



Tricetin and Tricin: An Overview of the Chemistry, Sources, Contents, and Pharmacological Properties of these Flavones

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Abstract

In this overview, information on the chemistry, sources, contents, and pharmacological properties of two flavones, namely, tricetin (TCT) and tricin (TC), is updated. TCT occurs mainly in honey and pollen of plant species belonging to the genus *Eucalyptus* of the family Myrtaceae. TC is found in monocotyledon species of the family Poaceae, occurring mainly in cereal crops such as oats, barley, rice, wheat, and corn, and in bamboo species. The chemical structure of TCT contains two hydroxyl (OH) groups at C5 and C7 of ring A and three OH groups at C3', C4', and C5' of ring B, with no methoxy (OCH₃) groups. TC has two OH groups at C5 and C7 of ring A, two OCH₃ groups at C3' and C5', and one OH group at C4' of ring B, i.e., at both sides of the C4' OH group. This renders greater bioavailability, higher metabolic stability, and better intestinal absorption to TC than TCT. In this overview, TCT and TC have eight and seven studies on anti-cancer properties, and 14 and 31 studies on other pharmacological properties, respectively. Both flavones are equally strong in terms of cytotoxicity towards cancer cells. With greater bioavailability, higher metabolic stability, and better intestinal absorption, the other pharmacological properties of TC are stronger than TCT, but not for anti-cancer properties.

Keywords: Hydroxylated Flavones, Methoxylated Flavones, Myricetin

Abbreviations: ABCG2 = ATP-binding cassette transporter G2, ABI = anaphase bridging index, AC = acute colitis, AChE = acetylcholinesterase, Akt = protein kinase B, AMPK = adenosine 5'-monophosphate activated protein kinase, AOM = azoxymethane, AP = acute pancreatitis, ATP = adenosine triphosphate, BaP = benzo(a)pyrene, BCRP = breast cancer resistance protein, CCL = CC motif ligand, CDK9 = cyclin-dependent kinase 9, COPD = chronic obstructive pulmonary disease, COX = cyclooxygenase, CXCL11 = C-X-C motif chemokine 11, DR = death receptor, DSS = dextran sulphate sodium, EC₅₀ = 50% effective concentration, Egr-1 = early growth response 1, ERK = extracellular signal-regulated kinase, FAK = focal-adhesion-kinase, HCMV = human cytomegalovirus, HEL = human embryonic lung, HIF = heterodimeric transcription factor, HMS = human mesenchymal stem, HO-1, heme oxygenase-1, HS = hepatic stellate, I/R = ischemia/reperfusion, IC₅₀ = 50% inhibitory concentration, IL = interleukin, JNK = c-Jun N-terminal kinase, LDL = low-density lipoprotein, LLC = Lewis lung carcinoma, LOX-1 = lectin-like ox-LDL, LPS = lipopolysaccharide, MAPK = mitogen-activated protein kinase, MDI = mixture of 3-isobutyl-1-methylxanthine (M), dexamethasone (D) and insulin (I), MDM = murine double minute, MI = mitotic index, MIC = minimum inhibitory concentration, MMP = matrix metalloproteinase, mTOR = mammalian target of rapamycin, MVT = membrane vascular transport, NF-κB = nuclear factor-kappa B, NO = nitric oxide, Nrf2 = nuclear factor erythroid 2-related factor 2, 6-OHDA = 6-hydroxydopamine, OVA = ovalbumin, PARP = poly (ADP-ribose) polymerase, PBMC = peripheral blood mononuclear cells, PD = Parkinson's disease, PDGF = platelet-derived growth factor, PGE₂ = prostaglandin E₂, PRKCA = protein kinase C alpha, PS = presenilin, RNA = ribonucleic acid, ROS = reactive oxygen species, S1P = sphingosine-1-phosphate, SP = specificity protein, SPHK = sphingosine kinase, SREBP-1 = sterol regulatory element-binding protein 1, T2D = type 2 diabetes, TG = triglyceride, TNF = tumor necrosis factor, UVB = ultraviolet B, and VEGFR = vascular endothelial growth factor receptor.

1. Introduction

Flavones are an important class of flavonoids. Found in more than 70 plant families, flavones have been reported in all plant parts including above- and below-ground, and vegetative and reproductive organs¹. Food sources of flavones are tea leaves, herbs, fruit juice, wine, honey, fruits, vegetables, cereals, and legumes².

Flavones (e.g., apigenin and luteolin) have a C₆–C₃–C₆ molecular formula consisting of a three-ring skeleton³. They differ from other flavonoids by having a C₂–C₃ double bond, a carbonyl group (ketone) at C₄ but lack a C₃ hydroxyl group at ring C²⁻⁴. In addition, most flavones have a hydroxyl group at C₅ and C₇ of ring A and/or C_{3'} and C_{4'} of ring B. Functionally, methylated flavones have better intestinal absorption and metabolic stability than non-methylated flavones⁵. Methylation also enhances their properties such as anticancer, immuno-modulation and antioxidant activities^{6,7}. Examples of methylated flavones with one methoxy group are diosmetin, acacetin and chrysoeriol^{8,9}. Examples of Polymethylated Flavones (PMFs) with five and six methoxy (OCH₃) groups are tangeretin and nobiletin, respectively¹⁰. Methylated flavones containing only one or two OCH₃ groups are metabolically more stable and superior in chemo preventive properties than PMF⁶.

Besides their functions in plant biochemistry, physiology, and ecology, flavones are important compounds for human health and nutrition. There is increasing scientific evidence for flavones having health-promoting functions such as antioxidant, antibacterial, antiviral, anti-inflammatory, anti-cancer, anti-estrogenic, anti-atherosclerotic, and anti-allergic activities^{1,3}.

In this overview, the information on the chemistry, sources, contents, and pharmacological properties of two flavones, namely, tricetin and tricetin, is updated. Their pharmacological properties are divided into anti-cancer and other pharmacological properties. Data used in the overview are procured from online databases including those of Google, Google Scholar, Science Direct, PubMed, PubMed Central, PubChem, and J-Stage.

2. Chemistry

Tricetin (TCT) or 5,7,3',4',5'-pentahydroxyflavone has a molecular formula of C₁₅H₁₀O₇ and a molecular

weight of 302.2 g/mol (Figure 1). The aglycone has two hydroxyl (OH) groups at C₅ and C₇ of ring A, and three OH groups at C_{3'}, C_{4'}, and C_{5'} of ring B. Ring C is oxygenated at position 1 and has a double bond at C₂ to C₃ and a carbonyl group (ketone) at C₄. The molecular structure of TCT is similar to myricetin except that the latter is a flavonol that bears an OH group at C₃. Rings A and B are benzene rings that form the benzoyl system and cinnamoyl system, respectively, while ring C is a heterocyclic system¹. Together, the three rings form the flavone backbone of TCT.

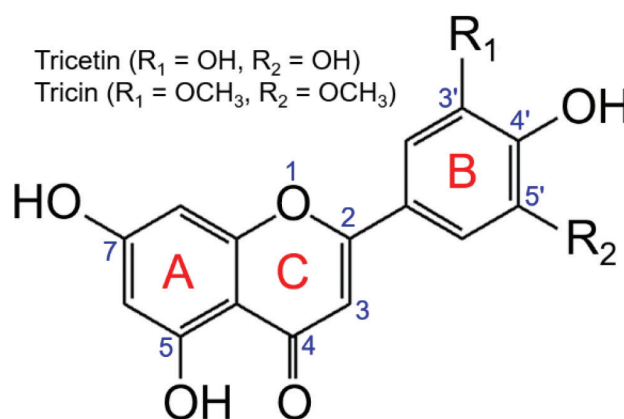


Figure 1. Molecular structure of tricetin and tricetin.

