



# Flavonoids: A Potent Substance in Anti-ulcer and Hepatoprotective Agents

G. S. Chakraborty, Pinkal Patel\*, Sachin Sharma, Rupal K. Jani and Snigdha Das Mandal

Faculty of Pharmacy, Parul Institute of Pharmacy and Research, Parul University, Vadodara, Gujarat, India; pinkpharmacy@gmail.com

## Abstract

Herbs have been used to treat various diseases in one form or another for the betterment of mankind since the ancient ages. As a result of this usage, the situation gradually deteriorated due to overexploitation, resulting in a decrease in natural resource production. Looking in to the benefits, the flora and fauna were rehabilitated through tissue culture, conservation, plantation and other means to ensure their availability in their natural form. Nature is the best chemist, as it aids in the treatment of ailments and provides the results of them. It is impossible to find any plant that has no medical use. Ulcer and liver diseases are at an alarming rate in the developing countries. A number of drugs, which are in the existing medications for peptic ulcer and hepatic diseases, are on the comeback list due to their adverse effects and drug interactions. Drugs, which are derived from the flora, have an important role in treatment and show a higher rate of acceptance. Flavonoids are the naturally occurring, low-molecular-weight molecules seen mostly distributed in the vegetable kingdom and hold an utmost thirst in declining the generation of reactive oxygen species. Secondary compounds like apigenin, silymarin, genistein, quercetin, kaempferol and catechins have a significant role in these diseases. This article focuses on the importance of flavonoids, which can serve as potent anti-ulcer and hepatoprotective agents.

**Keywords:** Flavonoids, Herbs, Liver, Secondary Metabolites, Ulcer

## 1. Introduction

Ulcers and diseases concerning the liver are now considered serious health problems. The liver is the vital organ for metabolism, secretion and storage. Nearly a thousand deaths are found to be due to hepatic failure and they become more severe if neglected. The observations from peptic ulcer groups are almost the same count as both are the silent processes of damaging the cells and tissues in our body system. The phenomenon of developing hepatocellular carcinoma are the general form, which correlates with the disease pattern<sup>1-3</sup>. They are due to the production of reactive species occurring which are with the damaged cells and further these cells form carbon-carbon bond with

the lipids, which are in tissues and gives the major strength<sup>4</sup>.

The formation of ulcers is an outbreak in the inflamed skin or in the mucus lining, making a path in the digestive canal. The majority of ulcers are due to imbalance, which occurs in the normal systematic process, and they are associated with a higher rate of aggression in the mucosal resistance<sup>5</sup>. Both the forms of ulcers namely gastric and duodenal ulcers form the corrosive action of pepsin and hydrochloric acid present in the mucosa linked to the upper gastrointestinal tract. Ulcers generally vary in the range of about 0.3cm to 0.5 cm or even bigger in size in diameter<sup>6</sup>. The most common cause of duodenal ulcers in adult males is due to lifestyle habits. The occurrence of gastric ulcers

\*Author for correspondence

occurs in the mid-age to older age to imbalance food habits and socio-economic class of pupils<sup>7</sup>.

On the other hand, the liver, being the largest organ in the system, holds a fundamental function that serves in the regulation of various processes involving carbohydrates, proteins, fatty substances, the removal of toxins present in the body, the secretion of fluids from gastric juice, and finally, vitamins. These elements make up a healthy liver in the body<sup>8,9</sup>.

- Owing to these functions hepatic ailments become a threat in the community and hold their position to date in the world<sup>10</sup>. The following factors are associated with liver diseases: Generation of higher quantities of free radicals, which fails its own defensive mechanism and damages the process of liver metabolism, leading to diseases such as fatty liver, jaundice, cirrhosis, etc.
- Poisoning with chemicals related to carbon tetrachloride, which in turn leads to the accumulation of lipid particles and eventually necrosis.
- Solvents such as ethanol, have a capacity to generate free radicals, which decrease the potential benefits of the enzymatic studies.

The classification of liver diseases is in three different sections, like<sup>11-14</sup>:

- Inflammatory cause of the liver
- Non-inflammatory, like hepatitis
- Degenerative, like cirrhosis.

The application of primitive methods was to be viewed as the best relief from ailments by having a mechanism based on the traditional system and usage of herbs by ancient people<sup>15</sup>. The available options in the treatment of these diseases were still limited and lacking because the root causes were different for different group of pupils<sup>16</sup>. For the search for newer drugs, the screening models were thoroughly assessed and are listed in Table 1.

There are a number of medicinal plants used commonly in the profile of dosing for liver disorders. This article aims to provide a complete list of plants and herbs that are beneficial in eradicating liver disorders (Table 2).

The need and importance of herbal drugs are generally growing due to their better acceptability and compatibility with mankind. Various medicinal plants

**Table 1.** Comparison between models<sup>17</sup>

| Anti-ulcer activity   | Hepatoprotective activity  |
|---|--|
| <ul style="list-style-type: none"> <li>• Histamine induced ulcers</li> <li>• Hydrochloric acid induced ulcers</li> <li>• Acetic acid induced ulcers</li> <li>• Ethanol induced ulcers<sup>18</sup></li> <li>• Aspirin induced ulcers<sup>19</sup></li> <li>• Water immersion stress induced ulcers<sup>20</sup></li> <li>• Pylorus ligation induced ulcers<sup>21</sup></li> <li>• Reserpine induced ulcers<sup>22</sup></li> <li>• Indomethacin induced ulcers<sup>23</sup></li> <li>• Serotonin induced ulcer<sup>24</sup></li> </ul> | <ul style="list-style-type: none"> <li>• Paracetamol (acetaminophen) induced hepatotoxicity</li> <li>• CCl<sub>4</sub> induced hepatotoxicity</li> <li>• Alcohol and Carbon tetrachloride induced hepatotoxicity</li> <li>• Carbon tetrachloride and paraffin induced hepatotoxicity</li> <li>• d-galactosamine/lipopolysaccharide (GalN/LPS) induces hepatotoxicity</li> <li>• Thioacetamide induced hepatotoxicity</li> <li>• Antitubercular drugs hepatotoxicity</li> </ul> |

