



Tabernaemontana divaricata: A Herbal Panacea

Sanjita Das*, Anupam Dubey and Divya

Noida Institute of Engineering and Technology (Pharmacy Institute),
Plot No. 19, Knowledge Park-II, Greater Noida, Uttar Pradesh - 201306, India; sanjita8@yahoo.co.in

Abstract

Tabernaemontana divaricata (pinwheel flower) is a flowering plant that can grow easily in gardens and along roadsides. This plant can be cultivated in every condition. No specific environmental condition is required for the growth of the plant. Growing evidence suggests that this plant has medicinal benefits for various diseases due to the presence of bioactive components in the plant. The plant is extensively found near Indian heritage to be used for worship. *T. divaricata* contains major alkaloids like apparicine, conophylline, coronardine, ibogamine, etc., exhibiting pharmacological activities. Their major pharmacological potential is against inflammation, pain, and other diseases. Plants' major activities, such as anti-diabetic, anti-inflammatory, antibacterial, antifungal, and so on, have been demonstrated by their responsible bioactive compounds. The review is to highlight the researchers' findings of different medicinal activities in *T. divaricata* along with the major responsible phytochemicals. There is a lot more scope for further research, which can be extended by the help of this review.

Keywords: Alkaloids, Bioactive Components, Latex, Pharmacological Activity, *T. divaricata*

1. Introduction

The plant which has medicinal value was always invaluable for humans and as the time passes, its utility will also increase. Compound that are found naturally are taken as safer in comparison to synthetic compound. Plants with medicinal value are more difficult to develop drug resistance than synthetic compounds¹. The plant *Tabernaemontana divaricata* also known as crepe jasmine and pinwheel flower, is from the family *Apocynaceae*. A plant that grows in spring is as graceful as it is evergreen. This plant can be found throughout South Asia as well as the countries of Southeast Asia. It is considered an ornamental plant and is abundantly found in Indian heritage to worship the god and goddess. The phytochemical, nonalkaloids

and alkaloids constituents such as flavonoids, phenylpropanoids, terpenoids, enzymes and steroids from the parts of the plant (stem, root, flower and leaves) have been reported. Along with its well-known analgesic and antidiarrheal properties, plant parts are also used as liver, spleen, and brain tonics; and it has been discovered that *T. divaricata* extract has antioxidant, anti-inflammatory, and reversible acetylcholinesterase inhibition properties²⁻⁴. A sticky milky liquid called latex comes out from the points of laticiferous tissue and contains secondary metabolites and proteins⁵⁻⁸. A milky fluid found in nature in 10% of all flowering plants is Latex⁶. The plants of the family *Apocynaceae* have latex as one of their pertinent features⁹. The easy and plentiful availability of *T. divaricata* as an ornamental plant and its medicinal importance created

*Author for correspondence

this review. In the present review, an attempt has been made to enlist the important scientifically proven pharmacological applications along with the major phytoconstituents responsible for these activities of *T. divaricata*.

2. Bioactive Components of Plant

T. divaricata, also known as pinwheel flower, is commonly found in small trees throughout South Asia^{10,11}. One of the investigators dug out the workings of the parts of the plant like stems and flowers to root extracts and extracts of leaves of *T. divaricata*. Moreover, the researcher screened 19,20-Dihydroervahanine and 19,20-Dihydrotabernamine along with isolated compounds like tabernaegantine A. It gave out the result that the respective compounds' extracts show slightly higher anti-acetylcholinesterase activity. Moreover, it has been shown in studies that the compounds which were isolated from *T. divaricata* show action against cell lines and the plant also contains many non-alkaloid compounds like enzymes, terpenoids and phenolic acid (Figure 1)¹².

3. Major Alkaloids of *T. divaricata* with Pharmacological Importance

3.1 12-hydroxy akuammicine

12-hydroxy akuammicine is a major alkaloid of *T. divaricata*. One of the investigators administered intravenously. The IP administration in mice and rats of 12-hydroxy akuammicine stopped the growth of ascites and alveolar lymphoma, at a concentration of 15-20 mg/kg/day for 10-20 days¹³.

3.2 19,20-Dihydrotabernamine and 19,20-Dihydroervahanine

The alkaloids mentioned above are generally seen in the roots of *T. divaricata*. Acetylcholinesterase activity can be witnessed in these alkaloids. The inhibitory effect of the alkaloids was proved to be specific and competitive, along with being capable of getting back to its previous position¹⁴.



Figure 1. Nonalkaloid occurring of *T. divaricata* with medicinal importance.

3.3 Apparicine

In vitro studies have shown that polio virus activity can be resisted and brought to a halt by apparicine at a certain concentration^{15,16}. In an *in vitro* study, this alkaloid also had antimicrobial activity against the microbes Salmonella, Pseudomonas, Escherichia, Shigella, Proteus, Staphylococcus and Corynebacterium at different concentrations¹⁷.

3.4 Catharanthine

The study by Ehrlich of ascites tumour cells, where it is easily detected by a biological model for the investigation of tumour cells¹⁸, catharanthine showed the aminoisobutyric acid kind of effects, an amino acid transporter in tumour cells. This revelation inferred that catharanthine might as well have anti-tumour properties¹⁹.

3.5 Conophylline

Conophylline, which comes from *T. divaricata* happens to be a vinca alkaloid. It has been demonstrated in experiments that it induces the differentiation of pancreatic precursor cells²⁰. Conophylline discourages and dissuades the development of cystic structures. Furthermore, they increased the number of insulin-positive cells in the rat pancreatic rudiment of organ culture²⁰. Conophylline has also been demonstrated to help increase insulin production in rat pancreatic acinar carcinoma cells²¹. Conophylline has been used as a health food lately for ameliorating and helping patients avoid obesity and diabetes. Some studies suggest it may lower blood glucose levels²². Conophylline also happens to be a new anti-tumour alkaloid²³⁻²⁶.

3.6 Coronaridine

Coronaridine is an alkaloid found in the parts of the plant of *T. divaricata* such as leaves and stems, along with bark and roots. It has been demonstrated that it influences central nervous and autonomic system activity as well¹³. In the writhing and pain response to

submerging the tail in hot water, one of the researchers²⁷ also demonstrated that coronaridine has both anti-inflammatory and analgesic activities in rats. This can be witnessed in mice as well as in the carrageenan-induced paw oedema method. An intravenous injection (coronaridine) has been shown to cause dose-related hypotension along with bradycardial responses in a regular model of rats^{28,29}.

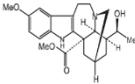
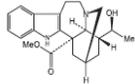
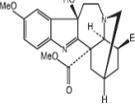
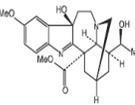
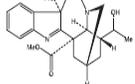
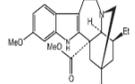
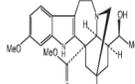
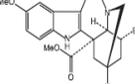
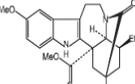
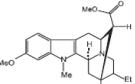
3.7 Dregamine

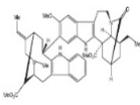
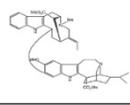
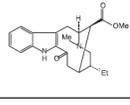
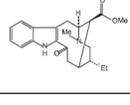
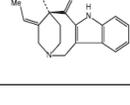
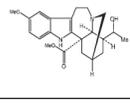
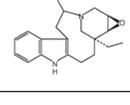
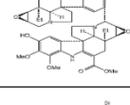
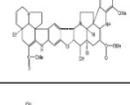
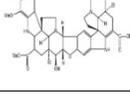
Dregamine can be found in the parts of the plant of *T. divaricata* such as leaves and stems, along with bark and roots. It has been demonstrated that dregamine has convulsive and respiration-related stimulating effects. It causes muscle pain in both *in vivo* and *in vitro* studies, apparently not very different from the activity of ibogaine. Dregamine has been used for the treatment of conditions such as muscular-related and nervous asthenia and respiratory depression³⁰.