Tricin (TC) or 5,7,4'-trihydroxy-3',5'-dimethoxy flavone has a C₁₇H₁₄O₇ molecular formula and 330.3 g/mol molecular weight (Figure 1). The aglycone has two hydroxyl groups at C₅ and C₇ of ring A. There is one OH group at C_{4'}, and on both sides are two OCH₃ groups at C_{3'} and C_{5'} of ring B. Similar to TCT, ring C of TC is also oxygenated at position 1, and has a double bond at C₂ to C₃ and a carbonyl (C=O) group at C₄. TC has one OH group at C_{4'} of the B ring and two OCH₃ groups at C_{3'} and C_{5'}, i.e., at both sides of the C_{4'} OH group. This renders greater bioavailability, better metabolic stability and higher intestinal absorption to TC¹¹.

Multiple OH groups in flavonoids confer substantial antioxidant activity, and methoxy groups increase lipophilicity and improve membrane partitioning¹². A double bond and C=O group increase activity by affording more stable flavonoids. In addition, methylated flavonoids have greater metabolic stability and better permeability to cell membranes during intestinal absorption^{5,7}. The structure-activity relationship of PMF

polymethoxylated flavones showed a correlation in the pattern of methoxylation, i.e., the number/position of OCH₃ groups, and anti-proliferative activity towards cancer cells^{13,14}. Towards the inhibition of HL60 leukemic cells, an increase in the number of OCH₃ groups at ring A enhanced the activity of PMFs, whereas an increase in OCH₃ groups at ring B reduced activity¹⁴.

3. Sources

TCT occurs in honey and pollen of 27 plant species (Table 1). They belong mostly to species of the genus *Eucalyptus* (11), and the family Myrtaceae (19). Besides

honey and pollen, TCT has been reported in the leaf of *Ginkgo biloba*, the resin of *Heliotropium zeylanicum*, the aerial part of *Inga fendleriana*, the fruit juice of *Morinda citrifolia*, the whole plants of *Potentilla discolor*, flower of *Punica granatum*, and root of *Rhodiola quadrifida*. The foliage of *Eucalyptus crebra* and *Morinda citrifolia* are shown in Figure 2.

TC is found in most monocotyledon species notably the family Poaceae²⁶. The aglycone is most commonly reported in cereal crops (bran, hull, husk, grain, lignin, and leaf) such as oats, barley, rice, wheat, and corn (Table 2). It is also found in the leaf of bamboo species that include *Phyllostachys* (2)

Table 1. Plant and product sources of tricetin

S. No.	Species	Common name	Family	Source	References
1	<i>Banksia ericifolia</i>	Heath	Myrtaceae	Honey	15
2	<i>Eucalyptus camaldulensis</i>	Red river gum	Myrtaceae	Honey	16
3	<i>E. crebra</i>	Narrow-leaf ironbark	Myrtaceae	Honey	17
4	<i>E. globoidia</i>	Stringybark	Myrtaceae	Honey	17
5	<i>E. globulus</i>	Southern blue gum	Myrtaceae	Pollen	18
6	<i>E. intermedia</i>	Bloodwood	Myrtaceae	Honey	17
7	<i>E. largiflorin</i>	Black box	Myrtaceae	Honey	17
8	<i>E. melliodora</i>	Yellow box	Myrtaceae	Honey	16
9	<i>E. moluccana</i>	Gum top	Myrtaceae	Honey	17
10	<i>E. nubila</i>	Blue top ironbark	Myrtaceae	Honey	17
11	<i>E. ochrophloia</i>	Yapunyah	Myrtaceae	Honey	17
12	<i>E. pilligaensis</i>	Mallee	Myrtaceae	Honey	16
13	<i>Ginkgo biloba</i>	Maiden hair tree	Ginkgoaceae	Leaf	19
14	<i>Guioa semiglauca</i>	Crow ash	Myrtaceae	Honey	15
15	<i>Helianthus annuus</i>	Sunflower	Asteraceae	Honey	15
16	<i>Heliotropium zeylanicum</i>	Heliotrope	Boraginaceae	Resin	20
17	<i>Inga fendleriana</i>	Shimbillo	Fabaceae	Aerial part	21
18	<i>Kunzea ericoides</i>	White tea tree	Myrtaceae	Pollen	18
19	<i>Leptospermum scoparium</i>	Broom tea tree	Myrtaceae	Pollen	18
20	<i>Lophostemon conferta</i>	Brush box	Myrtaceae	Honey	15
21	<i>Melaleuca quinquenervia</i>	Tea tree	Myrtaceae	Honey	15
22	<i>Metrosideros excelsa</i>	NZ Christmas tree	Myrtaceae	Pollen	18
23	<i>M. umbellata</i>	Southern Rata	Myrtaceae	Pollen	18
24	<i>Morinda citrifolia</i>	Noni	Rubiaceae	Fruit juice	22
25	<i>Potentilla discolor</i>	Cinquifol	Rosaceae	Whole plant	23
26	<i>Punica granatum</i>	Pomegranate	Lythraceae	Flower	24
27	<i>Rhodiola quadrifida</i>	Rhodiola	Crassulaceae	Root	25



Figure 2. (L–R) *Eucalyptus crebra*, *Morinda citrifolia*, *Avena sativa*, and *Sasa albo-marginata*.

Table 2. Plant and product sources of tricetin

S. No.	Species	Common name	Family	Source	References
1	<i>Agelaea pentagyna</i>	Agelaea	Connaraceae	Leaf	27
2	<i>Avena sativa</i>	Oat	Poaceae	Bran	26
				Hull	28
				Lignin	29
3	<i>Brachypodium distachyon</i>	Brachypodium	Poaceae	Lignin	29
4	<i>Casearia arborea</i>	Gia Verde	Salicaceae	Leaf	30
5	<i>Hordeum vulgare</i>	Barley	Poaceae	Leaf and grain	26
6	<i>Oryza sativa</i>	Rice	Poaceae	Bran	31,32
				Hull and straw	29
7	<i>Phyllostachys glauca</i>	Bamboo	Poaceae	Leaf	26
8	<i>P. nigra</i>	Bamboo	Poaceae	Leaf	26,33
9	<i>Sasa albo-marginata</i>	Bamboo	Poaceae	Leaf	34-38
10	<i>S. borealis</i>	Bamboo	Poaceae	Leaf	26
11	<i>S. senanensis</i>	Bamboo	Poaceae	Leaf	26
12	<i>S. veitchii</i>	Bamboo	Poaceae	Leaf	26
13	<i>Saccharum</i> sp.	Sugarcane	Poaceae	Bagasse and stem	29
14	<i>Triticum aestivum</i>	Wheat	Poaceae	Leaf, husk and bran	26
15	<i>T. durum</i>	Wheat	Poaceae	Straw	29
16	<i>Valeriana laxiflora</i>	Valerian	Caprifoliaceae	Root	39
17	<i>Zea mays</i>	Corn	Poaceae	Leaf and stem	26,29
18	<i>Zizania latifolia</i>	Wild rice	Poaceae	Aerial part	40,41

and *Sasa* (4) with *Sasa albo-marginata* most often reported. Cereal crops include the leaf, husk, and bran of wheat (*Triticum aestivum*); leaf and grain of barley (*Hordeum vulgare*); leaf and stem of corn (*Zea mays*); bran, hull, and lignin of oat (*Avena sativa*); and bran, hull, and straw of rice (*Oryza sativa*)²⁶. The foliage of *S. albo-marginata* and *A. sativa* are shown in Figure 2.