**Table 2.** Herbs comprising the potential use of hepatoprotective activity

| Sr. No. | Name of the plant                                     | Part used          | Extraction solvent           | Chemical constituent   | Animal model     | Hepatotoxic agent                       | References |
|---------|---|--------------------|------------------------------|--|------------------|---|------------|
| 1       | <i>Aerva lanata</i><br>(Amaranthaceae)                | Leaf,<br>Root      | Aqueous alcoholic extract    | Sitosteryl plamitate,<br>hentriaconten,                                    | Rat              | Paracetamol                             | 25,<br>26  |
| 2       | <i>Artemisia capillaris</i><br>(Asteraceae)           | Whole plants       | Aqueous ethanol extract      | Eupatolin,<br>Capillartemisin A  | Rat              | Carbon tetrachloride                    | 27         |
| 3       | <i>Aphanamixis polystachya</i><br>(Meliaceae)         | Leaf               | Extract of Ethanol           | Polyprenol, lutein   | Rat              | Carbon tetrachloride                    | 28         |
| 4       | <i>Allium hirtifolium</i><br>(Alliaceae)              | Leaf               | Aqueous alcoholic extract    | Shallomin, Quercetin and Kaempferol  | Rat              | Alloxan induced                         | 29,<br>30  |
| 5       | <i>Amorphophallus paeoniifolius</i> Linn<br>(Araceae) | Tubers             | Methanol and aqueous extract | Steroids and Flavonoids  | Rat              | Paracetamol                             | 31         |
| 6       | <i>Allium sativum</i><br>(Alliaceae)                  | Bulbs              | Ethanoliceextract            | Sapogenins, Saponins, Allicin  | Rat              | Cadmium                                 | 32,<br>33  |
| 7       | <i>Berberi vulgaris</i><br>(Berberidaceae)            | Fruit              | Methanolic extract           | Berberine, oxyacanthine, and flavonoids                                    | Rat              | Carbon tetrachloride                    | 34,<br>35  |
| 8       | <i>Calendula officinalis</i><br>(Asteraceae)          | Flower             | Methanolic extract           | $\alpha$ -thujene and T-muurolol, flavonol glycosides, flavonoids          | Albino Rat       | Carbon tetrachloride                    | 36,<br>37  |
| 9       | <i>Cercissili quastrum</i><br>(Leguminoseae)          | Whole plants       | Hydro alcoholic extract      | Myricitoxide, diterpenoids, triterpenoids                                  | Rat              | Carbon tetrachloride                    | 37         |
| 10      | <i>Citrullus lanatus</i><br>( Cucurbitaceae )         | Fruits             | Methanolic extract           | Triterpenes, flavanoids, saponins  | Rat              | Carbon tetrachloride                    | 38,<br>39  |
| 12      | <i>Daucus carota</i><br>(Apiaceae)                    | Seeds              | Methanolic Extract           | Triterpenes, flavonoids  | Wister rat       | Lindane                                 | 40, 41     |
| 13      | <i>Decalepis hamiltonii</i><br>(Asclepiadaceae)       | Roots              | Aqueous extract              | 2-Hydroxy-4-methoxybenzaldehyde<br>Vanillin, Borneol                       | Rat              | Ethanol                                 | 42, 43     |
| 14      | <i>Eclipta alba</i><br>(Asteraceae)                   | Leaves             | Alcoholic extract            | Terpenes, flavonoids   | Rat and mice     | Carbon tetrachloride                    | 44         |
| 15      | <i>Epaltis divaricata</i><br>(Compositae)             | Whole Plants       | Aqueous Extract              | Flavonoids, Ascorbic acid, carotenoids, tannis and lignins                 | Mice             | Carbon tetrachloride                    | 45, 46     |
| 16      | <i>Embllica officinalis</i><br>(Euphorbiaceae)        | Fruit              | Hydroalcoholic Extract       | kaempferol-3-Rhamnoglucoside<br>Quercetin-3-Rhamnoglucoside,<br>Stepposide | Rats             | Rifampicin, Isoniazide and pyrazinamide | 47         |
| 17      | <i>Hypericum perforatum</i><br>(Clusiaceae)           | Dried aerial parts | Alcoholic Extract            | Flamin, kaempferol, naringenin and isohelichrysin                          | Male albino Mice | Carbon tetrachloride                    | 48         |

|    |   |              |                        |                                       |                    |   |        |
|----|---|--------------|------------------------|---------------------------------------|--------------------|---|--------|
| 18 | <i>Lactuca indica</i><br>(Asteraceae)       | Aerial parts | 95% Methanolic Extract | Sterols and flavonoids                | Rat                | Carbon tetrachloride                        | 49     |
| 19 | <i>Mentha pieperata</i><br>(Labiatae)       | Leaves       | Aqueous Extract        | Myrcene, pipitone, eugenol, menthone. | Albino Wistar Rats | Carbon tetrachloride                        | 50     |
| 20 | <i>Silybum marianum</i><br>(Asteraceae)     | Whole plants | Ethanol extract        | Isoflavonoid and Silymarin            | Rats               | Carbon tetrachloride induced liver toxicity | 51, 52 |
| 21 | <i>Taraxacum officinale</i><br>(Asteraceae) | Roots        | Aqueous extract        | carotenoids, lutein, flavonoids       | Rats               | Carbon tetrachloride induced liver toxicity | 53, 54 |

