

THE THYROID GLAND AND OXIDATIVE STRESS; THE ROLE OF MELATONIN

KARBOWNIK M^{1,2} and LEWINSKI A^{1,2}

¹Department of Endocrinology and Isotope Therapy, Medical University of Lodz;² Polish Mother's Memorial Hospital - Research Institute, 281/289, Rzgowska St., 93-338 Lodz, Poland. E-mail : alewin@csk.am.lodz.pl

SUMMARY

Reactive oxygen species (ROS) and free radicals participate in physiological and pathological processes in the thyroid gland. For example, the role of hydrogen peroxide (H₂O₂) is crucial for thyroid hormone biosynthesis. Also other free radicals or reactive species, formed from iodine or tyrosine residues, are produced during thyroid hormone synthesis in physiological conditions. In turn, much evidence has been accumulated, showing that thyroid diseases, e.g., Graves' disease, non-toxic goitre formation or thyroid cancer, are accompanied by enhanced oxidative stress. The presence of some antioxidants has been found in the thyroid. Melatonin (N-acetyl-5-methoxytryptamine) - the main secretory product of the pineal gland is a well-known antioxidant and free radical scavenger, widely distributed in the organism. Mutual relationships between the pineal gland and the thyroid have for a long time been a subject of intensive research. The abundant to-date's evidence relates mostly to the inhibitory action of melatonin on the thyroid growth and function. Recently, experimental models have been developed, showing - among others - protective effects of melatonin against oxidative damage to lipids in the thyroid gland. Thus, free radicals and antioxidants, melatonin included, may participate in both physiological and pathological processes in the thyroid, what has already been partially documented.

Key words : Antioxidants; Free radicals; Melatonin; Oxidative stress; Thyroid gland.

INTRODUCTION

Reactive oxygen species (ROS) and free radicals are produced in living organisms, in certain amounts, under physiological conditions. An overproduction of ROS and free radicals causes oxidative stress and can lead to several diseases (1, 2). The most basic reaction of oxidative stress is Fenton reaction, in which iron [as ferrous ion (Fe²⁺)] participates: $Fe^{2+} + H_2O_2 \rightarrow Fe^{3+} + \cdot OH + OH^-$. Hydroxyl radical ($\cdot OH$), produced in this reaction, is the most toxic free radical.

Melatonin (N-acetyl-5-methoxytryptamine) is produced in the pineal gland, as well as in numerous other tissues and organs (3). The indoleamine possesses properties of a hormone but also demonstrates other numerous characteristics, allowing its action in cellular compartments of all tissues and organs (4). Melatonin is a well-known antioxidant and free radical scavenger, revealing preventive action against oxidative damage in different tissues (5-9).

Relationship between the pineal gland and the thyroid gland

In different animal species, a suppressive effect of melatonin on thyroid growth processes (10-14) and thyroid function (15-17) was shown. In contrast, thyroid hormones reveal mainly stimulatory effects on the pineal, as regards growth and secretory processes of this gland (18). Experimental evidence, concerning the relationship between the thyroid and the pineal gland, also in relation to oxidative stress (18-23) has recently been summarized.

Corresponding author : Prof. Andrzej Lewinski, M.D., Ph.D.,

The role of oxidative stress in the thyroid gland- experimental evidence

Hydrogen peroxide (H₂O₂)

H₂O₂ is produced in the thyroid gland (24, 25) by the NADPH oxidase system of the apical membrane of thyroid follicular cells (thyrocytes) (26, 27); divalent reduction of oxygen, without superoxide anion radical (O₂^{·-}) generation, is involved in this process (28).

H₂O₂ is an essential factor for thyroid hormone synthesis, acting as an electron acceptor at each step of this process, namely iodide oxidation and, next, its organification, and coupling reaction of iodotyrosines (23). This reactive species is crucial for thyroid peroxidase (TPO) activity. TPO is the key enzyme for thyroid hormone synthesis. It is a heme-dependent protein, thus it contains iron. On one hand, increased iron stores in the organism are associated with an increased risk of several diseases, cancer included (1, 29). On the other hand, however, iron is an essential element for metabolism in different tissues and organs, also in the thyroid. It has been shown in studies on animals and in human subjects that iron deficiency reduces TPO activity, enhances the consequences of iodine deficit, while iron supplementation improves the efficacy of iodine supplementation (30).

Beside the main three steps of thyroid hormone synthesis, H₂O₂ also participates in autocatalytic covalent heme binding to the apoprotein of TPO molecule, thereby stabilizing the activity of the enzyme (31). It has also been found that either ROS or free radicals decrease TPO activity in the thyroid gland, without influencing TPO mRNA; the authors have speculated that TPO inactivation occurs at the heme-linked histidine residue of the enzyme molecule; this residue plays a critical role for TPO activity due to the presence of heme-derived iron, which potentially constitutes a substrate for Fenton reaction (28). Thus, while H₂O₂ is necessary for thyroid hormone synthesis, this ROS when being in excess may inhibit TPO activity, with a subsequent inhibition of thyroid hormone formation (28).

There are presumptions that oxidative stress may be involved in the pathomechanism of hypothyroidism and goiter formation. Using dog thyroid cells, it has been found that thyrotropin (TSH) - the main hormone, stimulating secretory and growth processes in the thyroid - enhances H₂O₂ generation through the cyclic adenosine 3',5'-monophosphate (cAMP) cascade (25). Thus, in any conditions of increased blood TSH concentration, mainly in case of hypothyroidism and in chronic stimulation of the thyroid gland due to iodine deficiency, an increased production of H₂O₂ must take place with subsequently enhanced formation of free radicals (especially ·OH).

It is suggested that H₂O₂ participates in Wolff-Chaikoff 's effect, i.e. in a phenomenon which relies on the fact that iodides, when in excess, are able to inhibit iodine organification and subsequent thyroid hormone synthesis. Iodide was shown to strongly inhibit both protein iodination and H₂O₂ generation stimulated by TSH in dog thyroid slices (32). It is suggested that both apoptosis and necrosis occur in the thyroid gland via the mechanism involving H₂O₂, the former process resulting from lower, and the latter process from higher concentrations of this species under *in vitro* conditions (33). Because melatonin directly neutralizes H₂O₂ (34) and is the most effective scavenger of ·OH (4), the indoleamine might prevent pathological processes in the thyroid caused by an excessive amount of H₂O₂.