3.8 Ibogamine

Ibogamine is an indole alkaloid found in the parts of the plant of *T. divaricata* such as leaves and stems, along with bark and roots (Table 1). In the study, it has been suggested that it can be used to minimise the monosynaptic reflexes of the knee-jerk in some of the animals. Surprisingly, the mechanism of action had no effect on neuromuscular transmission or postsynaptic reflex arcs³¹. However, it is also interesting to note that it possibly had an effect by way of nicotinic receptor stoppage at the meeting point of the neuromuscular system³². Not only that, ibogamine could not also function as a very strong anticonvulsant agent, as it was demonstrated in a mouse model³³. The indole alkaloid ibogamine lately has been explored as a possibility that it can act as an agent that can fight and resist vigorously the drug withdrawal symptoms³⁴⁻³⁶. In rodent models of cocaine and opiate, studies done before the clinical investigations of ibogamine, administration done on its own infers a possibility that it is apparently an agent which is anti-addictive³⁷⁻⁴³.

Table 1. Classification of alkaloids on the basis of plant parts and class of alkaloids¹¹⁵

Parts of plant	Alkaloid	Structure of alkaloid	Class of alkaloids
 Whole Plant	Vocristine		Ibogan
	Heyneanine		
	Voacangine hydroxyindolenine		
	Voacristine hydroxyindolenine		
	19-Heyneanine hydroxyindolenine		
 Flowers	Isovoacangine		Ibogan
	Isovoacristine		
	19-Epivoacangine		
	3-Oxovoacangine		
		11-Methoxy-N-methyl-dihydropericyclivine	

 <p>Stem</p>	Conodusrine		Bis-indole
	Voacamine		
	3S-Cyanocoronaridine		Ibogane
	Dregamine		Corynanthean
	Conolidine		Aspidospermatan
 <p>Leaves</p>	19-Epivoacristine		Ibogane
	5-Hydroxyvoacphylline		Plumeran
	Conofoline		Bis-indole
	Conophyllidine		
	Conophylline		

4. Pharmacological Activities of Plant

4.1 Antioxidant Activity

Antioxidants are molecules or compounds that interrupt the movement of free radicals such as hydrogen peroxide to regulate the process of autoxidation or do the same by directly squeezing their formation^{43,44}. Constituents of therapeutic plants like phenolic diterpenes, carotenoids, volatile oils, flavonoids, phenolic acids and anthocyanidins prove to be and are used as antioxidants⁴⁵. These

compounds choose free radicals as their target by consuming molecules of oxygen, donating molecules of hydrogen, acting as reducing agents, or breaking up antioxidant chains^{44,46}. The various parts of species *T. divaricata* are made of a compound or extract of methanol, ethanol, aqueous, petroleum ether, hexane, octyl benzoate or octyl benzoic acid, chloroform, and digalactosyl deconate. The models used for the activity are fluorescence recovery after photobleaching, 2,2-diphenylpicrylhydrazyl assay - *in vitro*, H₂O₂ free radicals, minimising power - *in vitro*

along with superoxide anion radical scavenging, NO, Trolox equivalent antioxidant capacity assay), H₂O₂ scavenging, and A β 25 - 35 peptides, NOR crystal violet assay and LPO assay⁴⁷⁻⁵⁸.

4.2 Anti-inflammatory Activity

Inflammation is defined as the process by which an organism's body responds to an injury. Inflammation's development is often a result of infection, chemical or physical injury, injury to cells, and death^{59,60}. One of the researchers investigated the anti-inflammatory activity of flower extract obtained from *T. divaricata*. Models were prone to chronic formalin and acute carrageenan⁶¹. The parts such as leaves, stems, flowers, and other parts of the plant are made into compounds with ethyl acetate, aqueous, ethanol, methanol and hexane fraction. The models this activity uses are: minimization of interleukin-6 secretion and TNF production; carrageenan and formalin prompt-mice models. The models of mice along with croton oil bring on oedema in the models of mice^{53,55,61-63}.

4.3 Anti-microbial Activity

Compounds that are complex and do not allow the development of microorganisms at petite assemblage are called antimicrobials⁶⁴. Antimicrobials are often described as secondary metabolites. They are more or less regularly created and extracted either from therapeutic plants or even from microorganisms⁶⁵. One of the researchers has proved *T. divaricata* extracts

work as antibiotics. Monoterpenoid indole alkaloids, like voacamine type and 3-hydroxyiboga, are the compounds that are activated biologically and perform the function of antimicrobial agents, holding back the growth of bacteria, parasites and fungi⁶⁶.

4.4 Antifungal Activity

One of the researchers investigated and reported the antifungal activity of a compound that is pharmacologically active from *T. divaricata*. Coronaridine, a major compound from the plant *T. divaricata*, was found and cleared off from the method - ethanolic extraction of plant *T. divaricata*⁶⁷. The parts flower and leaves of the plant make a compound or extract of ethanol and methanol along with aqueous. The cell lines used for activity are *Penicillium chrysogenum* and *Malassezia furfur*, and the method which is used for antifungal activity is the Poisoned food technique *in vitro*⁶⁸⁻⁷⁰.

4.5 Anticancer Activity

Various leaf extracts in the study were made in a solvent containing methanol and ethyl acetate, along with chloroform and hexane, and were checked against these cell lines: HT-29-Human colorectal adenocarcinoma cell line, 502,713, HCT-15 is used for the colon, Michigan Cancer Foundation-7 (MCF-7) is for breast cancer, and Human prostatic carcinoma cell line (PC-3) is for prostate cancer⁴⁰. Different parts of the plant *T. divaricata* are made into an extract

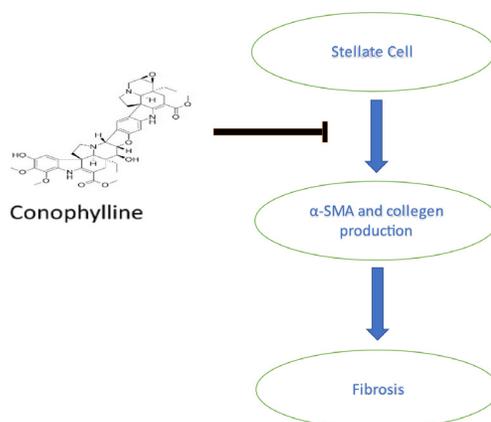


Figure 2. Mechanism of anticancer cancer activity of conophylline.

with different compounds like ethanol, hexane, etc. for anti-cancer activity (Figure 2). Different parts of the plant *T. divaricata* have several compounds that show anti-cancer activity. Root bark of plant contains 5 and 3- oxocoronidine, ibogamine, leaves of plant contains tabernaemontamine, mehranine, vocangine, voaphylline, and the root of plant contains major 2 alkaloid which is conodurine and tabernaegantine. For anticancer activity, various cell lines were used, including HCT 15 (isolated from a cancer patient's large intestine), col-2 and HT-29 for colon cancer, S-10 for sarcoma, V79 and LUP for lung cancer, Human Leukaemia-60, P-388, MOLT-4 (T-cell derived cell line) for blood cancer, and ZR-75-1 and BC- for breast cancer. It has been scientifically proven that conophylline of *T. divaricata* inhibits the stellate cells from α -SMA (α -smooth muscle actin) and collagen production and protects them from fibrosis^{67,71-84}.