**Table 3.** Herbs and shrubs with anti-ulcer activity

| SI No | Name of the plant                               | Part used           | Extraction solvent                 | Chemical constituent                                      | Animal Model           | Ulcer creating agent                        | References |
|-------|---|---------------------|------------------------------------|---|------------------------|---|------------|
| 1     | <i>Cynodon dactylon</i><br>(Poaceae)            | Aerial parts        | Alcoholic extract                  | Flavonoids  | Albino Rats            | Pylorus ligation                            | 55         |
| 2     | <i>Cucurbita pepo</i><br>(Cucurbitaceae)        | Seed                | Methanolic extract                 | Terpenoids, cucurbitacin                                  | Rat                    | Stress induced                              | 56         |
| 3     | <i>Boswellia serrata</i><br>(Burseraceae)       | Bark                | Petroleum ether                    | squalene, polyprenol, lutein                              | Male albino rat        | Aspirin                                     | 57         |
| 4     | <i>Pycnanthus angolensis</i><br>(Myristicaceae) | Bark                | Ethanol extract                    | Flavonones, epicatechin and (+)-catechin                  | Male Albino Wistar rat | Ethanol                                     | 58         |
| 5     | <i>Alstonia scholaris</i><br>(Apocyanaceae)     | Bark                | Ethanol Extract                    | Coumarins, flavonoids, phlobatannin, saponins and tannins | Male albino rat        | Pylorus ligation                            | 59         |
| 6     | <i>Asparagus racemosus</i><br>(Asparagaceae)    | Roots               | Methanolic extract                 | Shatavarin, flavonoid                                     | Male albino rats       | Indomethacin                                | 60         |
| 7     | <i>Azadirachta indica</i><br>(Meliaceae)        | Leaf                | Aqueous extract                    | Flavonoids, proteins                                      | Rat                    | Indomethacin treated, ethanol and histamine | 61, 62     |
| 8     | <i>Butea foandosa</i><br>(Fabaceae)             | Leaves              | Chloroform and ethanolic extract   | Butrin, flavonoids  | Rat                    | Hydrochloric acid                           | 63         |
| 9     | <i>Bauhinia variegata</i><br>(Fabaceae)         | Leaves              | Aqueous extract, ethanolic extract | Flavonoids  | Rat                    | Aspirin                                     | 64         |
| 10    | <i>Hibiscus Rosa</i><br>(Malvaceae)             | Leaves              | Methanol extract                   | Flavonoids, anthocyanins, quercetin                       | Rat                    | Pyloric ligation                            | 65         |
| 11    | <i>Murrya koenigii</i><br>(Rutaceae)            | Root Stem and Leave | Methanol extract                   | Monoterpenes  | Albino Rat             | Hydrochloric acid, indomethacin             | 66         |

|    |  |                                |                                       |  |            |  |        |
|----|--|--------------------------------|---------------------------------------|--|------------|--|--------|
| 12 | <i>Ocimum Sanctum</i><br>(Lamiaceae)                 | Leaves                         | Alcoholic extra                       | Alkaloids,<br>saponins,apigenin                                | Rat        | Aspirin and<br>ethanol                                 | 67     |
| 13 | <i>Moringa oleifera</i><br>(Moringaceae)             | Leaves                         | Alcoholic<br>extract                  | Alkaloids, flavonoids,<br>zeatin, kaempferom,<br>and terpenoid | Rat        | Aspirin and<br>ethanol<br>induced                      | 68     |
| 14 | <i>Sophorasub<br/>prostrata</i><br>(Fabaceae)        | Whole<br>plants                | Alcoholic<br>extract                  | Sophoradin   | Rat        | Pylorus ligated  | 69     |
| 15 | <i>Glicyrriza glabra</i><br>(Fabaceae)               | dried<br>roots and<br>rhizomes | Ethanol<br>extract                    | Flavonoids, Glabra   | Swiss mice | Water<br>immersion<br>and acetic acid<br>induced ulcer | 70     |
| 16 | <i>Sylibinmarium</i><br>(Asteraceae)                 | Whole<br>plants                | Methanolic<br>extract                 | Sylimarin  | Rat        | Ethanol, cold<br>resistance,<br>pylorous<br>ligation   | 71     |
| 17 | <i>Genistar umelica</i><br>(Fabaceae)                | Whole<br>plant                 | Methanolic<br>extract                 | Genistin, luteoline-7-<br>glycoside                            | Rat        | Pylorus<br>ligation                                    | 72, 73 |
| 18 | <i>Eucalypus<br/>maculate</i><br>(Myrtaceae)         | Leaves                         | Methanolic<br>extract                 | Quercetin  | Rat        | Cold resistance<br>and pylorus<br>ligation             | 74     |
| 19 | <i>Rhammus<br/>procubens</i><br>(Rhamnaceae)         | Whole<br>plants                | Aqueous and<br>Ethanollic<br>extracts | Kaempferol   | Rat        | Pylorous<br>ligation                                   | 75     |
| 20 | <i>Anacardium<br/>accidentate</i><br>(Anacardiaceae) | leaves                         | Hydroalcoholic<br>Extract             | Catechins  | Rat        | Pylorous<br>ligation<br>HCl<br>ethanol                 | 76, 77 |

are used traditionally in the treatment of peptic ulcers (Table 3).