Reactive nitrogen species

Nitric oxide synthase (NOS) catalyzes the formation of free radical-nitric oxide (NO·). An expression of mRNA for the three isoforms of NOS - brain (type I), endothelial (type III), and inducible (type II) has been detected in the rat thyroid gland (35-37). Nitric oxide has been found to inhibit cell proliferation in cultured human thyrocytes (38). Some data suggest that NO· participates in the regulation of thyroid hormone synthesis. While NO· inhibits TSH-stimulated iodide uptake by-probably-stimulation of guanylyl cyclase (GC) activity and cyclic GMP (cGMP) production in calf thyroid (39),

the radical has been found to stimulate thyroid peroxidase activity in monolayer cultures of primary human thyrocytes (40).

It is known that melatonin directly neutralizes NO^\cdot (41) and peroxyxynitrite anion (ONOO) (42), the latter being a highly destructive product of the interaction between the $\text{O}_2^{\cdot-}$ and NO^\cdot . Additionally, melatonin has been demonstrated to inhibit the activity of NOS (5). There is also an indirect evidence that melatonin detoxifies $\text{O}_2^{\cdot-}$ (43).

Other free radicals

Numerous free radicals and reactive species are undoubtedly involved in the complex process of thyroid hormone synthesis. According to experimental evidence or suggestions, the following radicals or reactive species are of special importance: tyrosine free radical (Tyr $^\cdot$), diiodotyrosyl residue radical (DIT $^\cdot$), diiodotyrosyl residue radical in thyroglobulin (Tg-DIT $^\cdot$), iodine radical (I^\cdot), iodonium ion (I^+), hypoiodous acid intermediate [$\text{IO}^\cdot(\text{IOH})$], and ascorbate radical (Asc $^\cdot$) (44).

Antioxidative defence mechanisms in the thyroid gland

An antioxidative defence system in the thyroid gland comprises both antioxidative enzymes and free radical scavengers. The presence of the following antioxidative enzymes in the thyroid gland has been documented: superoxide dismutase (SOD) (45, 46), glutathione peroxidase (GSH-Px) (47), and catalase (CAT) (48). It should be stressed here that melatonin has been documented to stimulate the activity of SOD, GSH-Px, and CAT (5, 6). Moreover, the presence of glutathione (GSH), a well known intracellular antioxidant, alpha- and gamma-tocopherols, and coenzyme Q has been found in human thyroid (49), and of ascorbic acid in hog thyroid (50). Ascorbic acid becomes a free radical itself in the process of scavenging free radicals in the thyroid gland (50).

Peroxioredoxins (Prxs), antioxidative proteins involved in the regulation of cell differentiation and proliferation, have been found in human thyroid follicular cells and in FRTL-5 cells (51). Using FRTL-5 cells, it has been documented that Prxs are involved in the process of H_2O_2 elimination, when this oxygen species is produced in response to TSH, and that they protect thyroid cells from H_2O_2 -induced apoptosis (51).

A positive immunostaining with antibodies against melatonin has been found in C cells in the rat thyroid gland (3, 52). It has not been examined till now whether melatonin is produced in thyroid follicular cells. However, because melatonin is able to reach any tissue or organ within very short time (5, 6), it is highly probable that the indoleamine is also available for the thyroid gland, when it is required to reveal antioxidative effects.

Oxidative changes in the thyroid gland under pathological conditions

The results obtained in humans and in animal models suggest that oxidative damage in the thyroid is accompanied by increased activities of antioxidative enzymes or increased production of antioxidants, what probably represents the defence mechanism (46, 47, 53). In case of human thyroid tissue - non-toxic nodular goitre, carcinoma (follicular and papillary) and follicular adenoma - the highest level of lipid peroxidation products was found in carcinomas and it was still increased in adenomas, when compared to the level of lipid peroxidation products in control tissue and in nodular goitre; those changes were accompanied by increased activities of antioxidative enzymes, such as SOD, GSH-Px, and CAT, especially in carcinomas (47).

A much higher amount of Prx I, measured by immunoblotting, has been found in follicular thyroid adenomas and carcinomas than in histopathologically unchanged thyroid tissue; additionally,

a significantly higher amount of Prx I has been found in thyroids collected from patients with Graves' disease than in normal thyroid tissue (54).

The presence of immunochemical staining of SOD type II (inducible), and of mRNA for this enzyme was found in the human papillary thyroid carcinoma but not in normal human thyroid tissue (55). On the contrary, a decreased expression of mRNA encoding for CAT, and copper and zinc SOD were found in anaplastic thyroid carcinoma, when compared to histopathologically unchanged thyroid tissue and to differentiated thyroid tumors (56).

Thus, in case of more advanced tumors or, generally, more serious diseases of the thyroid gland, more free radicals are generated, followed by an increased production of antioxidants or increased activities of antioxidative enzymes. However, at extremely advanced stages of diseases, like anaplastic carcinoma, the defence mechanisms remain inactive.

Regarding the protective effects of melatonin against oxidative stress in the thyroid gland, experimental data have unfortunately been very scarce. However, we have found recently a protective action of melatonin against lipid peroxidation in porcine thyroid. Because iron is present in TPO, and H_2O_2 is essential for TPO activity, the thyroid gland may - under pathological conditions - be exposed to excessive amounts of either Fe^{2+} or H_2O_2 , or both. Ferrous iron and H_2O_2 constitute substrates for Fenton reaction.

Using an *in vitro* model of Fenton reaction, we have induced oxidative damage to lipids in homogenates of porcine thyroids; we have found that Fe^{2+} plus H_2O_2 increased - in the Fe^{2+} concentration - dependent manner - the level of lipid peroxidation products [malondialdehyde + 4-hydroxyalkenals (MDA+4-HDA)]; at the same time, melatonin - in a concentration-dependent manner - decreased lipid peroxidation induced by Fenton reaction, with the lowest effective concentration of 0.25 mM (57). We would like to indicate that the above study is the first one showing lipid peroxidation induction by Fenton reaction and the protective influence of melatonin against oxidative damage in the thyroid tissue.

Involvement of free radicals and reactive species in thyroid enlargement

Several experimental data, as well as results of studies in humans, support the hypothesis that oxidative stress is involved in goitre formation. It is supposed that $NO\cdot$ participates in vascular control during goitre formation. In animal model of thiouracil - and low iodine diet - induced goitre, the increased vascularization of the gland was accompanied by an induction of genes encoding for NOS I and NOS III; during goiter involution, the expression for these genes returned to basal values (37).