4.6 Acetylcholinesterase Activity

One of the investigators, Ingkaninan *et al.*, found the acetylcholinesterase activity of *T. divaricata*'s methanolic extracts. Rats were used in the study as test models and were tested *in vivo*⁸⁵. Almost all parts of the plant show acetylcholinesterase activity, but latex, flowers and roots show high acetylcholinesterase activity. These parts of the plant make an extract with different compounds like methyl alcohol, ethyl alcohol and PBS. There are several compounds present in plants which are responsible for AChE activity, like 19,20 dihydro tabernamine, conodurine, nitrogen-methylofinine, conophylline etc. The method used for the activity is Ellman's method^{71,86-91}.

4.7 Anti-fertility Activity

An anti-fertility activity is shown in the ethanolic extract of *T. divaricata* in oestrogenic activity models in immature female rats. The rats that were used in this study were immature female Albino Wistar. Researchers performed the experiment and found the extract showed the existence of several compounds which have medicinal value. The compounds are carbohydrates, steroids, alkaloids, glycosides, flavonoids, tannins, and increased the uterus' weight tremendously. Some histopathological studies also showed normal architecture of the uterus in vehicle-treated rats. It tells

us about the surface epithelium that has next to no secretory activity⁹².

4.8 Anticonvulsant Activity

The plant is rich in alkaloids which have neurological activities like coronaridine, dregamine, ibogamine and many other alkaloids. Khan and Mukhram did the research for anti-convulsant activity of *T. divaricata* and found that the flowers of the plant have some compounds that are responsible for anti-convulsant activity. Researchers selected 2 models for the activity: the PTZ and MES induced convulsion method and the compounds epivocangine, vobasine, voaphylline and dihydro pericyclivine showed anti-seizure activity in rats and mice by using MES induced convulsions and the PTZ induced convulsions method. Researchers found that the methanolic extract of flowers can stop seizures in animals. It is due to the inhibition of sodium channels, but the exact mechanism is not known⁹³⁻¹⁰¹.

4.9 Antibacterial Activity

There has been major research going on for almost half a century to advance antibacterial medicines¹⁰². Roughly a quarter of the medicine in our time has plant-related compounds at its base, which is very significant¹⁰³. *T. divaricata* indole alkaloids exhibit a wide range of pharmacological activities, including antibacterial activity against both gram-positive and gram-negative bacteria¹⁰⁴. The different parts of the plant like bark, flowers, roots are made into an extract with different compounds like chloroform, ethanol, dether, methanol and major natural compounds which are present in the plant like alkaloids, dichloromethane, taberdivamines A and B were used for their anti-bacterial activity. The cell line used for the activity are *Staphylococcus aureus*, *Staphylococcus saprophyticus*, *Streptococcus agalctiae*, *Streptococcus pyogenes*, *Enterococcus faecalis*, *Salmonella typhi*, *Escherichia coli*, *Shigella boydii*, *Shigella dysenteriae*, *Pseudomonas aeruginosa*, *Bacillus cereus*, *Klebsiella sp.*, *Streptococcus uberis*, *Escherichia coli* (ATCC 25922), *Klebsiella pneumoniae* (ATCC 35657), *Salmonella typhimurium* (MTCC 441), *Shigella flexneri* (ATCC 29508), *Staphylococcus aureus* (ATCC25923), *Aeromonas hydrophila*, *Staphylococcus epidermidis*, *Gardnerella vaginalis*, *Streptococcus agalactiae*, *Propionibacterium acnes*, *Corynebacterium*

macbinleyi, *Bacillus subtilis*, *Enterococcus faecalis*, *Bacillus megaterium*, *Proteus mirabilis*, *Shigella flexneri* (BCH 995), *Shigella boydii* (8) which is a gram negative bacteria, *Shigella sonnei* (NK 840), *Shigella dysenteriae* (1), *Vibrio cholerae* (1023), *Vibrio cholerae* (575), *Vibrio cholerae* (1311), *Vibrio cholerae* (756), *Escherichia coli* (RH 07/12, 18/9, K88), *Streptococcus suis* (gram positive bacteria), *Salmonella species*, *Cornebacterium diphtheriae* (AP596), *Staphylococcus aureus* (ML 267), *Staphylococcus aureus* (MTCC 96, ATCC 6538), *Bacillus subtilis* (MTCC 441), *Bacillus pumilis* (8241) *Pseudomonas aeruginosa* (AP585 NLF), *Klebsiella pneumoniae* strains, *Salmonella Paratyphi*, *Lactobacillus*, *Proteus vulgaris*, and *Klebsiella aerogenes*^{67,94,105-114}.

4.10 Anthelmintic Activity

Helminths are commonly found infectious agents of humans in the developing world. The disease caused by them creates such chaos across the world that it is becoming difficult for countries to cope with it¹¹⁵. *T. divaricata* is one of the Indian medicinal plants that shows anti-helminthic activity¹¹⁶. The anti-helminthic activity was performed on a mature earth worm found in India called *Pheretima posthuman*. It was done so because it has a great resemblance not only anatomically but also physiologically with the human being's intestinal round worm parasites¹¹⁷. Chloroform and methanolic extracts of *T. divaricata* leaves were prepared, Radhika B and Vilasini S performed the experiment and observed anthelmintic activity of methanol extract is more potent compared to the chloroform extract¹¹⁵.

4.11 Gastrointestinal Effect

Khan *et al.* found the gastro and intestinal effects of extract with methanol of *T. divaricata* flowers. To evaluate the possible effects in the study, the model of a rat pyloric ligation method (gastric ulceration induced) and as a standard drug one of the proton-pump inhibitors was used. The study resulted in the discovery that the extract reduced the gastric juice amount, free and total acidities, and ulcer index, along with the pH of gastric acid produced¹¹⁸. Khan *et al.* also did one more test using a range of concentrations of the methanol extract from the *T. divaricata* flower. After measuring, parameters such as catalase and superoxide dismutase,

along with the mucin and total protein, when treated with extracts, displayed an index that declined¹¹⁹.

4.12 Anti-diabetic Activity

The activity of *T. divaricata* (antidiabetic) from the extract (methanolic) was applied to alloxan-prompted diabetic rats. The results that came out showed a great amount of antidiabetic activity. Moreover, an additional decrease can also be seen in the oxidative damage effect that can be seen in rats¹²⁰. Kanthlal *et al.* suggest in their study that the insulin receptors may be alerted by an extract (methanolic). Henceforth, it may result in the production of beta-stem cells in the subject of the test's pancreas. Antidiabetic activity can be seen by the compound conophylline, which is generally segregated from *T. divaricata*^{120,121}. Increased plasma levels in diabetic rats were observed in the study. A severe decline in blood glucose levels can also be seen in the results, which indicates antidiabetic activity as well¹²².