4. Contents

The content of TCT in *Eucalyptus intermedia* and *E. ochrophloia* honey was 24.6% and 27.4% out of 60.2% and 75.3% of total flavonoids, respectively¹⁷. The content of myricetin in *E. intermedia* honey was slightly higher at 35.6%. *E. ochrophloia* honey does

not contain myricetin. The total content of flavonoids in *Melaleuca quinquenervia* honey was 6.35 mg/100 g honey with TCT amounting to 1.0 mg/100 g honey¹⁵. The content of TCT in the ethanol aerial part extract of *Inga fendleriana* was 5.88 µg/mg²¹.

Cereal crops have been reported to be a rich source of TC in dry weight. Among the grasses, the highest content of TC (33.1, 32.7, and 28.0 mg/g of lignin) was reported in oats, wheat, and brachypodium, respectively²⁹. TC was a major bioactive compound in the ethyl acetate extract of oat hull²⁸. Its content was 18 mg/kg, constituting 9.6% of the total phenolic compounds. In Manchurian wild rice, the content of TC ranged from 16.5–25.0 mg/100 g depending on the enzyme treatment and duration of extraction⁴⁰. In different plant parts of winter wheat, the content of TC was the highest in the hull (772 mg/kg) followed by the leaf (253 mg/kg) and the bran (45 mg/kg)⁴². The content of TC in different cereal crops has been quantified to be 23.6, 21.5, and 17.9 µg/g in sprouts of rice, millet, and barley⁴³, and 1006 and 454 mg/kg in straws of wheat and rice, respectively⁴⁴. A comparison between the TC content in the bran of different rice varieties showed that *Njavara* (1930 mg/kg) has the highest content, followed by *Palakkadan Matta* (120 mg/kg) and *Sujatha* (48.6 mg/kg)⁴⁵.

5. Pharmacological Properties

Recently, the pharmacological properties of TCT against cancer and diabetes have been reviewed^{46,47}. Cancers involve breast, lung, and liver cancer, including adenocarcinoma, osteosarcoma and glioblastoma, and diabetes including associated disorders such as inflammation, osteosarcoma, glioblastoma, and atherosclerosis. Bioactivities of TCT involving multi-drug resistance, antioxidant and α-glucosidase inhibition have been briefly mentioned. TC possesses anti-allergy, anti-HIV, anti-inflammatory, antioxidant, antiulcer, anti-viral, anti-diabetic, anti-obesity, anti-tyrosinase, immuno-modulatory, antibacterial, antifungal, anti-histaminic, and anti-tubercular activities^{48,49}.

5.1 Anti-cancer Activities

TCT inhibited the growth of MCF-7 breast cancer cells with an IC₅₀ value of 32.2 µM⁵⁰. Against Hep

G2 and PLC/PRF/5 liver cancer cells, IC₅₀ values of TCT were 4.87 and 4.23 µM, respectively⁵¹. Breast, liver, lung, oral, and nasopharyngeal cancer cells including glioblastoma, osteosarcoma, and leukaemia are susceptible to TCT (Table 3).

TC inhibited the proliferation of MDA-MB-468 breast, MCF-7 breast, HT-29 colon, and SW480 colon cancer cells with IC₅₀ values of 65.7, 104, 55.2, and 105 µM³¹. Against HBL100 breast and HCEC colon non-cancer cells, inhibition was 77.3 and 84.5 µM, respectively. Inhibition of colony formation by TC towards SW480 colon (16 µM) and MDA-MB-468 breast (0.6 µM) cancer cells was stronger than caffeic acid and protocatechuic acid³¹. Colon, breast, pancreas, prostate, and lung cancer cells including glioma are susceptible to TC (Table 3).

5.2 Other Pharmacological Properties

Other pharmacological properties of TCT include anti-inflammatory, anti-diabetic, poly (ADP-ribose) polymerase (PARP) inhibitory, Breast Cancer Resistance Protein (BCRP) or Adenosine Triphosphate (ATP)-binding cassette transporter G2 (ABCG2) inhibitory, Acetylcholinesterase (AChE) inhibitory, anti-gastric, neuroprotective, lipid inhibitory, and antioxidant activities (Table 4). Anti-inflammatory (5) and anti-diabetic (2) properties represent the major activities of TCT.

Other pharmacological properties of TC include anti-viral, anti-inflammatory, anti-obesity, cyclooxygenase (COX) inhibitory, anti-tubercular, skin photoaging inhibitory, anti-hepatic stellate (HS) cells, anti-tyrosinase, anti-histaminic, anti-angiogenesis, anti-diabetic, anti-leishmanial, osteoblastogenesis, acute colitis amelioration, neuroprotective, immunoregulatory, c-Jun N-terminal kinase (JNK) inhibitory, and pneumonia attenuation activities (Table 4). Anti-viral (6), anti-inflammatory (5), anti-obesity (4), and cyclooxygenase (COX) inhibitory (2) properties represent the major activities of TCT.

5.3 Structure-activity Relationship

The chemical structure of TC contains two OCH₃ groups at C3' and C5', and one OH group at C4' of ring B, i.e., at both sides of the C4' OH group (Figure 1). TCT has three OH groups at C3', C4', and C5', with no OCH₃ groups. This renders greater bioavailability,

Table 3. Anti-cancer activities of tricetin (TCT) and tricin (TC)

Compound, cancer cell line and cancer type	Effect and mechanism	References
Tricetin		
MCF-7 (breast)	TCT inhibited the growth of cancer cells by inducing cell cycle arrest and apoptosis, conveyed by activation of caspase-9, inhibition of p53-MDM2, and stabilization of p53.	50
HepG2 and PLC/PRF/5 (liver)	TCT induced apoptosis of cancer cells <i>via</i> the mitochondrial and DR5 cell death pathways and mediated by ROS generation and JNK activation.	51
H460 (lung)	TCT reversed BaP-mediated bone resorption activity of cancer cells by suppressing cancer bone metastasis.	52
HSC-3, SCC-9 and OECM-1 (oral)	TCT suppressed the migration of cancer cells by reducing MMP-9 expression and down-regulating the MAPK signalling pathway.	53
HONE-1, NPC-39 and NPC-BM (nasopharyngeal)	TCT inhibited the migration of cancer cells by down-regulating PS-1 activity and inhibiting the Akt/GSK-3 β pathway.	54
GBM 8401 and U87 (glioblastoma)	TCT suppressed the migration and/or invasion of cancer cells by inhibiting MMP-2 <i>via</i> the modulation of SP-1 expression and transcriptional activity. When combined with an ERK inhibitor (ERK-DN), a better anti-invasive effect was observed than tricetin treatment alone.	55
HOS and U2OS (osteosarcoma)	TCT (up to 80 μ M) inhibited the migration and invasion of cancer cells by down-regulation of MMP-9 expression <i>via</i> p38 and Akt signalling pathways.	56
HL-60 (leukaemia)	TCT induced apoptosis of leukaemia cells through a ROS- and JNK-mediated pathway. The anticancer activity of tricetin was enhanced when combined with an ERK inhibitor.	57
Tricin		
Adenocarcinomas (colon)	Dietary administration of mice with TC suppressed AOM/DSS-induced colon carcinogenesis by inhibiting TNF- α in the early phase, and MI and ABI in the later phase.	58
HT-29 and Colon26-Luc (colon)	TC inhibited the viability and migration of cancer cells with IC ₅₀ values of 108 and 34 μ M. Inhibition involved the down-regulation of phosphorylated Akt, ERK1/2, and NF- κ B, and by significantly suppressing cell motility, respectively.	43
MDA-MB-468 (breast)	TC inhibited the growth of cancer cells by cell cycle arrest and not by apoptosis. However, the anti-cancer effect of TC was not reflected in nude mice bearing tumor <i>in vivo</i> .	59
HepG2 (liver) and IN383/12 (pancreas)	TC was cytotoxic to cancer cells with IC ₅₀ values of 15 and 7.5 μ M, respectively.	42
PC3 (prostate)	TC potentiated the effect of docetaxel by significantly decreasing miR-21 in the treatment of cancer cells.	60
LLC cells (lung)	TC inhibited the tumor growth primarily by suppressing PRKCA/SPHK/S1P and anti-apoptotic signalling.	61
C6 (glioma)	TC inhibited the proliferation and invasion of cancer cells by up-regulation of FAK-targeting microRNA-7 in cancer cells.	62