In another study, low iodine diet resulted as expected in an increased thyroid weight, increased DNA and protein content, and increased 3H -thymidine incorporation into DNA of thyroid follicular cells; all those changes were reduced by an administration of an antioxidant - vitamin E (58). It is worth stressing that the changes in parameters of growth processes after vitamin E treatment were not accompanied by any changes in either TSH or thyroid hormone concentrations (58). These results suggest that vitamin E has a direct antigoirogenic effect.

Further studies, performed by the same group of authors, support the above suggestion. They have observed that vitamin E deficiency resulted in a significant increase in epithelial cell necrosis during goitre development and involution, and in increased lipid peroxidation and decreased GSH-Px activity in the thyroid (59).

Studies in humans suggest that iodine deficiency is associated with an increased oxidative stress and decreased antioxidative defence. A decreased SOD activity was observed in endemic

goiter tissue (45). In iodine-deficient children with goitre, the activities of antioxidative enzymes (GSH-Px, CAT and SOD) in erythrocytes, and selenium concentration in plasma and erythrocytes, have been found to be significantly lower than their respective values in non-goiter and non iodine-deficient control subjects, and even, what is most important, than in non-goiter but iodine-deficient children (60). Thus, it seems plausible that low activities of antioxidant enzymes and low selenium concentrations contribute to thyroid enlargement in iodine-deficient subjects.

In another study, increased damage to genomic DNA (evaluated by the measurement of the concentration of different oxidized bases in peripheral blood) was found in children (aged 15-8 years) with goitre, living in the area of severe or moderate iodine deficiency; DNA damage was accompanied by decreased blood concentration of thyroid free hormones, decreased concentration of selenium,

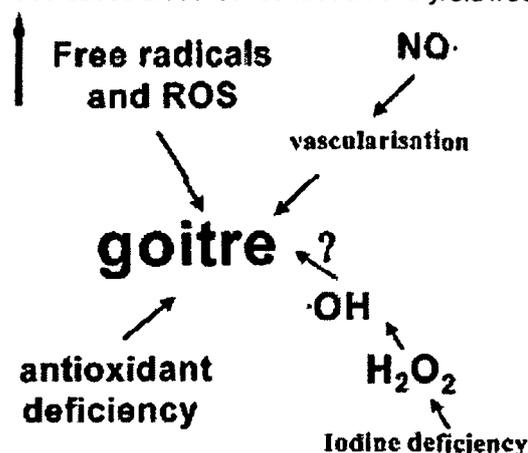


Fig. 1 The proposed mechanism - related to oxidative stress - of thyroid enlargement (goitrogenesis). H_2O_2 , hydrogen peroxide; $NO\cdot$, nitric oxide; $\cdot OH$, hydroxyl radical; ROS, reactive oxygen species; TSH, thyrotropin

decreased activity of GSH-Px and SOD in blood erythrocytes, and by decreased ioduria (61). The observation that iodine deficiency, resulting in goitre, leads to oxidative DNA damage, supports the view on an essential relationship between iodine deficiency and the increased incidence of thyroid cancer.

It has recently been found that melatonin concentration, measured at night, was significantly higher after than before the operation performed in patients with very large non-toxic nodular goitre (62). These results, for some reasons - unexpected, could be explained as follows: melatonin might actively be taken up by an enlarged thyroid with a subsequent decrease in blood concentration of the indoleamine before surgery.

The proposed mechanism - related to oxidative stress - of thyroid enlargement (goitrogenesis) is presented in Figure 1.

Thyroid autoantigens and oxidative stress

Thyroglobulin (Tg) and TPO belong to the main thyroid autoantigens. Thyroglobulin is stored extracellularly in the thyroid follicle lumen and is essential for thyroid hormone biosynthesis. During thyroid hormone synthesis, resulting from several oxidative reactions, the multimerized inactive form of Tg (mTg) is formed (63). It has recently been found that experimentally-induced oxidative stress participates in mTg fragmentation, producing Tg molecules with a recovered ability to store thyroid hormones; at the same time, the fragmentation of mTg precludes its excessive accumulation in the thyroid and, consequently, the thyroid enlargement (63).

Under experimental conditions of Fenton reaction or in the presence of ferric ions (Fe^{3+}) - both causing oxidative stress - a production of immunoreactive C-terminal fragments (40 kDa) of Tg

was observed and that process was accompanied by thyroid hormone synthesis (64). Using human thyroid cells, H_2O_2 -being in excess - was shown to produce the same 40 kDa immunoreactive fragments of Tg; it is worth stressing that both H_2O_2 and iodide have been indispensable to cause Tg fragmentation (65), what suggests that Tg cleavage occurs during thyroid hormone synthesis. Of great importance is the fact that immunoreactive Tg fragments have been found only in dead cells but not in living ones and that those fragments are able to enter living thyrocytes (65), what would start autoimmune response.

Concerning another thyroid autoantigen - TPO - worth mentioning are studies on oxidative stress and myeloperoxidase (MPO), which is very similar in structure and properties to TPO. Whole body ionizing radiation (800 cGy) resulted in a decreased level of glutathione, increased levels of lipid peroxidation products and increased levels of MPO (as the index of neutrophil infiltration) in different tissues, the changes which were reduced by a co-administration of melatonin (66). It is highly probable that melatonin is also able to modify the level of TPO protein under conditions of excessive oxidative stress. No studies have till now been performed, showing that oxidative stress is involved in autoimmunity directed against TPO. Some other results are worth mentioning, concerning the relationship between oxidative stress and thyroid autoimmunity.

It has been documented that free radicals are involved in interleukin - 1β - induced glycosaminoglycan production by retro-ocular fibroblasts and their accumulation in patients with Graves' disease (67). Furthermore, it has been shown that $NO\cdot$ is involved in interleukin - 1α - induced cytotoxicity in polarized human thyrocytes, suggesting that this free radical may promote the exposure of autoantigens to the immune system (68).

The relationship between oxidative stress and thyroid autoimmunity suggests a potential protective role of antioxidants in autoimmune diseases. However, the application of antioxidants in patients with autoimmune diseases or patients, predisposed genetically to these disorders, is still the point of discussion. As far as the influence of melatonin on the immune system is concerned, divergent results exist (69, 70). However, the precise effect, caused by the indoleamine in patients with autoimmune diseases, as well as the potential indications and contraindications for the treatment with this substance in such patients remain to be determined.