5. Conclusion

T. divaricata is easily and plentifully available in India as it is easily grown in houses, gardens, or even by the roadside. This plant has been used as a medicinal plant for many years. The present review revealed its scientifically proven anticancer, antioxidant, anti-fungal, anti-bacterial activities, etc. The present review concludes an assessment of the compound within the plant *T. divaricata* which shows pharmacological activities. Numerous compounds, such as conophylline, cornaridine, vocamine, vincristine etc., are often utilised for their curative effects. There are several alkaloids present in the plant that show different pharmacological activities like 19, 20-dihydrotabernamine and 19,20-dihydroervahanine A., which are found in the roots of the plant, show acetylcholinesterase activity, apparicine shows antimicrobial activity, catharanthine and conophylline show anti-cancer activity, anti-inflammatory and analgesic activity due to coronaridine found in almost all parts of the plant. This plant can also be useful for the treatment of Alzheimer's disease. The alkaloids like vocangine, coronaridine and isovacristine are observed to be present in *T. divaricata* and have parasymphomimetic activity. The review also enlisted different chemical compounds within different parts

of *T. divaricata* that are biologically active. It has a lot more room for further biological evolution activities in the future based on the details presented in this review. The easy and plentiful availability of the plant since ancient times needs further research to magnify insights about the biologically active compounds and relative pharmacological activities of this plant for socioeconomic benefits.

6. Acknowledgment

The authors of the present review acknowledge their gratitude to the Director, Noida Institute of Engineering and Technology (Pharmacy Institute) for his enormous support and for providing all kinds of facilities to get information on *T. divaricata*.

7. References

- Raj CN, Balasubramaniam A. Pharmacogostic and antimicrobial studies of the leaves of *Tabernaemontana divaricata* R. br. *Pharmacologyonline*. 2011; 2:1171-7.
- Pushpa B, Latha KP, Vaidya VP, Shruthi A, Shweatha A. Phytochemical analysis and antimicrobial evaluation of leaves extract of *Tabernaemontana coronaria*. *J Chem Pharm Res*. 2012; 4:3731-3.
- Gopinath SM, Suneetha TB, Mruganka VD, Ananda S. Evaluation of antibacterial activity of *Tabernaemontana divaricata* (L.) leaves against the causative organisms of bovine mastitis. *International Journal of Research in Phytochemistry and Pharmacology*. 2011; 1(4):211-3.
- Poornima K, Krishnan R, Aswathi KV, Gopalakrishnan VK. Toxicological evaluation of ethanolic extract of *Tabernaemontana coronaria* (L) R. Br. *Asian Pacific Journal of Tropical Disease*. 2012; 2:S679-84. [https://doi.org/10.1016/S2222-1808\(12\)60243-6](https://doi.org/10.1016/S2222-1808(12)60243-6)
- Konno K. Plant latex and other exudates as plant defense systems: roles of various defense chemicals and proteins contained therein. *Phytochemistry*. 2011; 72(13):1510-30. <https://doi.org/10.1016/j.phytochem.2011.02.016>
- Agrawal AA, Konno K. Latex: A model for understanding mechanisms, ecology, and evolution of plant defense against herbivory. *Annu. Rev. Ecol. Evol. Syst*. 2009; 40:311-31. <https://doi.org/10.1146/annurev.ecolsys.110308.120307>
- Ramos MV, Souza DP, Gomes MT, Freitas CD, Carvalho CP, Junior PA, Salas CE. A phytopathogenic cysteine peptidase from latex of wild rubber vine *Cryptostegia grandiflora*. *The Protein Journal*. 2014; 33(2):199-209. <https://doi.org/10.1007/s10930-014-9551-4>
- Freitas CD, Silva MZ, Bruno-Moreno F, Monteiro-Moreira AC, Moreira RA, Ramos MV. New constitutive latex osmotin-like proteins lacking antifungal activity. *Plant Physiology and Biochemistry*. 2015; 96:45-52. <https://doi.org/10.1016/j.plaphy.2015.07.012>
- Yagami T, Sato M, Nakamura A, Komiyama T, Kitagawa K, Akasawa A, Ikezawa Z. Plant defense-related enzymes as latex antigens. *Journal of allergy and clinical immunology*. 1998; 101(3):379-85. [https://doi.org/10.1016/S0091-6749\(98\)70251-9](https://doi.org/10.1016/S0091-6749(98)70251-9)
- Saha S. Phytochemical screening and Pharmacological Evaluation of the Methanol Extract of *Tabernaemontana divaricata* leaves.
- Tabernaemontana divaricata*. Natural Resource Conservation Service PLANTS Database USDA. Retrieved 7 December 2015.
- Lakhey P and Pathak J. 2020. *Tabernaemontana divaricata*. The IUCN Red List of Threatened Species 2020.
- Boligon AA, Piana M, Kubiça TF, Mario DN, Dalmolin TV, Bonez PC, Weiblen R, Lovato L, Alves SH, Campos MM, Athayde ML. HPLC analysis and antimicrobial, antimycobacterial and antiviral activities of *Tabernaemontana catharinensis* A. DC. *Journal of Applied Biomedicine*. 2015; 13(1):7-18. <https://doi.org/10.1016/j.jab.2014.01.004>
- Silveira DA, de Melo AF, Magalhaes PO, Fonseca-Bazzo YM. *Tabernaemontana* species: Promising sources of new useful drugs. In *Studies in Natural Products Chemistry*. 2017; 54:227-289. Elsevier. <https://doi.org/10.1016/B978-0-444-63929-5.00007-3>
- Andrade MT, Lima JA, Pinto AC, Rezende CM, Carvalho MP, Epifanio RA. Indole alkaloids from *Tabernaemontana australis* (Muell. Arg) Miens that inhibit acetylcholinesterase enzyme. *Bioorganic and Medicinal Chemistry*. 2005; 13(12):4092-5. <https://doi.org/10.1016/j.bmc.2005.03.045>
- Farnsworth NR, Svoboda GH, Blomster RN. Antiviral activity of selected *Catharanthus* alkaloids. *Journal of Pharmaceutical Sciences*. 1968; 57(12):2174-5. <https://doi.org/10.1002/jps.2600571235>
- Arens H, Borbe HO, Ulbrich B, Stockigt J. Detection of pericine, a new CNS-active indole alkaloid from *Picalima nitida* cell suspension culture by opiate receptor binding studies. *Planta Medica*. 1982; 46(12):210-4. <https://doi.org/10.1055/s-2007-971216>
- Siems WG, Grune T, Beierl B, Zollner H, Esterbauer H. The metabolism of 4-hydroxynonenal, a lipid peroxidation product, is dependent on tumor age in Ehrlich mouse ascites cells. *Free Radicals and Aging*. 1992; 124-35. https://doi.org/10.1007/978-3-0348-7460-1_13
- Fyfe MJ, Lofffield S, Goldman ID. A reduction in energy-dependent amino acid transport by microtubular inhibitors in Ehrlich ascites tumor cells. *Journal of Cellular Physiology*. 1975; 86(2):201-11. <https://doi.org/10.1002/jcp.1040860203>
- Ogata T, Li L, Yamada S, Yamamoto Y, Tanaka Y, Takei I, Umezawa K, Kojima I. Promotion of β -cell differen-