higher metabolic stability, and better intestinal absorption to TC than TCT¹¹. In this overview, TCT has eight studies on the anti-cancer and 14 studies on other pharmacological properties, while TC is reflected by seven anti-cancer studies and 31 studies on other pharmacological properties. In terms of cytotoxicity

towards cancer cells, it is apparent that TCT is slightly stronger than CT. It appears that the greater bioavailability, higher metabolic stability, and better intestinal absorption of TC than TCT are applicable for other pharmacological but not for anti-cancer properties.

Table 4. Other pharmacological properties of tricetin (TCT) and tricrin (TC)

Compound, bioactivity	Description of effect	Reference
Tricetin		
Anti-inflammatory	TCT inhibited PARP-1 nuclear enzyme in pulmonary epithelial cells by 80%. It ranked second to myricetin which displayed 93% inhibition.	63
	TCT displayed anti-inflammatory effects in a mouse model of LPS-induced acute pulmonary inflammation. When compared to those of fisetin, its effects were less pronounced.	64
	TCT protected chondrocytes against IL-1 β -induced inflammation in rats by suppressing the MAPK signalling pathway, suggesting its potential use in treating osteoarthritis.	65
	TCT protects vascular endothelial cells from vascular inflammation by inhibiting LOX-1, ERK1/2, and Egr-1, suggesting its potential use in modulating atherosclerosis.	66
	TCT protected acinar cells against AP in mice induced by cerulein. The processes involved the suppression of apoptosis and edema formation in the pancreas, and the reduction of amylase and lipase levels in the serum.	67
Anti-diabetic	TCT displayed stronger α -glucosidase inhibitory activity than acarbose, the anti-diabetic drug. TCT with the greatest number of hydroxyl groups had the strongest α -glucosidase, α -amylase, and lipase inhibitory activities when compared to other flavones.	24
	TCT was able to reverse the poor glucose uptake ability of the hyperglycemic cell model using HepG2 cells induced with high glucose. Metformin used as the positive control, had the strongest anti-diabetic effect on glucose uptake.	68
PARP inhibitory	TCT strongly reduced LPS-induced TNF- α concentration in the blood of COPD patients (-31%) and in IL-6 concentration in the blood of T2D patients (-29%).	69
BCRP/ABCG2 inhibitory	Using the MVT assay, TCT moderately inhibited BCRP/ABCG2 with an IC ₅₀ value of 0.41 μ M.	70
AChE inhibitory	Enzyme inhibition based on the Elman assay showed that TCT strongly inhibited AChE with an IC ₅₀ value of 18.3 μ g/ml. Inhibition was stronger than donepezil (22.0 μ g/ml) used as the standard.	71
Anti-gastric	TCT strongly inhibited H ⁺ , K ⁺ -ATPase gastric enzyme with an IC ₅₀ value of 0.31 μ M. Inhibition was stronger than myricetin (0.58 μ M). Oral administration of TCT (50 mg/kg) exerted significant inhibitory effects on gastric acid secretion in mice.	72
Neuroprotective	A PD model using <i>Caenorhabditis elegans</i> showed that TCT protected against neurotoxicity induced by 6-OHDA <i>via</i> suppression of the mitochondria-dependent apoptosis pathway and activation of the Nrf2/HO-1 signalling pathway.	73
Lipid inhibitory	Unlike flavonoids such as luteolin, diosmetin, and chrysoeriol which significantly decreased lipid accumulation, TCT up-regulated the levels of intracellular lipids. Assays were done with an MDI mixture.	74
Antioxidant	Among the flavonoids, TCT was a good antioxidant with negligible pro-oxidant activity, unlike myricetin which showed both pro-oxidant and antioxidant effects.	75
Tricrin		
Anti-viral	TC significantly suppressed HCMV replication in MRC-5 HEL cells <i>via</i> inhibition of COX-2 expression with an EC ₅₀ value of 0.51 μ M.	34
	TC possessed anti-influenza virus properties, ameliorated loss in body weight and improved survival rate in mice infected with the influenza A virus.	35
	TC displayed anti-HCMV effects in MRC-5 HEL cells by inhibiting CXCL11 gene expression.	36
	TC exerted anti-HCMV activity by attenuating the expression of aCCL2 and by inhibiting HCMV virion production.	37
	TC displayed anti-HCMV effects in HEL cells by inhibiting CCL5 induction needed for the growth of the virus.	76

Table 4. Continued...

Compound, bioactivity	Description of effect	Reference
	TC significantly suppressed HCMV replication in HEL fibroblast cells by inhibiting the kinase activity of CDK9.	38
Anti-inflammatory	TC exerted anti-inflammatory effects in LPS-stimulated RAW 264.7 cells by reducing LPS-induced NO, prostaglandin E ₂ , and intracellular ROS production.	77
	TC ameliorated LPS-induced inflammation in human PBMC by modulating MAPK and PI3K/Akt pathways, down-regulating NF- κ B signaling, and deactivating COX-2 and TNF- α .	32
	TC exhibited anti-inflammatory effects in LPS-stimulated RAW 264.7 cells <i>via</i> activation of AMPK and inhibition of NF- κ B.	78
	TC suppressed inflammation-related colon carcinogenesis in mice by significantly inhibiting TNF- α expression in the colon mucosa.	79
	TC (50 μ M) exerted anti-inflammatory activity in LPS-activated RAW 264.7 cells by reducing NO production and suppressing the NF- κ B pathway.	61
Anti-obesity	TC exhibited anti-adipogenic activity by significantly inhibiting TG accumulation in 3T3-L1 adipocytes without any cytotoxic effects.	28
	TC inhibited adipogenesis and lipogenesis by suppressing fat accumulation in 3T3-L1 pre-adipocytes <i>via</i> down-regulation of Akt/mTOR/S6K and Akt/mTORC1/SREBP-1 pathways.	80,81
	TC displayed an anti-obesity effect in obese mice given a high-fat diet by lowering body weight and adipogenesis, and by decreasing serum and hepatic TG levels. The mechanism involved the AMPK pathway.	82
COX inhibitory	TC inhibited COX enzymes and reduced intestinal carcinogenesis in <i>Apc^{Min}</i> mice. In mice on the TC diet, PGE ₂ levels in the small intestinal mucosa and blood were reduced by 34% and 40%, respectively.	83
	TC inhibited COX enzymes, with IC ₅₀ values of 1.0 μ M in both HT-29 and HCA-7 colon cancer cells, respectively.	84
Anti-tubercular	TC inhibited <i>Mycobacterium tuberculosis</i> with MIC and IC ₅₀ values of 58.5 and 20.2 μ g/ml, respectively.	39
Skin photoaging inhibitory	TC attenuated UVB-induced wrinkle formation in hairless SKH-1 mice by inhibiting the expressions of MMP-1 and MMP-3.	40
Anti-HS cells	TC inhibited the proliferation of cells <i>in vitro</i> by blocking tyrosine phosphorylation of the PDGF receptor and signalling pathways, suggesting its potential use in treating hepatic fibrosis.	85
Anti-tyrosinase	TC inhibited tyrosinase activity with an IC ₅₀ value of 0.27 mg/ml in a non-competitive manner. TC also quenched tyrosinase fluorescence by forming a complex.	86
Anti-histaminic	TC possessed potent anti-histaminic activity against exocytosis from rat leukaemia basophils (IC ₅₀ value of 4.8 μ M). Luteolin and scutellarein ranked second and third with weak inhibition (IC ₅₀ values of 58 and 67 μ M) respectively.	27
Anti-angiogenesis	TC efficiently suppressed tumor angiogenesis <i>in vitro</i> by down-regulating both VEGFR2 signalling and HIF-1 α activity.	87
Anti-diabetic	TC significantly increased glucose uptake in C2C12 myotubes of glucose-loaded mice by significantly lowering blood glucose levels.	88
Anti-leishmanial	TC exhibited potent inhibitory activity against intra-cellular <i>Leishmania infantum</i> amastigotes with an IC ₅₀ value of 56 μ M, suggesting its potential use for treating leishmaniasis.	30
Osteoblastogenesis	TC enhanced osteoblastogenesis in HMS cells <i>via</i> the regulation of Wnt/ β -catenin signalling.	89
AC amelioration	TC ameliorated AC in mice induced by DSS. TC improved colonic inflammation and modulated harmful microbiota in the gut.	61
Neuroprotective	TC attenuated cerebral I/R injury in nerve cells of male rats by inhibiting autophagy, apoptosis, and inflammation <i>via</i> regulation of the PI3K/Akt pathway.	90