## 2. Some Important Flavonoids

### 2.1 Luteolin

It belongs to the flavonoid group of the plant kingdom and is isolated from *Reseda luteola*. A yellow microcrystalline powder. Combination is used in traditional herbal remedies as potent scavenging properties<sup>78</sup> (Figure 1).

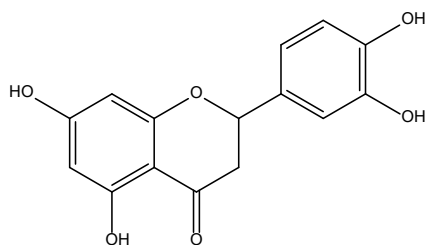


Figure 1. Luteolin.

### 2.2 Acacetin

This compound belongs to an O-methylated flavone found in *Robinia pseudoacacia*. Shows potential benefits in curing liver against chemical induced moiety<sup>79</sup> (Figure 2).

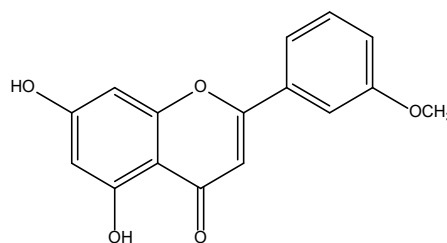


Figure 2. Acacetin.

### 2.3 Apigenin

Scientifically belonging to the flavone class. It combines the sugar portion with the non-sugar moiety. A yellow crystalline solid, which shows potential effects on ulcer activity<sup>80</sup> (Figure 3).

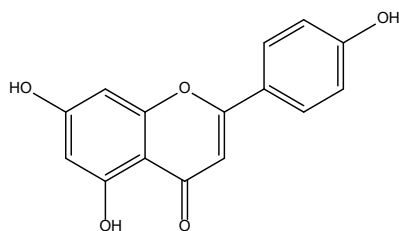


Figure 3. Apigenin.

### 2.4 Silymarine

A derived complex with a combination of silybin, silydianin, and silychris derived from the milk thistle plant<sup>81</sup> (Figure 4).

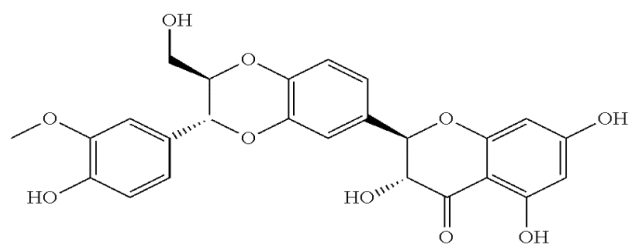


Figure 4. Silymarine.

### 2.5 Kaempferol

Belongs to the class of flavonols. Tan oily in nature with yellow crystalline solid with solubility in water and organic solvents. Possess a greater extent of antioxidant activity<sup>82</sup> (Figure 5).

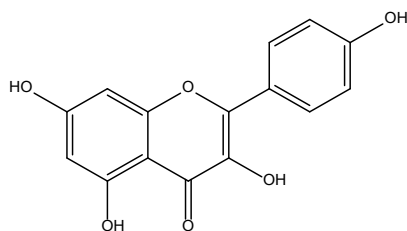


Figure 5. Kaempferol.

### 2.6 Salvigenin

Obtained from the plant *Dorema glabrum*. Salvigenin is a highly potent free radical scavenging molecule, and apart from that, the synergistic effects are seen as hepatoprotective activity and in tumour cells<sup>80</sup> (Figure 6).

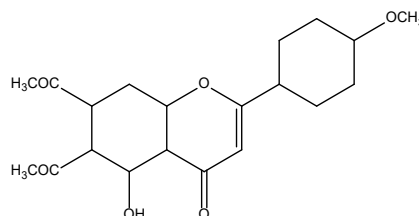


Figure 6. Salvigenin.

### 2.7 Quercetin

Commonly found fruits, vegetables, leaves, and grains. Possess a broader range in curing various pharmacological effects such as antioxidants, ROS etc.,<sup>83</sup> (Figure 7).

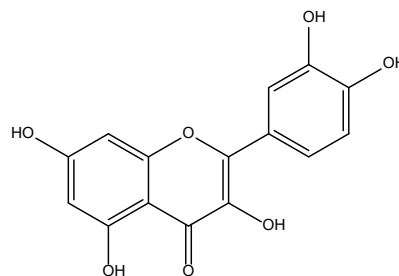


Figure 7. Quercetin.

## 3. Conclusion

Natural herbs and plants serve as the best remedy for the liver and ulcer healing properties due to their potential benefits, as they have fewer side effects and more beneficial parameters. These are due to the presence of metabolites, which show the potency of the activity. Owing to the importance of flavonoids, they are also useful in the degeneration of cells, as they are significantly involved in the process of oxidation developed by cells on intra- and extra-cellular parameters. Flavonoids are seen potential in the healing process of gastric juice owing to mucoprotective activity. Compounds like silymarin, quercetin, apigenin, salvigenin luteolin etc., have shown its importance in these diseases mainly.

Therefore, the search for newer medications along with these combinations can lead to the eradication of these diseases, which are silent in nature but can be fatal to humankind. Hence, the search is on for these drugs and active moiety to cure the ailments with maximum benefits.

