **20**: 31-34.
- King, A. B. S. 1994. *Heliothis/Helicoverpa* (Lepidoptera, Noctuidae). In: Matthews, G. A, Tunstall, J. P. (Eds.), *Insect Pests of Cotton*. 39-108. CAB International, Wallingford.
- Krieg, A., Groner, A., Huber, J. and Matter, M. 1980. Über die Wirkung von mittel- und langwellen ultravioletter strahlen (UV-B and UA-A) auf insektenpathogene Bakterien und deren Beeinflussung durch UV-Schutzstoffe. *Nachrichtenbl Deut pflanzenschutz dienst (Braunschweig)*, **32**: 100-105.
- Maheshbabu, S. 1990. Studies on the efficacy of *Heliothis armigera* (Hübner) nuclear polyhedrosis virus and its safety to certain non-target species. M.Sc (Ag.) thesis, Tamil Nadu Agric Univ, Coimbatore, India. 120 pp.
- Muthiah, C. and Rabindra, R. J. 1991. Control of gram pod borer (*Heliothis armigera*) on pigeonpea with controlled droplet application of NPV and effect of oral feeding of virus on mulberry silkworm (*Bombyx mori*). *Indian Journal of Agricultural Sciences*, **61**: 449-452.
- Parthasarathy, P. 2002. Evaluation of baculoviruses for the management of diamondback moth, *Plutella xylostella*. Ph. D thesis, Tamil Nadu Agricultural University, Coimbatore, India. 154 pp.
- Petrik, D. T., Iseli, A., Montelone, B. A., Van Etten, J. L. and Clem, R. J. 2003. Improving baculovirus resistance to UV inactivation: increased virulence resulting from expression of a DNA repair enzyme. *Journal of Invertebrate Pathology*, **82**: 50–56.
- Santharam, G., Azeez Basha, A., Balasubramanian, M., 1982. Lack of susceptibility of honey bees to three insect viruses. *Indian Bee Journal*, **44**: 35.
- Sathiah, N. 2001. Studies on improving the production and formulation of the nuclear polyhedrosis virus of the cotton bollworm, *Helicoverpa armigera* (Hübner), Ph. D thesis, Tamil Nadu Agricultural University, Coimbatore, India. 276 pp.
- Subramanian, S. 2003. Studies on the use of baculoviruses for the management of *Plutella xylostella* (Linn.) and *Spodoptera litura* (Fabr.) infesting cauliflower. Ph. D thesis, Tamil Nadu Agricultural University, Coimbatore, India. 152 pp.
- Tiwari, S. and Khan, M. A. 2004. Effect of endosulfan on per cent parasitisation by three species of *Trichogramma*. *Indian Journal of Entomology*, **66**: 135-137.

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