Table 4. Continued...

Compound, bioactivity	Description of effect	Reference
Immuno-regulatory	TC suppressed allergic responses in OVA-sensitized mice by reducing the Th2 cytokine level and increasing the Th1 cytokine level, suggesting its potential in treating allergy-related disorders.	41
Pneumonia attenuation	TC attenuated the progression of LPS-induced pneumonia by modulating Akt and MAPK signalling pathways. Against JNK1, its inhibition was 75.4% and 17.7 μ M in IC ₅₀ value.	91,92

6. Conclusion

TCT has been reported mainly in honey and pollen of *Eucalyptus* species while TC is dominant in cereal crops and bamboo species. Both flavones are equally strong in anti-cancer properties but TC has stronger other pharmacological properties than TCT. TC (5,7,4'-trihydroxy-3,5'-dimethoxyflavone) is a methylated flavone with two OCH₃ groups and three OH groups, while TCT (5,7,3,4,5'-pentahydroxyflavone) is a hydroxylated flavone five OH groups and no OCH₃ groups. Studies on the structure-activity relationship of TCT and TC by increasing and decreasing the number of OCH₃ groups, respectively, would be worthy of further investigation.

7. References

- Martens S, Mithöfer A. Flavones and flavone synthases. *Phytochemistry*. 2005; 66(20):2399-407. <https://doi.org/10.1016/j.phytochem.2005.07.013> PMID:16137727
- Hostetler GL, Ralston RA, Schwartz SJ. Flavones: Food sources, bioavailability, metabolism, and bioactivity. *Advances in Nutrition*. 2017; 8(3):423-35. <https://doi.org/10.3945/an.116.012948> PMID:28507008 PMCid:PMC5421117
- Singh M, Kaur M, Silakari O. Flavones: An important scaffold for medicinal chemistry. *European Journal of Medicinal Chemistry*. 2014; 84:206-39. <https://doi.org/10.1016/j.ejmech.2014.07.013> PMID:25019478
- Manach C, Scalbert A, Morand C, Rémésy C, Jiménez L. Polyphenols: Food sources and bioavailability. *American Journal of Clinical Nutrition*. 2004; 79(5):727-47. <https://doi.org/10.1093/ajcn/79.5.727> PMID:15113710
- Wen X, Walle T. Methylated flavonoids have greatly improved intestinal absorption and metabolic stability. *Drug Metabolism and Disposition*. 2006; 34(10):1786-92. <https://doi.org/10.1124/dmd.106.011122> PMID:16868069
- Walle T. Methoxylated flavones, a superior cancer chemopreventive flavonoid sub-class? *Seminars in Cancer Biology*. 2007; 17(5):354-62. <https://doi.org/10.1016/j.semcancer.2007.05.002> PMID:17574860 PMCid:PMC2024817
- Walle T. Methylation of dietary flavones increases their metabolic stability and chemopreventive effects. *International Journal of Molecular Sciences*. 2009; 10(11):5002-19. <https://doi.org/10.3390/ijms10115002> PMID:20087474 PMCid:PMC2808020
- Chan EWC, Ng YK, Tan CY, Alessandro L, Wong SK, Chan HT. Diosmetin and tamarixetin (methylated flavonoids): A review on their chemistry, sources, pharmacology, and anticancer properties. *Journal of Applied Pharmaceutical Science*. 2021; 11(3):22-8. <https://doi.org/10.7324/JAPS>
- Chan EWC, Wong SK, Chan HT. Acacetin and chrysoeriol: A short review of the chemistry, plant sources, bioactivities and structure-activity relationships of these methylated flavones. *Tropical Journal of Natural Product Research*. 2022; 6(1):1-7. <https://doi.org/10.26538/tjnpr/v6i1.1>
- Chan EWC, Soo OY, Tan YH, Wong SK, Chan HT. Nobiletin and tangeretin (citrus polymethoxyflavones): An overview on their chemistry, pharmacology and cytotoxic activities against breast cancer. *Journal of Chinese Pharmaceutical Sciences*. 2020; 29:443-54. <https://doi.org/10.5246/jcps.2020.07.042>
- Cai H, Boocock DJ, Steward WP, Gescher AJ. Tissue distribution in mice and metabolism in murine and human liver of apigenin and tricetin, flavones with putative cancer chemopreventive properties. *Cancer Chemotherapy and Pharmacology*. 2007; 60:257-66. <https://doi.org/10.1007/s00280-006-0368-5> PMID:17089164
- Heim KE, Tagliaferro AR, Bobilya DJ. Flavonoid antioxidants: Chemistry, metabolism and structure-activity relationships. *Journal of Nutritional Biochemistry*. 2002; 13(10):572-84. [https://doi.org/10.1016/S0955-2863\(02\)00208-5](https://doi.org/10.1016/S0955-2863(02)00208-5) PMID:12550068
- Kawaii S, Ikuina T, Hikima T, Tokiwano T, Yoshizawa Y. Relationship between structure and antiproliferative activity of polymethoxyflavones towards HL60 cells. *Anticancer Research*. 2012; 32(12):5239-44.
- Kawaii S, Ishikawa Y, Yoshizawa Y. Relationship between the structure of methoxylated and hydroxylated flavones and their antiproliferative activity in HL60 cells. *Anticancer Research*. 2018; 38(10):5679-84. <https://doi.org/10.21873/anticancer.12904> PMID:30275187
- Yao L, Jiang Y, Singanusong R, D'Arcy B, Datta N, Caffin N, et al. Flavonoids in Australian *Melaleuca*,