## 4. References

1. Eduardo MS, Eduardo MB, Isela AG, Maria TS, Jose GS, Mirandeli B, Angel MG, Manuel GL, Leopoldo AF and Jose AM. Review of natural products with hepatoprotective effects. *World J Gastroenterol.* 2014; 20(40):14787-1480. <https://doi.org/10.3748/wjg.v20.i40.14787>
2. Lin JH and Lu AY. Role of pharmacokinetics and metabolism in drug discovery and development. *Pharmacol Rev.* 1997; 49:403-449.
3. Dong WL, Yun TK, Yu-Jung J, Young-Eon K and Daeseok H. Anti-obesity effect of *Artemisia capillaris* extracts in high-fat diet-induced obese rats. *Molecules.* 2013; 18:9241-9252. <https://doi.org/10.3390/molecules18089241>
4. Handa SS and Sharma A. Hepatoprotective activity of Andrographolide from *Andrographis paniculata* against carbon tetrachloride. *Ind. J. Med. Res.* 1990; 92:276-92.
5. Sharma D and Bhatt S. A comprehensive review on ulcer potential of medicinal plants. *Inter National Journal of Pharmacy and Pharmaceutical Science.* 2014; 6(10):829-30.
6. Malfertheiner P, Chan FK and McColl KE. Peptic ulcer disease. *The Lancet.* 2009; 374(9699):1449-1461. [https://doi.org/10.1016/S0140-6736\(09\)60938-7](https://doi.org/10.1016/S0140-6736(09)60938-7)
7. Dey NC and Dey TK. A text book of pathology. Calcutta: New Central Book Agency. 2002.
8. Ward FM and Daly MJ. Hepatic disease. In: *Clinical Pharmacy and Therapeutics.* New York: Churchill Livingstone. 1999; 195-212.
9. Shanani S. Evaluation of hepatoprotective efficacy of APCL-A polyherbal formulation *in vivo* in rats. *Indian Drugs.* 1999; 36:628-631.
10. Asha VV and Pushpangadan P. Preliminary evaluation of the anti-hepatotoxic activity of *Phyllanthus kozhikodanus*, *Phyllanthus maderaspatensis* and *Solanum indicum*. *Fitoterapia.* 1998; 59:255-259.
11. Harshmohan. The liver, biliary tract and exocrine pancreas. *Text book of pathology.* 4th ed. New Delhi. Jaypee Brothers Medical Publishers (P) Ltd. 2002.
12. Gupta KA, Ganguly P, Majumder KU and Ghosal S. Hepatoprotective and antioxidant effect and steroidal saponins of *Solanum xanthocarpum* and *Solanum nigrum* in paracetamol induced hepatotoxicity in rats. *Pharmacologyonline.* 2009; 1:75.
13. Sandhir R and Gill K. Hepatoprotective effects of Liv-52 on ethanol induced liver damage in rats. *Ind. J. Expt. Biol.* 1999; 37:762-66.
14. Kumar CH, Ramesh A, Kumar JNS and Ishaq BM. A review on hepatoprotective activity of medicinal plants. *Int J Pharm Sci Res.* 2011; 2:501-515.
15. Subbarayappa V. The roots of ancient medicine: an historical outline. *Journal of Biosciences.* 2001; 26(2):135-143. <https://doi.org/10.1007/BF02703637>
16. Girish C and Pradhan SC. Indian herbal medicines in the treatment of liver diseases: problems and promises. *Fundamental and Clinical Pharmacology.* 2012; 26(2):180-189. <https://doi.org/10.1111/j.1472-8206.2011.01011.x>
17. Mukherjee P. Quality control of herbal drugs, Business horizons pharmaceutical publisher, New Delhi 4th ed. 2010. 529-541.
18. Thamotharan G, Shekar G and Ganesh T. Anti ulcerogenic effects of *Lanata camara* Linn leaves on *in vivo* test models in rats. *Asian J Pharm Clinical Res.* 2010; 3(3):57-60.
19. Goodman and Gilman. *The pharmacological basis of therapeutics*, 9th ed. New York: Mc Graw-Hill. 1996.
20. Vinod N, Albina A, Gopalakrishna HN and Dorababu P. Evaluation of the anti-ulcer activity of NR-ANX-C (a polyherbal formulation) in aspirin and pyloric ligation induced gastric ulcers in albino rats. *Indian J Med Res.* 2010; 132:218-223.
21. Al-Yahya MA, Rafatullah S, Mossa JS and Ageel AM. Gastric anti-secretory, antiulcer and cytoprotective properties of ethanolic extract of *Alpinia galangal* Willd in Rats. *Phytotherapy Res.* 1990; 4(3): 112-4. <https://doi.org/10.1002/ptr.2650040308>
22. Rang HP. *Pharmacology: Churchill Livingstone Elsevier.* 2009
23. Kokate CK, Purohit AP and Gokhale SB. *Pharmacognosy.* 13<sup>th</sup> ed. Pune: NiraliPrakashan Publisher. 2007
24. Warriar PK, Nambiar VP and Raman KC. *Indian medicinal plants: A compendium of 500 species.* Hyderabad: Orient Longman Publisher. 1994.
25. Vendrichelvan T, Jegdeesan M, Palaniappan MS, Murali NP and Sasikumar K. Diuretic and anti inflammatory activity of *Arena lanata* in rat. *Indian Journal of pharmaceutical science.* 2000; 62(4):300-302.
26. Rajesh, Chitra K and Padmaa MP. *Aervalanata* (Linn) juss ex Shult-An Review. *Indian journal of natural product and resources.* 2011; 2(1):5-9.
27. Consolacion SY, Ragasa MA, Leonora TA, Vincent AS, Maria N, Lorraine G, Bugayong SD, Jacinto WL and Shen CC. Terpenoids and sterol from *Aphanamixis polystachya*. *Journal of Chemical and Pharmaceutical Research.* 2014; 6(6):65-68.