- tiation by conophylline in fetal and neonatal rat pancreas. *Diabetes*. 2004; 53(10):2596-602. <https://doi.org/10.2337/diabetes.53.10.2596>
21. Takatsuna H, Umezawa K. Screening of bioactive metabolites for pancreatic regeneration chemotherapy. *Biomedicine and Pharmacotherapy*. 2004; 58(10):610-3. <https://doi.org/10.1016/j.biopha.2004.10.003>
 22. Kojima I, Umezawa K. Conophylline: A novel differentiation inducer for pancreatic β cells. *The international journal of biochemistry and cell biology*. 2006; 38(5-6):923-30. <https://doi.org/10.1016/j.biocel.2005.09.019>
 23. Atsumi S, Nagasawa A, Koyano T, Kowithayakornd T, Umezawa K. Suppression of TGF- β signaling by conophylline via upregulation of c-Jun expression. *Cellular and Molecular Life Sciences CMLS*. 2003; 60(11):2516-25. <https://doi.org/10.1007/s00018-003-3299-x>
 24. Gohda J, Inoue JI, Umezawa K. Down-regulation of TNF- α receptors by conophylline in human T-cell leukemia cells. *International Journal of Oncology*. 2003; 23(5):1373-9. <https://doi.org/10.3892/ijo.23.5.1373>
 25. Irie T, Kubushiro K, Suzuki K, Tsukazaki K, Umezawa K, Nozawa S. Inhibition of attachment and chemotactic invasion of uterine endometrial cancer cells by a new vinca alkaloid, conophylline. *Anticancer Research*. 1999; 19(4B):3061-6.
 26. Umezawa K, Taniguchi T, Toi M, Ohse T, Tsutsumi N, Yamamoto T, Koyano T, Ishizuka M. Growth inhibition of K-ras-expressing tumours by a new vinca alkaloid, conophylline, in nude mice. *Drugs under Experimental and Clinical Research*. 1996; 22(2):35-40.
 27. Taesotikul T, Panthong A, Kanjanapothi D, Verpoorte R, Scheffer JJ. Anti-inflammatory, antipyretic and antinociceptive activities of *Tabernaemontana pandacaqui* Poir. *Journal of Ethnopharmacology*. 2003; 84(1):31-5. [https://doi.org/10.1016/S0378-8741\(02\)00264-7](https://doi.org/10.1016/S0378-8741(02)00264-7)
 28. Taesotikul T, Panthong A, Kanjanapothi D, Verpoorte R, Scheffer JJ. Cardiovascular effects of *Tabernaemontana pandacaqui*. *Journal of Ethnopharmacology*. 1989; 27(1-2):107-19. [https://doi.org/10.1016/0378-8741\(89\)90083-4](https://doi.org/10.1016/0378-8741(89)90083-4)
 29. Taesotikul T, Panthong A, Kanjanapothi D, Verpoorte R, Scheffer JJ. Cardiovascular activity of the crude alkaloidal fraction from *Tabernaemontana pandacaqui* in the rat. *Journal of Ethnopharmacology*. 1998; 59(3):131-7. [https://doi.org/10.1016/S0378-8741\(97\)00116-5](https://doi.org/10.1016/S0378-8741(97)00116-5)
 30. Mehrotra PK, Kamboj VP. Hormonal Profile of Coronaridine Hydrochloride-an Antifertility Agent of Plant Origin1. *Planta Medica*. 1978; 33(04):345-9. <https://doi.org/10.1055/s-0028-1097389>
 31. Schneider JA, Sigg EB. Neuropharmacological studies on ibogaine, an indole alkaloid with central-stimulant properties. *Annals of the New York Academy of Sciences*. 1957; 66(3):765-76. <https://doi.org/10.1111/j.1749-6632.1957.tb40765.x>
 32. Pace CJ, Glick SD, Maisonneuve IM, He LW, Jokiel PA, Kuehne ME, Fleck MW. Novel iboga alkaloid congeners block nicotinic receptors and reduce drug self-administration. *European Journal of Pharmacology*. 2004; 492(2-3):159-67. <https://doi.org/10.1016/j.ejphar.2004.03.062>
 33. Chen G and B Bohner. A study of central nervous system stimulants. *The Journal of Pharmacology and Experimental Therapeutics*. 1958; 123(3):212-5.
 34. Goutarel R, Gollnhofer O, Sillans R. Pharmacodynamics and therapeutic applications of iboga and ibogaine. *Psychedelic Monographs and Essays*. 1993; 6:71-111.
 35. Lotsof HS, inventor; Lotsof Howard S. Rapid method for interrupting the narcotic addiction syndrome. United States Patent US. 1985; 4499096.
 36. Prachayasakul W, Pongchaidecha A, Chattipakorn N, Chattipakorn S. Ethnobotany and ethnopharmacology of *Tabernaemontana divaricata*. *Indian Journal of Medical Research*. 2008; 127(4):317-36.
 37. Aceto MD, Bowman ER, Harris LS, May EL. Dependence studies of new compounds in the rhesus monkey and mouse (1991). NIDA Research Monograph. 1992; 119:513-58.
 38. Cappendijk SL, Dzoljic MR. Inhibitory effects of ibogaine on cocaine self-administration in rats. *European Journal of Pharmacology*. 1993; 241(2-3):261-5. [https://doi.org/10.1016/0014-2999\(93\)90212-Z](https://doi.org/10.1016/0014-2999(93)90212-Z)
 39. Glick SD, Rossman K, Rao NC, Maisonneuve IM, Carlson JN. Effects of ibogaine on acute signs of morphine withdrawal in rats: Independence from tremor. *Neuropharmacology*. 1992; 31(5):497-500. [https://doi.org/10.1016/0028-3908\(92\)90089-8](https://doi.org/10.1016/0028-3908(92)90089-8)
 40. Glick SD, Rossman K, Steindorf S, Maisonneuve IM, Carlson JN. Effects and aftereffects of ibogaine on morphine self-administration in rats. *European Journal of Pharmacology*. 1991; 195(3):341-5. [https://doi.org/10.1016/0014-2999\(91\)90474-5](https://doi.org/10.1016/0014-2999(91)90474-5)
 41. Glick SD, Maisonneuve IM. Mechanisms of antiaddictive actions of ibogaine a. *Annals of the New York Academy of Sciences*. 1998; 844(1):214-26. <https://doi.org/10.1111/j.1749-6632.1998.tb08237.x>
 42. Sershen H, Hashim A, Lajtha A. Ibogaine reduces preference for cocaine consumption in C57BL/6By mice. *Pharmacology Biochemistry and Behavior*. 1994; 47(1):13-9. [https://doi.org/10.1016/0091-3057\(94\)90105-8](https://doi.org/10.1016/0091-3057(94)90105-8)
 43. Sershen H, Hashim A, Lajtha A. Ibogaine and cocaine abuse: pharmacological interactions at dopamine and serotonin receptors. *Brain research bulletin*. 1997; 42(3):161-8. [https://doi.org/10.1016/S0361-9230\(96\)00296-1](https://doi.org/10.1016/S0361-9230(96)00296-1)
 44. Fierascu RC, Ortan A, Fierascu IC, Fierascu I. In vitro and in vivo evaluation of antioxidant properties of wild-growing