- Guioa, Lophostemon, Banksia and Helianthus honeys and their potential for floral authentication. *Food Research International*. 2004; 37(2):166-74. <https://doi.org/10.1016/j.foodres.2003.11.004>
16. Martos I, Ferreres F, Yao L, D'Arcy B, Caffin N, Tomás-Barberán FA. Flavonoids in monospecific *Eucalyptus* honeys from Australia. *Journal of Agricultural and Food Chemistry*. 2000; 48(10):4744-8. <https://doi.org/10.1021/jf000277i> PMID:11052728
 17. Yao L, Jiang Y, D'Arcy B, Singanusong R, Datta N, Caffin N, *et al.* Quantitative high-performance liquid chromatography analyses of flavonoids in Australian *Eucalyptus* honeys. *Journal of Agricultural and Food Chemistry*. 2004; 52(2):210-4. <https://doi.org/10.1021/jf034990u> PMID:14733497
 18. Campos MG, Webby RF, Markham KR. The unique occurrence of the flavone aglycone tricetin in Myrtaceae pollen. *Zeitschrift für Naturforschung C*. 2002; 57(9-10):944-6. <https://doi.org/10.1515/znc-2002-9-1031> PMID:12440738
 19. Liu L, Wang Y, Zhang J, Wang S. Advances in the chemical constituents and chemical analysis of *Ginkgo biloba* leaf, extract, and phytopharmaceuticals. *Journal of Pharmaceutical and Biomedical Analysis*. 2021; 193. <https://doi.org/10.1016/j.jpba.2020.113704> PMID:33157480
 20. Singh B, Sahu PM, Sharma RA. Flavonoids from *Heliotropium subulatum* exudate and their evaluation for antioxidant, antineoplastic and cytotoxic activities II. *Cytotechnology*. 2017; 69(1):103-15. <https://doi.org/10.1007/s10616-016-0041-8> PMID:27905025 PMCid:PMC5264626
 21. Pistelli L, Bertoli A, Noccioli C, Mendez J, Musmanno RA, Di Maggio T, *et al.* Antimicrobial activity of *Inga fendleriana* extracts and isolated flavonoids. *Natural Product Communications*. 2009; 4(12):1679-83. <https://doi.org/10.1177/1934578X0900401214>
 22. Lee D, Yu JS, Huang P, Qader M, Manavalan A, Wu X, *et al.* Identification of anti-inflammatory compounds from Hawaiian noni (*Morinda citrifolia* L.) fruit juice. *Molecules*. 2020; 25(21):4968-79. <https://doi.org/10.3390/molecules25214968> PMID:33121016 PMCid:PMC7662328
 23. Wang N, Zhu F, Shen M, Qiu L, Tang M, Xia H, *et al.* Network pharmacology-based analysis on bioactive anti-diabetic compounds in *Potentilla discolor* Bunge. *Journal of Ethnopharmacology*. 2019; 241. <https://doi.org/10.1016/j.jep.2019.111905> PMID:31022565
 24. Wu S, Tian L. A new flavone glucoside together with known ellagitannins and flavones with anti-diabetic and anti-obesity activities from the flowers of pomegranate (*Punica granatum*). *Natural Product Research*. 2019; 33(2):252-7. <https://doi.org/10.1080/14786419.2018.1446009> PMID:29502447
 25. Yoshikawa M, Shimada H, Shimoda H, Murakami N, Yamahara J, Matsuda H. Bioactive constituents of Chinese natural medicines. II. *Rhodiola radix*. (1). Chemical structures and antiallergic activity of rhodiocyanosides A and B from the underground part of *Rhodiola quadrifida* (Pall.) Fisch. *et Mey.* (Crassulaceae). *Chemical and Pharmaceutical Bulletin*. 1996; 44(11):2086-91. <https://doi.org/10.1248/cpb.44.2086> PMID:8945774
 26. Li M, Pu Y, Yoo CG, Ragauskas AJ. The occurrence of tricetin and its derivatives in plants. *Green Chemistry*. 2016; 18(6):1439-54. <https://doi.org/10.1039/C5GC03062E>
 27. Kuwabara H, Mouri K, Otsuka H, Kasai R, Yamasaki K. Tricetin from a Malagasy Connaraceae plant with potent antihistaminic activity. *Journal of Natural Products*. 2003; 66(9):1273-5. <https://doi.org/10.1021/np030020p> PMID:14510616
 28. Lee D, Park HY, Kim S, Park Y, Bang MH, Imm JY. Anti-adipogenic effect of oat hull extract containing tricetin on 3T3-L1 adipocytes. *Process Biochemistry*. 2015; 50(12):2314-21. <https://doi.org/10.1016/j.procbio.2015.09.019>
 29. Lan W, Rencoret J, Lu F, Karlen SD, Smith BG, Harris PJ, *et al.* Tricin-lignins: Occurrence and quantitation of tricetin in relation to phylogeny. *Plant Journal*. 2016; 88(6): 1046-57. <https://doi.org/10.1111/tpj.13315> PMID:27553717
 30. Santos AL, Yamamoto ES, Passero LF, Laurenti MD, Martins LF, Lima ML, *et al.* Anti-leishmanial activity and immunomodulatory effects of tricetin isolated from leaves of *Casearia arborea* (Salicaceae). *Chemistry and Biodiversity*. 2017; 14(5). <https://doi.org/10.1002/cbdv.201600458> PMID:28054741
 31. Hudson EA, Dinh PA, Kokubun T, Simmonds MS, Gescher A. Characterization of potentially chemopreventive phenols in extracts of brown rice that inhibit the growth of human breast and colon cancer cells. *Cancer Epidemiology, Biomarkers and Prevention*. 2000; 9(11):1163-70.
 32. Shalini V, Pushpan CK, Sindhu G, Jayalekshmy A, Helen A. Tricetin, flavonoid from Njavara reduces inflammatory responses in hPBMCs by modulating the p38MAPK and PI3K/Akt pathways and prevents inflammation associated endothelial dysfunction in HUVECs. *Immunobiology*. 2016; 221(2):137-44. <https://doi.org/10.1016/j.imbio.2015.09.016> PMID:26514297
 33. Jiao J, Zhang Y, Liu C, Liu JE, Wu X, Zhang Y. Separation and purification of tricetin from an antioxidant product derived from bamboo leaves. *Journal of Agricultural and Food Chemistry*. 2007; 55(25):10086-92. <https://doi.org/10.1021/jf0716533> PMID:18001030
 34. Akuzawa K, Yamada R, Li Z, Li Y, Sadanari H, Matsubara K, *et al.* Inhibitory effects of tricetin derivative from *Sasa albo-marginata* on replication of human cytomegalovirus. *Antiviral Research*. 2011; 91(3):296-303. <https://doi.org/10.1016/j.antiviral.2011.06.014> PMID:21745500
 35. Yazawa K, Kurokawa M, Obuchi M, Li Y, Yamada R, Sadanari H, *et al.* Anti-influenza virus activity of tricetin, 4',5,7-trihydroxy-3',5'-dimethoxyflavone. *Antiviral Chemistry and Chemotherapy*. 2011; 22(1):1-11. <https://doi.org/10.3851/IMP1782> PMID:21860068