28. Gole MK and Dasgupta S. Role of plant metabolites in toxic liver injury. *Asia Pacific Journal of Clinical Nutrition*. 2002; 11:48-50. <https://doi.org/10.1046/j.1440-6047.2002.00265.x>
29. Kazemi S, Asgary S, Moshtaghian J, Rafieian M, Adelnia A and Shamsi F. Liver-protective effects of hydroalcoholic extract of *Allium hirtifolium* Boiss. In rats with alloxan-induced diabetes mellitus. *Arya Atheroscler*. 2010; 6:11-15.
30. Rose P, Whiteman M, Moore PK and Zhu YZ. Bioactive S-alk(en)ylcysteine sulfoxide metabolites in the genus *Allium*: the chemistry of potential therapeutic agents. *Nat. Prod. Rep*. 2005; 22:351-368. <https://doi.org/10.1039/b417639c>
31. Hurkadale PJ, Shelar PA, Palled SG, Mandavkar YD and Khedkar AS. Hepatoprotective activity of *Amorphophallus paeoniifolius* tubers against paracetamol-induced liver damage in rats. *Asian Pacific Journal of Tropical Biomedicine*. 2012; 238-242. [https://doi.org/10.1016/S2221-1691\(12\)60167-1](https://doi.org/10.1016/S2221-1691(12)60167-1)
32. Amagase H, Petesch BL, Matsuura H, Kasuga S and Itakura Y. Intake of garlic and its bioactive components. *J. Nutr*. 2001; 131:955S-962S. <https://doi.org/10.1093/jn/131.3.955S>
33. Obioha UE, Suru SM, Ola-Mudathir KF and Faremi TY. Hepatoprotective potentials of onion and garlic extracts on cadmium-induced oxidative damage in rats. *Biol. Trace Elem. Res*. 2009; 129:143-156. <https://doi.org/10.1007/s12011-008-8276-7>
34. Fatehi M, Saleh TM, Fatehi-Hassanabad Z, Farrokhfal K, Jafarzadeh M and Davodi SA. Pharmacological study on *Berberis vulgaris* fruit extract. *J. Ethnopharmacol*. 2005; 102:46-52. <https://doi.org/10.1016/j.jep.2005.05.019>
35. Domitrovic R, Jakovac H and Blagojevic G. Hepatoprotective activity of berberine is mediated by inhibition of TNF- $\alpha$ , COX-2, and iNOS expression in CCl<sub>4</sub>-intoxicated mice. *Toxicology*. 2011; 280:33-43. <https://doi.org/10.1016/j.tox.2010.11.005>
36. Preethi KC and Kuttan R. Hepato and reno protective action of *Calendula officinalis* L. flower extract. *Indian J. Exp. Biol*. 2009; 47:163-168.
37. Valan MF, Britto AJD and Venkataraman R. Phytoconstituents with hepatoprotective activity. *Int. J. Chem. Sci*. 2010; 8(3):1421-1432.
38. Bartalis J. Hepatoprotective activity of cucurbitacin [dissertation]. Brookings: South Dakota State University. 2005.
39. Altas S, Kizil G, Kizil M, Ketani A and Haris PI. Protective effect of Diyarbakir watermelon juice on carbon tetrachloride-induced toxicity in rats. *Food Chem. Toxicol*. 2011; 49:2433-2438. <https://doi.org/10.1016/j.fct.2011.06.064>
40. Balasubramaniam P, Pari L and Menon VP. Protective effect of carrot (*Daucus carota* L.) against lindane-induced hepatotoxicity in rats. *Phytotherapy Research*. 1998; 12:434-436. [https://doi.org/10.1002/\(SICI\)1099-1573\(199809\)12:6<434::AID-PTR310>3.0.CO;2-U](https://doi.org/10.1002/(SICI)1099-1573(199809)12:6<434::AID-PTR310>3.0.CO;2-U)
41. Vasudevan M, Gunnam KK and Parle M. Antinociceptive and anti-inflammatory properties of *Daucus carota* seeds extract. *J. Health Sci*. 2006; 52:598-606. <https://doi.org/10.1248/jhs.52.598>
42. Srivastava A and Shivanandappa T. Hepatoprotective effect of the aqueous extract of the roots of *Decalepis hamiltonii* against ethanol-induced oxidative stress in rats. *Hepatology Research*. 2006; 35:267-275. <https://doi.org/10.1016/j.hep-res.2006.04.011>
43. Harish R, Divakar S, Srivastava A and Shivanand T. Isolation of Antioxidant Compounds from the Methanolic Extract of the Roots of *Decalepis hamiltonii*(Wight and Arn.). *J. Agric. Food Chem*. 2005; 53:7709-7714. <https://doi.org/10.1021/jf051047c>
44. Singh B, Saxena AK, Chandan BK, Agarwal SG, Bhatia MS and Anand KK. Hepatoprotective effect of ethanolic extract of *Eclipta alba* on experimental liver damage in rats and mice. *Phytotherapy Research*. 1993; 7:154-158. <https://doi.org/10.1002/ptr.2650070212>
45. Hewawasam RP, Jayatilaka KAPW, Pathirana C and Mudduwa LKB. Hepatoprotective effect of *Epilates divaricata* extract on carbon tetrachloride induced hepatotoxicity in mice. *Indian J Med Res*. 2004; 120:30-34.
46. Rusu MA, Tamas M, Puica C, Roman I and Sabadas M. The hepatoprotective action of ten herbal extracts in CCl<sub>4</sub> intoxicated liver. *Phytotherapy Research*. 2005; 19:744-749. <https://doi.org/10.1002/ptr.1625>
47. Tasduq SA, Kaiser P, Gupta DK, Kapahi BK, Jyotsna S Maheshwari HS and Johri RK. Protective effect of a 50 hydroalcoholic fruit extract of *Embllica officinalis* against anti-tuberculosis drugs induced liver toxicity. *Phytotherapy Research*. 2005; 19:193-197. <https://doi.org/10.1002/ptr.1631>
48. Ozturk Y, Aydin S, Baser KHC, Kirimer N and Kurtar O. Hepatoprotective activity of *Hypericum perforatum* L. alcoholic extract in rodents. *Phytotherapy Research*. 1992; 6:44-46. <https://doi.org/10.1002/ptr.2650060111>
49. Ki HK, Young HK, Kang R and Lee. Isolation of hepatoprotective phenylpropanoid from *Lactuca indica*. *Natural Product Sciences*. 2010; 16(1):6-9.
50. Shah PP and Mello PMD. A review of medicinal uses and pharmacological effect of *Mentha piperita*. *Natural product radiance*. 2014; 3(4):214-221.
51. Salam OM, Sleem AA, Omara EA and Hassan NS. Hepatoprotective effects of misoprostol and silymarin on carbon tetrachloride-induced hepatic damage in rats. *Fundam. Clin. Pharmacol*. 2009; 23:179-188. <https://doi.org/10.1111/j.1472-8206.2008.00654.x>
52. Kim SH, Cheon HJ, Yun N, Oh ST, Shin E, Shim KS and Lee SM. Protective effect of a mixture of Aloe vera and *Silybum marianum* against carbon tetrachloride-induced acute