- plants. A short review. *Current Opinion in Food Science*. 2018; 24:1-8. <https://doi.org/10.1016/j.cofs.2018.08.006>
45. Toghueo RM, Boyom FF. Endophytes from ethno-pharmacological plants: Sources of novel antioxidants-A systematic review. *Biocatalysis and Agricultural Biotechnology*. 2019; 22:101430. <https://doi.org/10.1016/j.bcab.2019.101430>
46. Shori AB. Screening of antidiabetic and antioxidant activities of medicinal plants. *Journal of Integrative Medicine*. 2015; 13(5):297-305. [https://doi.org/10.1016/S2095-4964\(15\)60193-5](https://doi.org/10.1016/S2095-4964(15)60193-5)
47. Jain S, Jain A, Jain N, Jain DK, Balekar N. Phytochemical investigation and evaluation of in vitro free radical scavenging activity of *Tabernaemontana divaricata* Linn. *Natural Product Research*. 2010; 24(3):300-4. <https://doi.org/10.1080/14786410903237123>
48. Wasupongpun W, Premkaisorn P. Evaluation of Antioxidant Activity of Eleven Thai Medicinal Herbs. *Sci. J*. 2010; 26:29-38.
49. Rumzhum NN, Rahman MM, Kazal MK. Antioxidant and cytotoxic potential of methanol extract of *Tabernaemontana divaricata* leaves. *International Current Pharmaceutical Journal*. 2012; 1(2):27-31. <https://doi.org/10.3329/icpj.v1i2.9446>
50. Venkatachalapathi S, Saranya C, Ravi S. Isolation and Characterization of Bio Active Compounds from *Tabernaemontana divaricata* and a Study of its Antioxidant and Antibacterial Activity. *Indo Am. J. Pharm. Res*. 2014; 4(5):2401-6.
51. Khan MS. Gastroprotective effect of *Tabernaemontana divaricata* (Linn.) R. Br. Flower methanolic extract in wistar rats. *British Journal of Pharmaceutical Research*. 2011; 1(3):88. <https://doi.org/10.9734/BJPR/2011/347>
52. Choudhary RK, Saroha AE, Swarnkar PL. Screening of endogenous antioxidants in some medicinal plants. *Toxicological and Environmental Chemistry*. 2011; 93(4):656-64. <https://doi.org/10.1080/02772248.2010.551122>
53. Mueller M, Janneon K, Puttipan R, Unger FM, Viernstein H, Okonogi S. Anti-inflammatory, antibacterial, and antioxidant activities of Thai medicinal plants. *Int. J. Pharm. Pharm. Sci*. 2015; 7(11):123-8.
54. Anbukkarasi M, Thomas PA, Sheu JR, Geraldine P. In vitro antioxidant and anticataractogenic potential of silver nanoparticles biosynthesized using an ethanolic extract of *Tabernaemontana divaricata* leaves. *Biomedicine and Pharmacotherapy*. 2017; 91:467-75. <https://doi.org/10.1016/j.biopha.2017.04.079>
55. Anbukkarasi M, Sundararajan M, Venkadeswaran K, Ruban VV, Anand T, Geraldine P. Antihypercholesterolemic, anti-oxidative and anti-inflammatory potential of an extract of the plant *Tabernaemontana divaricata* in experimental rats fed an atherogenic diet. *Biocatalysis and Agricultural Biotechnology*. 2019; 19:101115. <https://doi.org/10.1016/j.bcab.2019.101115>
56. Kalaimagal C. In vitro antioxidant activity in ethanolic leaf extract of (L.). *International Journal of Bio-Pharma Research*. 2019; 8(6):2602-6.
57. Santhi R, Annapurani S. Preliminary evaluation of In vitro and In vivo antioxidative and antitumor activities of flavonoid extract of *Tabernaemontana divaricata* leaves in Ehrlich's lymphoma and Dalton's lymphoma ascites model. *Journal of Cancer Research and Therapeutics*. 2020; 16(1):78. https://doi.org/10.4103/jcrt.JCRT_445_17
58. Khongsombat O. Inhibitory effects of *Tabernaemontana divaricata* root extract on oxidative stress and neuronal loss induced by amyloid β 25-35 peptide in mice. *Journal of Traditional and Complementary Medicine*. 2018; 8(1):184-9. <https://doi.org/10.1016/j.jtcme.2017.05.009>
59. Elgorashi EE, McGaw LJ. African plants with in vitro anti-inflammatory activities: A review. *South African Journal of Botany*. 2019; 126:142-69. <https://doi.org/10.1016/j.sajb.2019.06.034>
60. Calixto JB, Otuki MF, Santos AR. Anti-inflammatory compounds of plant origin. Part I. Action on arachidonic acid pathway, nitric oxide and nuclear factor κ B (NF- κ B). *Planta Medica*. 2003; 69(11):973-83. <https://doi.org/10.1055/s-2003-45141>
61. Jain S, Sharma P, Ghule S, Jain A, Jain N. In vivo anti-inflammatory activity of *Tabernaemontana divaricata* leaf extract on male albino mice. *Chinese Journal of Natural Medicines*. 2013; 11(5):472-6. [https://doi.org/10.1016/S1875-5364\(13\)60086-2](https://doi.org/10.1016/S1875-5364(13)60086-2)
62. Jolly C, Thambi P, Kuzhivelil B, Sabu M. Antioxidant and anti-inflammatory activities of the flowers of *Tabernaemontana coronaria* (L) R. Br. *Indian J. Pharm. Sci*. 2006; 68:352. <https://doi.org/10.4103/0250-474X.26675>
63. Kanthlal SK, Suresh V, Arunachalam G, Frank PR, Kameshwaran S. In vivo evaluation of analgesic and antipyretic activity of aerial parts of *Tabernaemontana divaricata* in experimental animal models. *Pharmacologyonline*. 2011; 3:1127-33.
64. Bhadane BS, Patil MP, Maheshwari VL, Patil RH. Ethnopharmacology, phytochemistry, and biotechnological advances of family Apocynaceae: A review. *Phytotherapy Research*. 2018; 32(7):1181-210. <https://doi.org/10.1002/ptr.6066>
65. Ncube NS, Afolayan AJ, Okoh AI. Assessment techniques of antimicrobial properties of natural compounds of plant origin: current methods and future trends. *African Journal of Biotechnology*. 2008; 7(12). <https://doi.org/10.5897/AJB07.613>
66. Marinho FF, Simoes AO, Barcellos T, Moura S. Brazilian *Tabernaemontana* genus: Indole alkaloids and phytochemi-