36. Murayama T, Li Y, Takahashi T, Yamada R, Matsubara K, Tsuchida Y, *et al.* Anti-cytomegalovirus effects of tricrin are dependent on CXCL11. *Microbes and Infection*. 2012; 14(12):1086-92. <https://doi.org/10.1016/j.micinf.2012.05.017> PMID:22683667
37. Akai Y, Sadanari H, Takemoto M, Uchide N, Daikoku T, Mukaida N, *et al.* Inhibition of human cytomegalovirus replication by tricrin is associated with depressed CCL2 expression. *Antiviral Research*. 2017; 148:15-9. <https://doi.org/10.1016/j.antiviral.2017.09.018> PMID:28965916
38. Sadanari H, Fujimoto KJ, Sugihara Y, Ishida T, Takemoto M, Daikoku T, *et al.* The anti-human cytomegalovirus drug tricrin inhibits cyclin-dependent kinase 9. *FEBS Open Bio*. 2018; 8(4):646-54. <https://doi.org/10.1002/2211-5463.12398> PMID:29632816 PMCID:PMC5881553
39. Gu JQ, Wang Y, Franzblau SG, Montenegro G, Yang D, Timmermann BN. Anti-tubercular constituents of *Valeriana laxiflora*. *Planta Medica*. 2004; 70(6):509-14. <https://doi.org/10.1055/s-2004-827149> PMID:15229801
40. Moon JM, Park SH, Jhee KH, Yang SA. Protection against UVB-induced wrinkle formation in SKH-1 hairless mice: Efficacy of tricrin isolated from enzyme-treated *Zizania latifolia* extract. *Molecules*. 2018; 23(9):2254-66. <https://doi.org/10.3390/molecules27133978> PMID:19223822 PMCID:PMC6254026
41. Lee JY, Park SH, Jhee KH, Yang SA. *Zizania latifolia* and its major compound tricrin regulate immune responses in OVA-treated mice. *Molecules*. 2022; 27(13):3978-90. <https://doi.org/10.3390/molecules27133978> PMID:35807220 PMCID:PMC9268014
42. Moheb A, Grondin M, Ibrahim RK, Roy R, Sarhan F. Winter wheat hull (husk) is a valuable source for tricrin, a potential selective cytotoxic agent. *Food Chemistry*. 2013; 138(2-3):931-7. <https://doi.org/10.1016/j.foodchem.2012.09.129> PMID:23411198
43. Yue GG, Gao S, Lee JK, Chan YY, Wong EC, Zheng T, *et al.* A natural flavone tricrin from grains can alleviate tumor growth and lung metastasis in colorectal tumor mice. *Molecules*. 2020; 25(16):3730-45. <https://doi.org/10.3390/molecules25163730> PMID:32824166 PMCID:PMC7463810
44. Zheng T, Wong EC, Yue GG, Li XX, Wu KH, Lau DT, *et al.* Identification and quantification of tricrin present in medicinal herbs, plant foods and by-products using UPLC-QTOF-MS. *Chemical Papers*. 2021; 75(9):4579-88. <https://doi.org/10.1007/s11696-021-01651-6>
45. Mohanlal S, Parvathy R, Shalini V, Helen A, Jayalekshmy A. Isolation, characterization and quantification of tricrin and flavonolignans in the medicinal rice Njavara (*Oryza sativa* L.), as compared to staple varieties. *Plant Foods for Human Nutrition*. 2011; 66:91-6. <https://doi.org/10.1007/s11130-011-0217-5> PMID:21373805
46. Patel DK. Potential benefits of tricetin in medicine for the treatment of cancers and other health-related disorders: Medicinal importance and therapeutic benefit. *Journal of Natural Products*. 2022; 12(6):12-9. <https://doi.org/10.2174/2210315512666211221113117>
47. Patel DK, Patel K. Biological potential of tricetin on diabetes disease: Pharmacological approaches in the medicine through scientific data analysis. *Metabolism: Clinical and Experimental*. 2022; 128. <https://doi.org/10.1016/j.metabol.2021.155052>
48. Zhou JM, Ibrahim RK. Tricrin – A potential multi-functional nutraceutical. *Phytochemistry Reviews*. 2010; 9(3):413-24. <https://doi.org/10.1007/s11101-009-9161-5>
49. Jiang B, Song J, Jin Y. A flavonoid monomer tricrin in gramineous plants: Metabolism, bio/chemosynthesis, biological properties, and toxicology. *Food Chemistry*. 2020; 320. <https://doi.org/10.1016/j.foodchem.2020.126617> PMID:32247167
50. Hsu YL, Uen YH, Chen Y, Liang HL, Kuo PL. Tricetin, a dietary flavonoid, inhibits proliferation of human breast adenocarcinoma MCF-7 cells by blocking cell cycle progression and inducing apoptosis. *Journal of Agricultural and Food Chemistry*. 2009; 57(18):8688-95. <https://doi.org/10.1021/jf901053x> PMID:19705844
51. Hsu YL, Hou MF, Tsai EM, Kuo PL. Tricetin, a dietary flavonoid, induces apoptosis through the reactive oxygen species/c-Jun NH2-terminal kinase pathway in human liver cancer cells. *Journal of Agricultural and Food Chemistry*. 2010; 58(23):12547-56. <https://doi.org/10.1021/jf103159r> PMID:21067180
52. Hung JY, Chang WA, Tsai YM, Hsu YL, Chiang HH, Chou SH, *et al.* Tricetin, a dietary flavonoid, suppresses benzo(a)pyrene induced human non small cell lung cancer bone metastasis. *International Journal of Oncology*. 2015; 46(5):1985-93. <https://doi.org/10.3892/ijo.2015.2915> PMID:25738754
53. Chung TT, Chuang CY, Teng YH, Hsieh MJ, Lai JC, Chuang YT, *et al.* Tricetin suppresses human oral cancer cell migration by reducing matrix metalloproteinase-9 expression through the mitogen-activated protein kinase signalling pathway. *Environmental Toxicology*. 2017; 32(11):2392-9. <https://doi.org/10.1002/tox.22452> PMID:28731287
54. Ho HY, Lin FC, Chen PN, Chen MK, Hsin CH, Yang SF, *et al.* Tricetin suppresses migration and presenilin-1 expression of nasopharyngeal carcinoma through Akt/GSK-3 β pathway. *American Journal of Chinese Medicine*. 2020; 48(5):1203-20. <https://doi.org/10.1142/S0192415X20500597> PMID:32668971
55. Chao R, Chow JM, Hsieh YH, Chen CK, Lee WJ, Hsieh FK, *et al.* Tricetin suppresses the migration/invasion of human glioblastoma multiforme cells by inhibiting matrix metalloproteinase-2 through modulation of the expression and transcriptional activity of specificity protein 1. *Expert Opinion on Therapeutic Targets*. 2015; 19(10):1293-306. <https://doi.org/10.1517/14728222.2015.1075509> PMID:26245494
56. Chang PY, Hsieh MJ, Hsieh YS, Chen PN, Yang JS, Lo FC, *et al.* Tricetin inhibits human osteosarcoma cells metastasis