- hepatotoxicity and liver fibrosis. J. Pharmacol. Sci. 2009; 109:119-127. <https://doi.org/10.1254/jphs.08189FP>
53. Fallah H, Zarrei M, Ziai M, Mehrazma M, Alavian SM and Kianbakht S. The effects of *Taraxacum officinale* L. and *Berberis vulgaris* L. root extracts on carbon tetrachloride induced liver toxicity in rats. J. Med. Plants. 2010; 9:45-52.
54. Mahesh A, Jeyachandran R, Cindrella L, Thangadurai D, Veerapur V and Muralidhara Rao D. Hepatocurative potential of sesquiterpene lactones of *Taraxacum officinale* on carbon tetrachloride induced liver toxicity in mice. Acta Biol. Hung. 2010; 61:175-190. <https://doi.org/10.1556/ABiol.61.2010.2.6>
55. Kondaveeti SB, Ivvala AS, Dadduru K, Basha SS and Sailaja I. Indigenous effect of cynodonectylon in experimental induced ulcer and gastric secretion. Internal Research Journal of Pharmacy. 2012; 3(5):301-304.
56. Gill NS and Bali M. Isolation of antiulcer Cucurbitane type triterpenoids from the seed of *Cucurbita pepo*. Research Journal of Phytochemistry. 2011; 5(2):72-79. <https://doi.org/10.3923/rjphyto.2011.70.79>
57. Khaja Z, Mangamoori LN, Muna A and Mohammed I. Evaluation of antiulcer activity of *Boswellia serrata* bark extracts using aspirin induced ulcer model in albino rats. J. Med. Allied Sci. 2011; 1(1):14 -20.
58. Margaret OS and Adepero OA. Antinociceptive and antiulcer activities of *Pycnanthus angolensis*, Revista Brasileira de Farmacognosia. 2015; 25:252-257. <https://doi.org/10.1016/j.bjp.2015.05.004>
59. Arulmozhi S, Papiya MM, Sathiyarayanan L and Prasad A. Analgesic, anti-inflammatory and anti-ulcerogenic activities of fractions from *Alstonia scholaris*. Pharmacologia. 2012; 3:132-7. <https://doi.org/10.5567/pharmacologia.2012.132.137>
60. Bhatnagar M and Sisodia SS. Antisecretory and antiulcer activity of *Asparagus racemosus* against indomethacin plus pyloric ligation-induced gastric ulcer in rats. J. Herb. Pharmacother. 2006; 6(1):13-20. [https://doi.org/10.1080/J157v06n01\\_02](https://doi.org/10.1080/J157v06n01_02)
61. Chattopadhyay I, Nandi B, Chatterjee R, Biswas K, Bandyopadhyay U and Banerjee RK. Mechanism of antiulcer effect of Neem (*Azadirachta indica*) leaf extract: effect on H<sup>+</sup>-K<sup>+</sup>-ATPase, oxidative damage and apoptosis. Inflammo. Pharma. 2004; 12(2):153-76. <https://doi.org/10.1163/1568560041352257>
62. Kottaimuthu R. Ethnobotany of the Valaiyans of Karandamalai, Dindigul District, Tamil Nadu. Ethnobotanical Leaflets. 2009; 12:195-203.
63. Ramesh L and Ranirukmini RK. Antiulcerogenic study of different extracts of *Butea frondosa* Roxb in albino mice. J. Pharmacog. 2010; 1(1):6-9.
64. Rajkapoor B, Jayakar B, Anandar R and Kavinani S. Antiulcer activity of *Bauhinia variegata* Linn. Journal of Natural Remedies. 2003; 3(2):215-217.
65. Srivastava S, Jiaswal J, Gautam H, Sharma S and Rao CHV. Antiulcer activity of methanolic extract of *Hibiscus rosa-sinensis* leaves. Int. J. Pharm. Pharm Sci. 2013; 5(3):829-30.
66. Mitra PK. Comparative Evaluation of Anti-Ulcer Activity of Root Stem and Leave of *Murraya koenigii* (Linn.) Spreng in Rats. Journal of Medicinal Plants Studies. 2013; 1(3):158-165. <https://doi.org/10.14259/pm.v1i1.12>
67. Dharmani P, Kulchibhota VK, Mauriya R, Srivastav S, Sharma S and Patil G. Evaluation of anti ulcerogenic and ulcer healing property of *Ocimum sanctum* Linn. J. Ethnopharmacol. 2004; 93:197-206. <https://doi.org/10.1016/j.jep.2004.02.029>
68. Verma VK, Singh N, Saxena P and Singh R. Anti-Ulcer and Antioxidant Activity of *Moringa oleifera* (Lam) Leaves against Aspirin and Ethanol Induced Gastric Ulcer in Rats. Res. J. of Pharmaceuticals. 2012; 2(2):46-57.
69. Sasajima M, Nakane S, Saziki R, Saotome H, Hatayama K, Kyogoku K and Tanaka I. Studies on the anti-ulcer effects of isoprenyl flavonoids. Nihon Yakurigaku Zasshi. 1978; 74(8):897-905. <https://doi.org/10.1254/fjp.74.897>
70. Vahid N, Ehsan A, Mostafa M, Hadi KG and Amina J. Antiulcer Properties of *Glycyrrhiza glabra* L. Extract on experimental models of gastric ulcer in Mice. Iranian Journal of Pharmaceutical Research. 2015; 14 (4):1163-1170.
71. Tumova L, Rimakova J, Tuma J and Dusek J. *Silybum marianum* in vitro flavonolignan production. Plant Soil Environ. 2006; 52(10):454-8. <https://doi.org/10.17221/3466-PSE>
72. Rainova L, Nakov N, Bogdanova S, Minkov E and Staneva Stoytcheva D. Ulceroprotective activity of the avonoids of *Genistarumelica* Vel. Phytother Res. 1988; 2:137-139. <https://doi.org/10.1002/ptr.2650020307>
73. Rao CV, Ojha SK, Radhakrishnan K, Govindarajan R, Rastogi S, Mehrotra S and Pushpangadan PJ. Antiulcer activity of *Uleria salicifolia* rhizome extract. Ethnopharmacol. 2004; 91(2):243-9. <https://doi.org/10.1016/j.jep.2003.12.020>
74. Willoughby GA and Hayward AC. Degradation of delignified *Eucalyptus maculate* by *Cellvibrio mixtus*. Letters in Applied Microbiology. 1988; 6(5):99-103. <https://doi.org/10.1111/j.1472-765X.1988.tb01224.x>
75. Goel RK and Pendey VK. Anti-inflammatory and antiulcer effects of kaempferol, a flavone, isolated from *Rhamnus procumbens*. Indian J. Exp. Biol. 1988; 26:121-24.
76. Konan NA and Bacchi EM. Antiulcerogenic effect and acute toxicity of a hydroethanolic extract from the cashew (*Anacardium occidentale* L.) leaves. J. Ethnopharmacol. 2007; 112(2):237-42. <https://doi.org/10.1016/j.jep.2007.03.003>
77. Adeleye OE, Okediran BS, Rahman SA and Ajibola ES. Effect of intragastric administration of crude aqueous leaf