Effects of thyroid hormones on oxidative processes

Thyroid hormones regulate energy metabolism, among others, by their effects on mitochondria (71), in which free radicals and ROS are produced. Under *in vitro* conditions, thyroid hormones may act as antioxidants; thyroid hormones [3,5,3'-triiodothyronine, T_3 ; and 3,5,3',5'-tetraiodothyronine (thyroxine), T_4] and their structural analogues (L-thyronine, T_0 ; 3,5,3'-triiodothyroacetic acid, TA_3 ; 3,5,3',5'-tetraiodothyroacetic acid, TA_4) were found to reveal antioxidative properties (72-74). For example, using different models of oxidative stress, it was shown that all the above mentioned hormones or their analogues revealed a capacity to scavenge free radicals, $\cdot OH$ and $O_2\cdot^-$ included (shown for T_3), and that T_0 , T_3 , and TA_3 effectively prevented the formation of lipid peroxidation products - conjugated dienes and thiobarbituric acid reactive substances - during LDL oxidation (74). It was documented in the same study that the 4'-hydroxydiphenylether structure of thyroid compounds was necessary for their free radical scavenging activity. This has been confirmed by antioxidant effects of T_0 , which has the basic structure of thyroid hormones but does not possess iodine atom; conversely, monoiodotyrosine (MIT), which possesses the mono-iodosubstituted phenolic ring of T_3 , does not reveal any antioxidative effects (74).

Effects of thyroid hormone excess

Changes in the values of oxidative parameters in different tissues were observed, due to thyroid hormone excess resulting from either thyroid diseases or treatment with thyroid hormones. In blood samples, collected from hyperthyroid patients suffering from Graves' disease, the levels of thiobarbituric acid-reacting substances and conjugated dienes (both being parameters of oxidative stress), and the activities of main antioxidative enzymes (SOD, CAT and GSH-Px) were significantly higher than those in healthy subjects: all those changes were - to a differential degree - restored in result of antithyroid drug treatment (75).

Similarly, a significant increase in malondialdehyde (MDA) concentration and a slight increase in Schiff's bases and conjugated dienes concentration (all being parameters of oxidative damage to lipids) were found in blood serum collected from hyperthyroid patients with Graves' disease; treatment with methimazole (an antithyroid drug) brought about a decrease in the values of indices of oxidative stress, accompanied by normalization of thyroid hormone and TSH concentration (76).

In mononuclear cells, collected from hyperthyroid patients with Graves' disease, an increased level of 8-oxo-7,8-dihydro-2'-deoxyguanosine (8-oxo-dGuo) (the parameter of DNA oxidative damage) was found; additionally, 8-oxo-dGuo level correlated significantly with free T_4 concentration (77). It is presumed that untreated hyperthyroidism contributes to oxidative damage to DNA.

An induction of oxidative stress was also observed in experimental models of hyperthyroidism. The treatment of rats with thyroid hormones caused a production of $O_2^{\cdot -}$ and H_2O_2 in liver mitochondria (78) and of NO^{\cdot} in liver cytosol (79). Expectedly, T_3 -injection increased lipid peroxidation and protein oxidation in rat liver; it is worth stressing that the damage to lipids preceded the damage to protein molecules in response to thyroid hormone, what suggests different susceptibility of target molecules to excessive amounts of thyroid hormones (80).

In animal models of hyper- and hypothyroidism, T_3 - treatment in rats brought about a significant increase in lipid peroxidation in the liver, the heart, and skeletal muscles, whereas methimazole administration (causing hypothyroidism) did not influence the level of lipid peroxidation products in those organs; at the same time, a decrease in the whole antioxidant capacity of tissues and an increase in GSH-Px and glutathione reductase (GSH-Rd) were observed in only some examined organs in both hyper- and hypothyroid rats (81).

In other animal models of hyper- and hypothyroidism, $L-T_4$ treatment in *OF1* female mice resulted in an increased sensitivity to lipid peroxidation, an increased level of oxidized GSH (GSSG), and an increased ratio GSSG/GSH, as well as increased oxidative damage to mitochondrial DNA in the heart; interestingly, a treatment with an antithyroid drug - propylthiouracil (PTU) - did not influence oxidative damage to lipids and decreased GSSG concentration and the GSSG/GSH ratio, as well as reduced oxidative damage to mitochondrial DNA (82). The latter finding is of great importance, suggesting protective effects of PTU against oxidative stress. In agreement with this, $L-T_4$, but not PTU, treatment in *OF1* female mice increased lipid peroxidation in the skeletal muscle (83).

In the study, performed at our laboratory, a 2-week-treatment with $L-T_4$ (100 μ g/kg B.W., for 14 days) resulted in an increased concentration of free fractions of both thyroid hormones; interestingly, a co-treatment with melatonin (5 mg/kg B.W., for 7 days) completely prevented the increase in free T_3 concentration (84). Thus, melatonin has abolished the increase in the concentration of this thyroid hormone, which is directly responsible for tissue effects of hyperthyroidism. At the same time, it was observed that melatonin injection resulted in decreased basal values of conjugated dienes and Schiff's bases in rat kidney (84). It has recently been found that T_3 causes mutagenic action through induction of oxidative stress (85).

Effects of thyroid hormone deficiency

It has been observed that hypothyroidism, resulting from either PTU-treatment or thyroidectomy, causes a decrease in H_2O_2 generation (86). Additionally, hypothyroidism resulted from thyroidectomy, prevented post-ischemic lipid peroxidation in rat kidney (87).

Thus, it is still unclear, if the above-mentioned protective effects of PTU against oxidative stress (82) are really related to direct antioxidant action of the drug or to hypothyroidism, or if they result from both mechanisms. Indeed, PTU has been shown to react directly with $\cdot OH$ and, to a lesser extent, with O_2^- , protecting against lipid peroxidation (88), and to prevent the formation of hypochlorous acid (another oxygen species) (89).

On the other hand, however, it should be expected that hypothyroid status, which constitutes a pathological condition, leads, sooner or later, to oxidative damage of biological molecules. In agreement with the last assumption, progressive hypothyroidism, observed early in postnatal rats, enhanced oxidative processes; increased SOD and CAT activities, and decreased GSH level were accompanied by an increased amount of $\cdot OH$, enhanced protein carbonylation and lipid peroxidation in rat brain (90).

It is known that in aging humans, the function of several endocrine glands declines progressively, resulting in decreased concentrations of different hormones, among others of melatonin and of thyroid hormones. The study, performed in perimenopausal and menopausal women, with initial low levels of blood melatonin, revealed that melatonin treatment for 3-6 months resulted in a significant increase in thyroid hormone concentrations (91). Thus, melatonin reveals a recovery effect on thyroid function towards a more juvenile pattern of regulation (91); it is not excluded that the described effect is a direct one because a similar treatment with melatonin in aging patients did not result in any changes of TSH concentration (92).