- cal activities. *Fitoterapia*. 2016; 114:127-37. <https://doi.org/10.1016/j.fitote.2016.09.002>
67. Singh B, A Sharma R, K Vyas G. Antimicrobial, anti-neoplastic and cytotoxic activities of indole alkaloids from *Tabernaemontana divaricata* (L.) R. Br. *Current Pharmaceutical Analysis*. 2011; 7(2):125-32. <https://doi.org/10.2174/157341211795684844>
 68. Kumari S, Mazumder A, Bhattacharya S. Pharmacognostical and antimicrobial studies of the stem of *Tabernaemontana divaricata* Linn. *Int. J. Pharm. Sci.* 2015; 7:101-4.
 69. Rakkimuthu R, Nithiyakamatchi R, Sathishkumar P, Ananda Kumar AM, Sowmiya D. In vitro antifungal activity of formulated floral extracts against *Malassezia furfur*. *Int. J. Anal. Exp. Modal Anal.* 2019; 6:1-0.
 70. Satapathy R, Beura S. Management of *Colletotrichum gloeosporioides* (Penz.) Causing Cashew Anthracnose through Botanicals. *Int. J. Curr. Microbiol. Appl. Sci.* 2018; 7:3539-43. <https://doi.org/10.20546/ijcmas.2018.709.439>
 71. Thind TS, Agrawal SK, Saxena AK, Arora S. Studies on cytotoxic, hydroxyl radical scavenging and topoisomerase inhibitory activities of extracts of *Tabernaemontana divaricata* (L.) R. Br. ex Roem. and Schult. *Food and Chemical Toxicology*. 2008; 46(8):2922-7. <https://doi.org/10.1016/j.fct.2008.05.036>
 72. Rumzhum NN, Rahman MM, Kazal MK. Antioxidant and cytotoxic potential of methanol extract of *Tabernaemontana divaricata* leaves. *International Current Pharmaceutical Journal*. 2012; 1(2):27-31. <https://doi.org/10.3329/icpj.v1i2.9446>
 73. Thombre, R.Jagtap, R.Patil, N. Evaluation of phytoconstituents, antibacterial, antioxidant and cytotoxic activity of *Vitex negundo* L. and *Tabernaemontana divaricata* L. *International Journal of Pharma and Biological Sciences*. 2013; 4:389-96.
 74. Lee CC, Houghton P. Cytotoxicity of plants from Malaysia and Thailand used traditionally to treat cancer. *Journal of Ethnopharmacology*. 2005; 100(3):237-43. <https://doi.org/10.1016/j.jep.2005.01.064>
 75. Kumar A, Selvakumar S. Antiproliferative efficacy of *Tabernaemontana divaricata* against HEP2 cell line and Vero cell line. *Pharmacognosy Magazine*. 2015; 11(Suppl 1):S46. <https://doi.org/10.4103/0973-1296.157682>
 76. Hullatti K, Pathade N, Mandavkar Y, Godavarthi A, Biradi M. Bioactivity-guided isolation of cytotoxic constituents from three medicinal plants. *Pharmaceutical Biology*. 2013; 51(5):601-6. <https://doi.org/10.3109/13880209.2012.753919>
 77. Poornima K, Gopalakrishnan VK. Anticancer activity of *Tabernaemontana coronaria* against carcinogen induced clear cell renal cell carcinoma. *Chinese Journal of Biology*. 2014; 2014. <https://doi.org/10.1155/2014/584074>
 78. Bao MF, Yan JM, Cheng GG, Li XY, Liu YP, Li Y, Cai XH, Luo XD. Cytotoxic indole alkaloids from *Tabernaemontana divaricata*. *Journal of Natural Products*. 2013; 76(8):1406-12. <https://doi.org/10.1021/np400130y>
 79. Guo LL, He HP, Di YT, Li SF, Cheng YY, Yang W, Li Y, Yu JP, Zhang Y, Hao XJ. Indole alkaloids from *Ervatamia chinensis*. *Phytochemistry*. 2012; 74:140-5. <https://doi.org/10.1016/j.phytochem.2011.11.002>
 80. Gunasekera SP, Cordell G, Farnsworth NR. Anticancer indole alkaloids of *Ervatamia heyneana*. *Phytochemistry*. 1980; 19(6):1213-8. [https://doi.org/10.1016/0031-9422\(80\)83086-X](https://doi.org/10.1016/0031-9422(80)83086-X)
 81. Ohishi K, Toume K, Arai MA, Sadhu SK, Ahmed F and Ishibashi M. Coronaridine, an iboga type alkaloid from *Tabernaemontana divaricata*, inhibits the Wnt signaling pathway by decreasing β -catenin mRNA expression. *Bioorganic and Medicinal Chemistry Letters*. 2015; 25(18):3937-3940. <https://doi.org/10.1016/j.bmcl.2015.07.036>
 82. Mavuduru S, Kriti K, Mishra A, Ghosh M. Isolation of Anticancer Agents from *Tabernaemontana divaricata* (L.) R. Br. ex Roem. and Schult.
 83. Dantu AS, Shankarguru P, Ramya DD, Vedha HB. Evaluation of in vitro anticancer activity of hydroalcoholic extract of *Tabernaemontana divaricata*. *Asian J Pharm Clin Res*. 2012; 5(3):59-61.
 84. Doshi GM, Kanad PP, Azad N, Desai A, Somani RR, Chaskar PK. In vitro Cytotoxicity Studies on *Tabernaemontana divaricata* leaves extracts by sulforhodamine B assay method. *Int. J. Pharm. Sci. Rev. Res*. 2017; 45:179-82.
 85. Kam TS, Pang HS, Lim TM. Biologically active indole and bisindole alkaloids from *Tabernaemontana divaricata*. *Organic and Biomolecular Chemistry*. 2003; 1(8):1292-7. <https://doi.org/10.1039/b301167d>
 86. Ingkaninan K, Changwijit K, Suwanborirux K. Vobasinyliboga bisindole alkaloids, potent acetylcholinesterase inhibitors from *Tabernaemontana divaricata* root. *Journal of Pharmacy and Pharmacology*. 2006; 58(6):847-52. <https://doi.org/10.1211/jpp.58.6.0015>
 87. Chaiyana W, Schripsema J, Ingkaninan K, Okonogi S. 3'-R/S-Hydroxyvoacamine, a potent acetylcholinesterase inhibitor from *Tabernaemontana divaricata*. *Phytomedicine*. 2013; 20(6):543-8. <https://doi.org/10.1016/j.phymed.2012.12.016>
 88. Chattipakorn S, Pongpanparadorn A, Pratchayasakul W, Pongchaidacha A, Ingkaninan K, Chattipakorn N. *Tabernaemontana divaricata* extract inhibits neuronal acetylcholinesterase activity in rats. *Journal of Ethnopharmacology*. 2007; 110(1):61-8. <https://doi.org/10.1016/j.jep.2006.09.007>
 89. Ingkaninan K, Temkitthawon P, Chuenchom K, Yuyaem T, Thongnoi W. Screening for acetylcholinesterase inhibitory activity in plants used in Thai traditional rejuvenating and