- extract of *Anacardium occidentale* on gastric acid secretion in rats. Niger J. Physiol. Sci. 2010; 25(1):59-62.
78. Barnaulov OD, Manicheva OA, Zapesochaya GG, Shelyuto VL and Glyzin VI. Effects of certain flavonoids on the ulcerogenic action of reserpine in mice. Khim. Farm. Zh. 1982; 16:300-303. <https://doi.org/10.1007/BF00777270>
79. Salama MM, Ezzat SM and Sleem AA. A new hepatoprotective flavone glycoside from the flowers of *Onopordum alexandrinum* growing in Egypt. Z. Naturforsch C. 2011; 66(5-6):251-259. <https://doi.org/10.1515/znc-2011-5-608>
80. Okuyama E, Yamazaki M and Ishii Y. Isolation and identification of ursolic acid-related compounds as the principles of glechoma hederacea having an antiulcerogenic activity. Shoyakugaku Zasshi. 1983; 37:52-55.
81. Wollenweber E, Dorr M and Rustiyan A. Doremaacheri, the first umbelliferous plant found to produce exudate flavonoids. Phytochem. 1995; 38:1417. [https://doi.org/10.1016/0031-9422\(94\)00840-P](https://doi.org/10.1016/0031-9422(94)00840-P)
82. Majid AS, Najme KF, Nafiseh A, Ali F, Ebrahim AA and Mahmoud RK. Medicinal plants with hepatoprotective activity in Iranian folk medicine. Asian Pacific Journal of Tropical Biomedicine. 2015; 5(2):146-157 [https://doi.org/10.1016/S2221-1691\(15\)30159-3](https://doi.org/10.1016/S2221-1691(15)30159-3)
83. Xi Chen. Protective effects of quercetin on liver injury induced by ethanol. Pharmacognosy Magazine. 2010; 6(22):136-141. <https://doi.org/10.4103/0973-1296.62900>