Radioiodine (^{131}I) - induced oxidative stress

Radioiodine (^{131}I) therapy is a treatment of choice in differentiated thyroid cancer (after total thyroidectomy) and in hyperthyroidism. In one of the studies, the parameters of oxidative stress were evaluated in patients subjected to total thyroidectomy because of thyroid carcinoma, before (at that time patients were hypothyroid) and after ^{131}I treatment (93). The level of lipid peroxidation products, measured in blood erythrocytes, significantly increased after thyroidectomy, comparing to values in the controls; a further increase in lipid peroxidation was found after ^{131}I treatment. Whereas GSH level and GSH-Px and GSH-Rd activities were lower in thyroidectomized patients than those in the control subjects, the values of those parameters significantly increased after ^{131}I treatment.

In another study, a treatment with ^{131}I , using either higher (in case of thyroid cancer) or lower (in case of hyperthyroidism) activities, resulted in increased levels of isoprostanes (products of oxidative injury to lipids) in blood plasma, serum and urine; the damaging effect of ^{131}I was significantly higher and longer lasting after higher-activity therapy (94). The above findings are in agreement with other numerous data, indicating that radiation causes oxidative damage to tissues, while antioxidants, like melatonin, may prevent it (8). Because of the potential role of ionizing radiation in the pathogenesis of thyroid cancer, the studies on protective effects of melatonin against radiation-induced oxidative stress and thyroid cancer seem to be of special value. It is worth mentioning here that melatonin effectively reduced histoenzymological changes in the rat thyroid gland caused by exposure to γ -radiation (95).

Summing up, oxidative stress plays a significant role in the thyroid gland under both physiological and pathological conditions. Melatonin, as an antioxidant and widely distributed molecule, may constitute a modulator of physiological processes in the thyroid, and under pathological

conditions may protect against oxidative damage in the gland and in other tissues. This assumption requires further experimental evidence.

ACKNOWLEDGEMENTS

Some parts of the research were supported by the Medical University project No. 502-11-806.

REFERENCES

- 1 Dreher D and Junod AF (1996). Role of oxygen free radicals in cancer development. *Eur J Cancer* **32**: 30-38.
- 2 Droge W (2002). Free radicals in the physiological control of cell function. *Physiol Rev* **82**: 47-95.
- 3 Kvetnoy IM (2002). Extrapineal melatonin in pathology: new perspectives for diagnosis, prognosis, and treatment of illness. *Neuroendocrinol Lett* **23**(Suppl 1): 92-96.
- 4 Tan DX, Chen LD, Poeggeler B, Manchester LC and Reiter RJ (1993). Melatonin: a potent, endogenous hydroxyl radical scavenger. *Endocrine J* **1**: 57-60.
- 5 Reiter RJ (1998). Oxidative damage in the central nervous system: Protection by melatonin. *Prog Neurobiol* **56**: 359-384.
- 6 Reiter RJ, Tan DX, Qi W, Manchester LC, Karbownik M and Calvo JR (2000). Pharmacology and physiology of melatonin in the reduction of oxidative stress *in vivo*. *Biol Signals Recept* **9**: 160-171.
- 7 Hardeland R, Poeggeler B, Niebergall R and Zelosko V (2003). Oxidation of melatonin by carbonate radicals and chemiluminescence emitted during pyrrole ring cleavage. *J Pineal Res* **34**: 17-25.
- 8 Karbownik M and Reiter RJ (2000). Antioxidative effects of melatonin in protection against cellular damage caused by ionizing radiation. *Proc Soc Exp Biol Med* **225**: 9-22.
- 9 Karbownik M and Reiter RJ (2002). Melatonin protects against oxidative stress caused by δ -aminolevulinic acid: Implication for cancer reduction, *Cancer Invest* **20**: 276-286.
- 10 Lewinski A, Vaughan MK, Champney TH, Reiter RJ and Smith NKR (1984). Dark exposure inhibits the mitotic activity of thyroid follicular cells in male mice with intact pineal. *Experientia* **40**: 1284-1285.
- 11 Lewinski A, Wajs E and Krotewicz M (1993). Melatonin and other indolic substances: their influence on thyroid growth and secretion. In: Touitou Y, Arendt J and Pevet P (eds.), *Melatonin and the Pineal Gland - From Basic Science to Clinical Application*, Excerpta Medica. Amsterdam, pp. 265-268.
- 12 Lewinski A, Wajs E, Modrzejewska H, Klencki M, Karbownik M and Greger J (1994). Inhibitory influence of melatonin on thymidine kinase activity in the rat thyroid lobes incubated *in vitro*. *Neuroendocrinol Lett* **16**: 221-226.
- 13 Lewinski A and Sewerynek E (1986). Melatonin inhibits the basal and TSH-stimulated mitotic activity of thyroid follicular cells *in vivo* and in organ culture. *J Pineal Res* **3**: 291-299.
- 14 Wajs E and Lewinski A (1992). Inhibitory influence of late-afternoon melatonin injections and the counter-inhibitory effect of melatonin pellets on thyroid growth processes in male Wistar rats; comparison with effects of other indole substances. *J Pineal Res* **13**: 158-166.
- 15 Krotewicz M, Lewinski A and Wajs E (1992). The inhibitory effect of late afternoon melatonin injections, but not of melatonin-containing subcutaneous implants, on thyroid hormone secretion in male Wistar rats. *Neuroendocrinol Lett* **14**: 405-411.