- neurotonic remedies. *Journal of Ethnopharmacology*. 2003; 89(2-3):261-4. <https://doi.org/10.1016/j.jep.2003.08.008>
90. Nakdook W, Khongsombat O, Taepavarapruk P, Taepavarapruk N, Ingkaninan K. The effects of *Tabernaemontana divaricata* root extract on amyloid β -peptide 25-35 peptides induced cognitive deficits in mice. *Journal of Ethnopharmacology*. 2010; 130(1):122-6. <https://doi.org/10.1016/j.jep.2010.04.027>
91. Singh MK, Usha R, Hithayshree KR, Bindhu OS. Hemostatic potential of latex proteases from *Tabernaemontana divaricata* (L.) R. Br. ex. Roem. and Schult. and *Artocarpus altilis* (Parkinson ex. FA Zorn) Forsberg. *Journal of Thrombosis and Thrombolysis*. 2015; 39(1):43-9. <https://doi.org/10.1007/s11239-013-1012-y>
92. Jain S, Jain A, Deb L, Dutt KR, Jain DK. Evaluation of antifertility activity of *Tabernaemontana divaricata* (Linn) R. Br. leaves in rats. *Natural Product Research*. 2010; 24(9):855-60. <https://doi.org/10.1080/14786410903314385>
93. Gomez Gonzalez C, Rodriguez C, Janetina S. Hecubine: Two novel alkaloids. *Revista Cubana de Farmacia*. 1978; 12:177-83.
94. Arambewela LS, Ranatunge T. Indole alkaloids from *Tabernaemontana divaricata*. *Phytochemistry*. 1991; 30(5):1740-1. [https://doi.org/10.1016/0031-9422\(91\)84254-P](https://doi.org/10.1016/0031-9422(91)84254-P)
95. Pawelka KH, Stockigt J. Indole alkaloids from cell suspension cultures of *Tabernaemontana divaricata* and *Tabernaemontana iboga*. *Plant Cell Reports*. 1983; 2(2):105-7. <https://doi.org/10.1007/BF00270178>
96. Gorman M, Neuss N, Cone NJ, Deyrup JA. Alkaloids from Apocynaceae. III. 1 Alkaloids of *Tabernaemontana* and *Ervatamia*. The Structure of Coronaridine, A New Alkaloid Related to Ibogamine. *Journal of the American Chemical Society*. 1960; 82(5):1142-5. <https://doi.org/10.1021/ja01490a031>
97. Elkeiy M. Separation, Isolation and Identification of Certain. *Journal of Pharmaceutical Sciences of the United Arab Republic*. 1966; 7.
98. Gomez-Gonzalez C, Polo CN, Rodriguez SC, Mendez AP. Phytochemistry of *Ervatamia coronaria* Stapf (IV). Fractionation of the total bases present in the flowers in an acidity gradient. *Cuban Magazine of Pharmacy*. 1981; 15(3):192-9.
99. Gomez Gonzalez C, Martinez J. Phytochemistry of *Ervatamia coronaria* Stapf.(II). Hecubine and voaphylline: Two alkaloids present in leaves. *Revista Cubana de Farmacia*. 1976; 10:45-54.
100. Raj K, Shoeb A, Kapil RS, Popli SP. Alkaloids of *Tabernaemontana divaricata*. *Phytochemistry*. 1974; 13(8):1621-2. [https://doi.org/10.1016/0031-9422\(74\)80344-4](https://doi.org/10.1016/0031-9422(74)80344-4)
101. Pathak S, Wanjari MM, Jain SK, Tripathi M. Evaluation of antiseizure activity of essential oil from roots of *Angelica archangelica* Linn. in mice. *Indian Journal of Pharmaceutical Sciences*. 2010; 72(3):371. <https://doi.org/10.4103/0250-474X.70487>
102. Zorofchian Moghadamtousi S, Abdul Kadir H, Hassandarvish P, Tajik H, Abubakar S, Zandi K. A review on antibacterial, antiviral, and antifungal activity of curcumin. *BioMed Research International*. 2014; 2014. <https://doi.org/10.1155/2014/186864>
103. Malla B, Gauchan DP, Chhetri RB. An ethnobotanical study of medicinal plants used by ethnic people in Parbat district of western Nepal. *Journal of Ethnopharmacology*. 2015; 165:103-17. <https://doi.org/10.1016/j.jep.2014.12.057>
104. Medeiros MR, de Melo Prado LA, Fernandes VC, Figueiredo SS, Coppede J, Martins J, Fiori GM, Martinez-Rossi NM, Belebony RO, Contini SH, Pereira PS. Antimicrobial activities of indole alkaloids from *Tabernaemontana catharinensis*. *Natural Product Communications*. 2011; 6(2):1934578X1100600209. <https://doi.org/10.1177/1934578X1100600209>
105. Kumari S, Mazumder A, Bhattacharya S. Pharmacognostical and antimicrobial studies of the stem of *Tabernaemontana divaricata* Linn. *Int. J. Pharm. Sci*. 2015; 7:101-4.
106. Ashikur RM, Hasanuzzaman MD, Mofizur RM, Zahan SI, Muhuri RS. Evaluation of antibacterial activity of study of leaves of *Tabernaemontana divaricata* (L.). *Int. Res. J. Pharm*. 2011; 2:123-7.
107. Gopinath SM, Suneetha TB, Mruganka VD, Ananda S. Evaluation of antibacterial activity of *Tabernaemontana divaricata* (L.) leaves against the causative organisms of bovine mastitis. *International Journal of Research in Phytochemistry and Pharmacology*. 2011; 1(4):211-3.
108. Pushpa B, Latha KP, Vaidya VP, Shruthi A, Shweath C. In vitro anthelmintic activity of leaves extracts of *Tabernaemontana coronaria*. *International Journal of ChemTech Research*. 2011; 3(4):1788-90.
109. Shaker IA, Inampudi S, Rayapu V. Antimicrobial activity assay of *Tabernaemontana coronaria*. *Int. J. Bioassays (IJB)*. 2012; 1:4-5.
110. Haniffa MA, Kavitha K. Antibacterial activity of medicinal herbs against the fish pathogen *Aeromonas hydrophila*. *Journal of Agricultural Technology*. 2012; 8(1):205-11.
111. Sumitha J, Padmalatha C, Singh AR. Antibacterial efficacy of *Moringa oleifera* and *Tabernaemontana divaricata* flower extracts on ocular pathogens. *Int. J. Curr. Microbiol. Appl. Sci*. 2015; 4:203-16.
112. Raja A, Ashokkumar S, Marthandam RP, Jayachandiran J, Khatiwada CP, Kaviyarasu K, Raman RG, Swaminathan M. Eco-friendly preparation of zinc oxide nanoparticles using *Tabernaemontana divaricata* and its photocatalytic and antimicrobial activity. *Journal of Photochemistry*

- and Photobiology B: Biology. 2018; 181:53-8. <https://doi.org/10.1016/j.jphotobiol.2018.02.011>
113. Zhu WT, Zhao Q, Huo ZQ, Hao XJ, Yang M, Zhang Y. Taberdivamines A and B, two new quaternary indole alkaloids from *Tabernaemontana divaricata*. *Tetrahedron Letters*. 2020; 61(44):152400. <https://doi.org/10.1016/j.tetlet.2020.152400>
114. Radhika B. Comparative study of soxhlation and maceration extracts of *Tabernaemontana divaricata* leaves for antibacterial activity. *J. Nat. Prod. Plant Resour.* 2017; 7:34-9.
115. Radhika B, Vilasini S. Anti-helminthic activity of *Tabernaemontana divaricata* leaves. *International Journal of Pharmacy and Biological Sciences*. 2016; 6(4):54-8. <https://doi.org/10.21276/ijpbs.2016.6.4.8>
116. King CH. Lifting the burden of schistosomiasis-defining elements of infection-associated disease and the benefits of antiparasite treatment. *The Journal of Infectious Diseases*. 2007; 196(5):653-5. <https://doi.org/10.1086/520522>
117. Chittaragi A, Kodiyalmath J. A comparative study on anthelmintic activity of various solvent extracts of *Clavaria rosea*. *Journal of Pharmacognosy and Phytochemistry*. 2014; 3(3):29-32.
118. Khan MS. Gastroprotective effect of *Tabernaemontana divaricata* (Linn.) R. Br. Flower methanolic extract in wistar rats. *British Journal of Pharmaceutical Research*. 2011; 1(3):88. <https://doi.org/10.9734/BJPR/2011/347>
119. Ali Khan MS, Mat Jais AM, Afreen A. Prostaglandin analogous and antioxidant activity mediated gastroprotective action of *Tabernaemontana divaricata* (L.) R. Br. flower methanolic extract against chemically induced gastric ulcers in rats. *BioMed Research International*. 2013; 2013. <https://doi.org/10.1155/2013/185476>
120. Kanthlal SK, Kumar BA, Joseph J, Aravind R, Frank PR. Amelioration of oxidative stress by *Tabernaemontana divaricata* on alloxan-induced diabetic rats. *Ancient Science of Life*. 2014; 33(4):222. <https://doi.org/10.4103/0257-7941.147429>
121. Kojima I, Umezawa K. Conophylline: A novel differentiation inducer for pancreatic β cells. *The International Journal of Biochemistry and Cell Biology*. 2006; 38(5-6):923-30. <https://doi.org/10.1016/j.biocel.2005.09.019>
122. Fujii M, Takei I, Umezawa K. Antidiabetic effect of orally administered conophylline-containing plant extract on streptozotocin-treated and Goto-Kakizaki rats. *Biomedicine and Pharmacotherapy*. 2009; 63(10):710-6. <https://doi.org/10.1016/j.biopha.2009.01.006>