- 16 Krotewicz M and Lewinski A (1994). Effects of pinealectomy and of late afternoon injections of pineal indole substances on thyroid hormone secretion in male Wistar rats. *Biochem Lett* **50**: 101-107.
- 17 Wright ML, Cuthbert KL, Donohue MJ, Solano SD and Proctor KL (2000). Direct influence of melatonin on the thyroid and comparison with prolactin. *J Exp Zool* **286**: 625-631.
- 18 Lewinski A, Sewerynek E and Karbownik M (2002). Melatonin from the past into the future - our own experience. In: Haldar C, Singaravel M and Maitra SK (eds.), *Treatise on Pineal Gland and Melatonin*, Science Publishers Inc. Enfield (NH), USA, and Plymouth, UK pp. 157-175.
- 19 Lewinski A (1990). Some aspects of the pineal-thyroid interrelationship and their possible involvement in the regulation of function and growth of these two glands. In: Reiter RJ and Lukaszuk A (eds.), *Advances in Pineal Research*, Vol. IV, John Libbey and Company Ltd., London, pp. 175-188
- 20 Lewinski A, Wajs E, Klencki M, Krotewicz M and Bilinski P (1996). Melatonin- induced inhibition of thyroid growth processes and possible mechanisms of that phenomenon. In: Haldar C (ed.), *Recent Researches in Biology*, Vol. 1, *The Pineal Gland: Its Molecular Signals*, Hindustan Publishing Corporation, New Delhi, pp. 27-32.
- 21 Lewinski A, Wajs E, Klencki M, Karbownik M, Gesing A, Sewerynek E, Slowinska- Klencka D, Skowronska-Jozwiak E, Bilinski P and Krotewicz M (1997). Pineal-thyroid interrelationships update. In: Webb SM, Puig - Doming M, Moller M and Pevet E (eds.), *Pineal Update from Molecular Mechanisms to Clinical Implications*, PJD Publications Limited, Westbury, New York, pp. 173-181.
- 22 Lewinski A and Karbownik M (2002). Melatonin and the thyroid gland. *Neuroendocrinol Lett* **23** (suppl 1): 73-78.
- 23 Karbownik M and Lewinski A (2003). The role of oxidative stress in physiological and pathological processes in the thyroid gland; possible involvement in pineal-thyroid interactions. *Neuroendocrinol Lett* **24**: 293-303.
- 24 Bjorkman U and Ekholm R (1984). Generation of H₂O₂ in isolated porcine thyroid follicles. *Endocrinology* **115**: 392-398.
- 25 Raspe E and Dumont JE (1995). Tonic modulation of dog thyrocyte H₂O₂ generation and I⁻ uptake by thyrotropin through the cyclic adenosine 3',5'- monophosphate cascade. *Endocrinology* **135**: 965-973.
- 26 Dupuy C, Ohayon R, Valent A, Noel-Hudson MS, Deme D and Virion A (1999). Purification of a novel flavoprotein involved in the thyroid NADPH oxidase: cloning of the porcine and human cDNAs. *J Biol Chem* **274**: 37265-37269.
- 27 DeDeken X, Wang D, Many MC, Costagliola S, Libert F, Vassart G, Dumont JE and Miot F (2000). Cloning of two human thyroid cDNAs encoding new members of the NADPH oxidase family. *J Biol Chem* **275**: 23227-23233.
- 28 Sugawara M, Sugawara Y, Wen K and Giulivi C (2002). Generation of oxygen free radicals in thyroid cells and inhibition of thyroid peroxidase. *Exp Biol Med* **227**: 141-146.
- 29 Eaton JW and Qian M (2002). Molecular bases of cellular iron toxicity. *Free Radic Biol Med* **32**: 833-840.
- 30 Zimmermann M and Kohrle J (2002). The impact of iron and selenium deficiencies on iodine and thyroid metabolism: biochemistry and relevance to public health. *Thyroid* **12**: 867-878.

- 31 Fayadat L, Niccoli-Sire P, Lanet J and Franc J-L (1999). Role of heme in intracellular trafficking of thyroperoxidase and involvement of H₂O₂ generated at the apical surface of thyroid cells in autocatalytic covalent heme binding. *J Biol Chem* **274**: 10533-10538.
- 32 Corvailain B, Van Sande J and Dumont JE (1988). Inhibition by iodide binding to proteins: the "Wolff-Chaikoff's effect is caused by inhibition of H₂O₂ generation. *Biochem Biophys Res Commun* **154**: 1287-1292.
- 33 Riou C, Remy C, Rabilloud R, Rousset B and Fontlupt P (1998). H₂O₂ induces apoptosis of pig thyrocytes in culture. *J Endocrinol* **156**: 315-322.
- 34 Tan DX, Manchester LC, Reiter RJ, Plummer BF, Limson J, Weintraub ST and Qi W (2000). Melatonin directly scavenges hydrogen peroxide: a potentially new metabolic pathway of melatonin biotransformation. *Free Radic Biol Med* **29**: 1177-1185.
- 35 Esteves RZ, van Sande J and Dumont JE (1992). Nitric oxide as a signal in thyroid. *Mol Cell Endocrinol* **90**: R1-3.
- 36 Millatt LJ, Jackson R, Williams BC and Whitley GS (1993). Nitric oxide stimulates cyclic GMP in human thyrocytes. *J Mol Endocrinol* **10**: 163-169.
- 37 Colin IM, Nava E, Toussaint D, Maiter DM, VanDenhove MF, Luscher TF, Ketelslegers JM, Deneef JF and Jameson JL (1995). Expression of nitric oxide synthase isoforms in the thyroid gland: evidence for a role of nitric oxide in vasculatur control during goiter formation. *Endocrinology* **136**: 5283-5290.
- 38 Motohashi S, Kasai K, Banba N, Hattori Y and Shimoda S (1996). Nitric oxide inhibits cell growth in cultured human thyrocytes. *Life Sci* **59**: PL227-234.
- 39 Bocanera LV, Krawiec L, Silberschmidt D, Pignatoro O, Juvenal GJ, Pregliasco LB and Pisarev MA (1997). Role of cyclic 3'5'guanosine monophosphate and nitric oxide in the regulation of iodide uptake in calf thyroid cells. *J Endocrinol* **155**: 451-457.
- 40 Millatt LJ, Johnstone AP and Whitley GS (1998). Nitric oxide enhances thyroid peroxidase activity in primary human thyrocytes. *Life Sci* **63**: PL373-380.
- 41 Noda Y, Mori A, Liburdy R and Packer L (1999). Melatonin and its precursors scavenge nitric oxide. *J Pineal Res* **27**: 159-163.
- 42 Gilad E, Cuzzocrea S, Zingarelli B, Salzman AL and Szabo C (1997). Melatonin is a scavenger of peroxynitrite. *Life Sci* **60**: PL 169-74.
- 43 Bromme H-J, Ebelt H, Peschke D and Peschke E (1999). Alloxan acts as a prooxidant only under reducing conditions: influence of melatonin. *Cell Mol Life Sci* **55**: 487-493.
- 44 Taurog A (2000). Hormone synthesis. In: Braverman LE and Utiger RD (eds.), *The Thyroid*, Lippincott Williams and Wilkins, Philadelphia, pp. 52-90.
- 45 Sugawara M, Kita T, Lee ED, Takamatsu J, Hagen GA, Kuma K and Medeiros-Neto GA (1988). Deficiency of thyroid superoxide dismutase in endemic goiter tissue. *J Clin Endocrinol Metab* **67**: 1156-1161.
- 46 Iwase K, Nagasaka A, Kato K, Ohtani S, Tsujimura T, Inagaki A, Jimbo S, Nakai QA, Masunaga R, Hamada M, Mano T, Kotake M and Miura K (1993). Localization of Cu/Zn and Mn superoxide dismutase in various thyroid disorders. *Acta Endocrinol* **129**: 573-578.
- 47 Sadani GR and Nadkarni GD (1996). Role of tissue antioxidant defence in thyroid cancers. *Cancer Lett* **109**: 231-235.

- 48 Rhee SG (1999). Redox signaling: hydrogen peroxide as intracellular messenger. *Exp Mol Med* **31**: 53-59.
- 49 Mano T, Iwase K, Hayashi R, Hayakawa N, Uchimura K, Makino M, Nagata M, Sawai Y, Oda N, Hamada M, Aono T, Nakai A, Nagasaka A and Itoh M (1998). Vitamin E and coenzyme Q concentrations in the thyroid tissues of patients with various thyroid disorders. *Am J Med Sci* **315**: 230-232.
- 50 Nakamura M and Ohtaki S (1993). Formation and reduction of ascorbate radicals by hog thyroid microsomes. *Arch Biochem Biophys* **305**: 84-90.
- 51 Kim H, Lee T-H, Park ES, Suh JM, Park SJ, Chung HK, Kwon O-Y, Kim YK, Ro HK and Shong M (2000). Role of peroxiredoxins in regulating intracellular hydrogen peroxide and hydrogen peroxide-induced apoptosis in thyroid cells. *J Biol Chem* **275**: 18266-18270.
- 52 Kvetnoy IM and Yuzhakov VV (1993). Extrapyneal melatonin: advances in microscopical identification of hormones in endocrine and nonendocrine cells. *Microsc Anal* **21**: 27-29
- 53 Mano T, Shinohara R, Iwase K, Kotake M, Hamada M, Uchimura K, Hayakawa N, Hayashi R, Nakai A, Ishizuki Y and Nagasaka A (1997) Changes in free radical scavengers and lipid peroxide in thyroid glands of various thyroid disorders. *Horm Metab Res* **29**: 351-354.
- 54 Yanagawa T, Ishikawa T, Ishii T, Tabuchi K, Iwasa S, Bannai S, Omura K, Suzuki H and Yoshida H (1999). Peroxiredoxin I expression in human thyroid tumors. *Cancer Lett* **145**: 127-132.
- 55 Kitano H, Kitanishi T, Nakanishi Y, Suzuki M, Taleuchi E, Yazawa Y, Kitajima K, Kimura H and Tooyama I (1999). Expression of inducible nitric oxide synthase in human thyroid papillary carcinoma. *Thyroid* **9**: 113-117.
- 56 Hasegawa Y, Takano T, Miyauchi A, Matsuzuka F, Yoshida H, Kuma K and Amino N (2003). Decreased expression of catalase mRNA in thyroid anaplastic carcinoma. *Jpn J Clin Oncol* **33**: 6-9.
- 57 Karbownik M and Lewinski A (2003). Melatonin reduces Fenton reaction- induced lipid peroxidation in porcine thyroid tissue. *J Cell Biochem* **90**: 806-811.
- 58 Mutaku JF, Many MC, Colin I, Deneff JF and van den Hove MF (1998). Antigoitrogenic effect of combined supplementation with dl-alpha-tocopherol, ascorbic acid and beta-carotene and of dl-alpha-tocopherol alone in the rat. *J Endocrinol* **156**: 551-561.
- 59 Mutaku JF, Poma JF, Many MC, Deneff JF and Van Den Hove MF (2002). Cell necrosis and apoptosis are differentially regulated during goitre development and iodine-induced involution. *J Endocrinol* **172**: 375-386.
- 60 Giray B, Hincal F, Tezic T, Okten A and Gedik Y (2001). Status of selenium and antioxidant enzymes of goitrous children is lower than healthy controls and nongoitrous children with high iodine deficiency. *Biol Trace Elem Res* **82**: 35-52.
- 61 Giray B and Hincal F (2002). Oxidative DNA base damage, antioxidant enzyme activities and selenium status in highly iodine-deficient goitrous children. *Free Rad Res* **36**: 55-62.
- 62 Karasek M, Stankiewicz A, Bandurska-Stankiewicz E, Zylinska K, Pawlikowski M and Kuzdak K (2000). Melatonin concentrations in patients with large goiter before and after surgery. *Neuroendocrinol Lett* **21**: 437-439.
- 63 Delom F, Lejeune P-J, Vinet L, Carayon P and Mallet B (1999). Involvement of oxidative reactions and extracellular protein chaperons in the rescue of misassembled thyroglobulin in the follicular lumen. *Biochem Biophys Res Commun* **255**: 438-443.

- 64 Duthoit C, Estienne V, Delom F, Durand-Gorde J-M, Mallet B, Carayon P and Ruf J (2000). Production of immunoreactive thyroglobulin C-terminal fragments during thyroid hormone synthesis. *Endocrinology* **141**: 2518-2525.
- 65 Duthoit C, Estienne V, Giraud A, Durand-Gorde J-M, Rasmussen AK, Feldt-Rasmussen U, Carayon P and Ruf J (2001). Hydrogen peroxide-induced production of a 40 kDa immunoreactive thyroglobulin fragment in human thyroid cells: the onset of thyroid autoimmunity. *Biochem J* **360**: 557-562.
- 66 Sener G, Jahovic N, Tosun O, Atasoy BM and Yegen BC (2003). Melatonin ameliorates ionizing radiation-induced oxidative organ damage in rats. *Life Sci* **74**: 563-572.
- 67 Lu R, Wang P, Wartofsky L, Sutton BD, Zweier JL, Bahn RS, Garrity J and Burman KD (1999). Oxygen free radicals in interleukin-1beta-induced glycosaminoglycan production by retro-ocular fibroblasts from normal subjects and Graves' ophthalmopathy patients. *Thyroid* **9**: 297-303.
- 68 Van den Hove M-F, Stenoiu MS, Croizet K, Couvreur M, Courtoy PJ, Devuyt O and Colin IM (2002). Nitric oxide is involved in interleukin-1 α - induced cytotoxicity in polarised human thyrocytes. *J Endocrinol* **173**: 177-185.
- 69 Lewinski A, Zelazowski P, Sewerynek E, Zerek-Melen G, Szkudlinski M and Zelazowska E (1989). Melatonin-induced suppression of human lymphocyte natural killer activity *in vitro*. *J Pineal Res* **7**: 153-164.
- 70 Guerrero JM and Reiter RJ (2002). Melatonin-immune system relationship. *Curr Top Med Chem* **2**: 167-179.
- 71 Goglia F, Silvestri E and Lanni A (2002). Thyroid hormones and mitochondria. *Biosci Rep* **22**: 17-32.
- 72 Hanna AN, Feller DR, Witiak DT and Newman HAI (1993). Inhibition of low density lipoprotein oxidation by thyronines and probucol. *Biochem Pharmacol* **45**: 753-762.
- 73 Chomard P, Seguin C, Loireau A, Autissier N and Artur Y (1998). Effects of iodotyrosines, thyronines, iodothyroacetic acids and thyromimetic analogues on *in vitro* copper-induced oxidation of low-density lipoproteins. *Biochem Pharmacol* **55**: 1591-1601.
- 74 Oziol L, Faure P, Vergely C, Rochette L, Artur Y and Chomard P (2001). *In vitro* free radical scavenging capacity of thyroid hormones and structural analogues. *J Endocrinol* **170**: 197-206.
- 75 Komosinska-Vassev K, Olczyk K, Kucharz EJ, Marcisz C, Winsz-Szczotka K and Kotulska A (2000). Free radical activity and antioxidant defense mechanism in patients with hyperthyroidism due to Graves' disease during therapy. *Clin Chim Acta* **300**: 107-117.
- 76 Sewerynek E, Wiktorska J, Nowak D and Lewinski A (2000). Methimazole protection against oxidative stress induced by hyperthyroidism in Graves' disease. *Endocr Regul* **34**: 83-89.
- 77 Hara H, Sato R and Ban Y (2001). Production of 8-OHdG and cytochrome c by cultured human mononuclear cells in patients with autoimmune thyroid disease. *Endocrine J* **48**: 671-675.
- 78 Fernandez V and Videla LA (1993). Influence of hyperthyroidism on superoxide radical and hydrogen peroxide production by rat liver submitochondrial particles. *Free Radic Res Commun* **18**: 329-335.
- 79 Fernandez V, Cornejo P, Tapia G and Videla LA (1997). Influence of hyperthyroidism on the activity of liver nitric oxide synthase in the rat. *Nitric Oxide* **1**: 463-468.

- 80 Tapia G, Cornejo P, Fernandez V and Videla LA (1999). Protein oxidation in thyroid hormone-induced liver oxidative stress: relation to lipid peroxidation. *Toxicol Lett* **106**: 209-214.
- 81 Venditti P, Balestrieri M, Di Meo S and De Leo T (1997). Effect of thyroid state on lipid peroxidation, antioxidant defences, and susceptibility to oxidative stress in rat tissues. *J Endocrinol* **155**: 151-157.
- 82 Gredilla R, Torres ML, Portero-Otin M, Pamplona R and Barja G (2001). Influence of hyper- and hypothyroidism on lipid peroxidation, unsaturation of phospholipids, glutathione system and oxidative damage to nuclear and mitochondrial DNA in mice skeletal muscle. *Mol Cell Biochem* **221**: 41-48.
- 83 Gredilla R, Barja G and Lopez-Torres M (2001). Thyroid hormone-induced oxidative damage on lipids, glutathione and DNA in the mouse heart. *Free Radic Res* **35**: 417-425.
- 84 Sewerynek E, Wiktorska J and Lewinski A (1999). Effects of melatonin on the oxidative stress induced by thyrotoxicosis in rats. *Neuroendocrinol Lett* **20**: 157-161.
- 85 Djelic N and Anderson D (2003). The effect of the antioxidant catalase on oestrogens, triiodothyronine, and noradrenaline in the Comet assay. *Teratog Carcinog Mutagen (Suppl)* **2**: 69-81.
- 86 Swaroop A and Ramasarma T (1985). Heat exposure and hypothyroid conditions decrease hydrogen peroxide generation in liver mitochondria. *Biochem J* **226**: 403-408.
- 87 Paller MS (1986). Hypothyroidism protects against free radical damage in ischemic acute renal failure. *Kidney Int* **29**: 1162-1166.
- 88 Hicks M, Wong LS and Day RO (1992). Antioxidant activity of propylthiouracil. *Biochem Pharmacol* **43**: 439-444.
- 89 Ross AD, Dey I, Janes N and Israel Y (1998). Effect of antithyroid drugs on hydroxyl radical formation and α -1-proteinase inhibitor inactivation by neutrophils: therapeutic implications. *J Pharmacol Exp Ther* **285**: 1233-1238.
- 90 Rahaman SO, Ghosh S, Mohanakumar KP, Das S and Sarkar PK (2001). Hypothyroidism in the developing rat brain is associated with marked oxidative stress and aberrant intraneuronal accumulation of neurofilaments. *Neurosci Res* **40**: 273-279.
- 91 Bellipanni G, Bianchi P, Pierpaoli W, Bulian D and Ilyia E (2001). Effects of melatonin in perimenopausal and menopausal women: a randomized and placebo controlled study. *Exp Gerontol* **36**: 297-310.
- 92 Siegrist C, Benedetti C, Orlando A, Beltran JM, Tuchscher L and Nosedá CM (2001). Lack of changes in serum prolactin, FSH, TSH, and estradiol after melatonin treatment in doses that improve sleep and reduce benzodiazepine consumption in sleep-disturbed, middle-aged, and elderly patients. *J Pineal Res* **30**: 34-42.
- 93 Konukoglu D, Hatemi HH, Arikan S, Demir M and Akcay T (1998). Radioiodine treatment and oxidative stress in thyroidectomised patients for differentiated thyroid cancers. *Pharmacol Res* **38**: 311-315.
- 94 Wolfram RM, Budinsky AC, Palumbo B, Palumbo R and Sinzinger H (2002). Radioiodine therapy induces dose-dependent *in vivo* oxidation injury: evidence by increased isoprostane 8-epi-PGF(2 α). *J Nucl Med* **43**: 1254-1258.
- 95 Kundurovic Z and Sceповic M (1989). Histoenzymological reactions of the thyroid gland in irradiated and previously melatonin-treated irradiated rats. *Acta Medica Jugosl* **43**: 337-347.