- by transcriptionally repressing MMP-9 via p38 and Akt pathways. *Environmental Toxicology*. 2017; 32(8):2032-40. <https://doi.org/10.1002/tox.22380> PMID:27860196
57. Chien MH, Chow JM, Lee WJ, Chen HY, Tan P, Wen YC, *et al.* Tricetin induces apoptosis of human leukemic HL-60 cells through a reactive oxygen species-mediated c-Jun N-terminal kinase activation pathway. *International Journal of Molecular Sciences*. 2017; 18(8):1667-81. <https://doi.org/10.3390/ijms18081667> PMID:28758971 PMCID:PMC5578057
58. Oyama T, Yasui Y, Sugie S, Koketsu M, Watanabe K, Tanaka T. Dietary tricetin suppresses inflammation-related colon carcinogenesis in male Crj: CD-1 mice. *Cancer Prevention Research*. 2009; 2(12):1031-8. <https://doi.org/10.1158/1940-6207.CAPR-09-0061> PMID:19934339
59. Cai H, Hudson EA, Mann P, Verschoyle RD, Greaves P, Manson MM, *et al.* Growth-inhibitory and cell cycle-arresting properties of the rice bran constituent tricetin in human-derived breast cancer cells *in vitro* and in nude mice *in vivo*. *British Journal of Cancer*. 2004; 91(7):1364-71. <https://doi.org/10.1038/sj.bjc.6602124> PMID:15316567 PMCID:PMC2410014
60. Ghasemi S, Lorigooini Z, Wibowo J, Amini-Khoei H. Tricetin isolated from *Allium atrovioleaceum* potentiated the effect of docetaxel on PC3 cell proliferation: Role of miR-21. *Natural Product Research*. 2019; 33(12):1828-31. <https://doi.org/10.1080/14786419.2018.1437439> PMID:29447469
61. Li XX, Chen SG, Yue GG, Kwok HF, Lee JK, Zheng T, *et al.* Natural flavone tricetin exerted anti-inflammatory activity in macrophage *via* NF- κ B pathway and ameliorated acute colitis in mice. *Phytomedicine*. 2021; 90. <https://doi.org/10.1016/j.phymed.2021.153625> PMID:34256329
62. Chung DJ, Wang CJ, Yeh CW, Tseng TH. Inhibition of the proliferation and invasion of C6 glioma cells by tricetin *via* the upregulation of focal-adhesion-kinase-targeting microRNA-7. *Journal of Agricultural and Food Chemistry*. 2018; 66(26):6708-16. <https://doi.org/10.1021/acs.jafc.8b00604> PMID:29877083
63. Geraets L, Moonen HJ, Brauers K, Wouters EF, Bast A, Hageman GJ. Dietary flavones and flavonols are inhibitors of poly (ADP-ribose) polymerase-1 in pulmonary epithelial cells. *Journal of Nutrition*. 2007; 137(10):2190-5. <https://doi.org/10.1093/jn/137.10.2190> PMID:17884996
64. Geraets L, Haegens A, Brauers K, Haydock JA, Vernooij JH, Wouters EF, *et al.* Inhibition of LPS-induced pulmonary inflammation by specific flavonoids. *Biochemical and Biophysical Research Communications*. 2009; 382(3):598-603. <https://doi.org/10.1016/j.bbrc.2009.03.071> PMID:19292976
65. Sun FF, Hu PF, Xiong Y, Bao JP, Qian J, Wu LD. Tricetin protects rat chondrocytes against IL-1 β -induced inflammation and apoptosis. *Oxidative Medicine and Cellular Longevity*. 2019. <https://doi.org/10.1155/2019/4695381> PMID:31231454 PMCID:PMC6512055
66. Cai L, Zhang X, Hou M, Gao F. Natural flavone tricetin suppresses oxidized LDL-induced endothelial inflammation mediated by Egr-1. *International Immunopharmacology*. 2020; 80. <https://doi.org/10.1016/j.intimp.2020.106224> PMID:31991371
67. Nagy-Pénczes M, Hajnády Z, Regdon Z, Demény MÁ, Kovács K, El-Hamoly T, *et al.* Tricetin reduces inflammation and acinar cell injury in cerulein-induced acute pancreatitis: The role of oxidative stress-induced DNA damage signalling. *Biomedicines*. 2022; 10(6):1371-91. <https://doi.org/10.3390/biomedicines10061371> PMID:35740393 PMCID:PMC9219693
68. Wang N, Zhu F, Shen M, Qiu L, Tang M, Xia H, *et al.* Network pharmacology-based analysis on bioactive anti-diabetic compounds in *Potentilla discolor* Bunge. *Journal of Ethnopharmacology*. 2019; 241. <https://doi.org/10.1016/j.jep.2019.111905> PMID:31022565
69. Weseler AR, Geraets L, Moonen HJ, Manders RJ, van Loon LJ, Pennings HJ, *et al.* Poly (ADP-ribose) polymerase-1 inhibiting flavonoids attenuate cytokine release in blood from male patients with chronic obstructive pulmonary disease or Type 2 diabetes. *Journal of Nutrition*. 2009; 139(5):952-57. <https://doi.org/10.3945/jn.108.102756> PMID:19321592
70. Tan KW, Li Y, Paxton JW, Birch NP, Scheepens A. Identification of novel dietary phytochemicals inhibiting the efflux transporter breast cancer resistance protein (BCRP/ABCG2). *Food Chemistry*. 2013; 138:2267-74. <https://doi.org/10.1016/j.foodchem.2012.12.021> PMID:23497885
71. Kuppusamy A, Arumugam M, George S. Combining *in silico* and *in vitro* approaches to evaluate the acetylcholinesterase inhibitory profile of some commercially available flavonoids in the management of Alzheimer's disease. *International Journal of Biological Macromolecules*. 2017; 95: 199-203. <https://doi.org/10.1016/j.ijbiomac.2016.11.062> PMID:27871793
72. Miyazaki Y, Ichimura A, Sato S, Fujii T, Oishi S, Sakai H, *et al.* The natural flavonoid myricetin inhibits gastric H⁺,K⁺-ATPase. *European Journal of Pharmacology*. 2018; 820:217-21. <https://doi.org/10.1016/j.ejphar.2017.12.042> PMID:29274333
73. Ren J, Yuan L, Wang W, Zhang M, Wang Q, Li S, *et al.* Tricetin protects against 6-OHDA-induced neurotoxicity in Parkinson's disease model by activating Nrf2/HO-1 signalling pathway and preventing the mitochondria-dependent apoptosis pathway. *Toxicology and Applied Pharmacology*. 2019; 378. <https://doi.org/10.1016/j.taap.2019.114617> PMID:31176653
74. Nishina A, Ukiya M, Fukatsu M, Koketsu M, Ninomiya M, Sato D, *et al.* Effects of various 5,7-dihydroxyflavone analogs on adipogenesis in 3T3-L1 cells. *Biological and Pharmaceutical Bulletin*. 2015; 38(11):1794-800. <https://doi.org/10.1248/bpb.b15-00489> PMID:26521830
75. Chobot V, Hadacek F, Bachmann G, Weckwerth W, Kubicova L. *In vitro* evaluation of pro- and antioxidant effects of flavonoid tricetin in comparison to myricetin.

- Molecules. 2020; 25(24):5850-61. <https://doi.org/10.3390/molecules25245850> PMID:33322312 PMCID:PMC7768484
76. Itoh A, Sadanari H, Takemoto M, Matsubara K, Daikoku T, Murayama T. Tricin inhibits the CCL5 induction required for efficient growth of human cytomegalovirus. *Microbiology and Immunology*. 2018; 62(5):341-7. <https://doi.org/10.1111/1348-0421.12590> PMID:29603339
77. Kang BM, An BK, Jung WS, Jung HK, Cho JH, Cho HW, *et al*. Anti-inflammatory effect of tricin isolated from *Alopecurus aequalis* Sobol. on the LPS-induced inflammatory response in RAW 264.7 cells. *International Journal of Molecular Medicine*. 2016; 38(5):1614-20. <https://doi.org/10.3892/ijmm.2016.2765> PMID:28025993
78. Lee D, Imm JY. AMP kinase activation and inhibition of nuclear factor-kappa B (NF- κ B) translocation contribute to the anti-inflammatory effect of tricin. *Journal of Food Biochemistry*. 2017; 41(2). <https://doi.org/10.1111/jfbc.12293>
79. Tanaka T, Oyama T, Sugie S. Dietary tricin suppresses inflammation-related colon carcinogenesis in mice. *Journal of Nutritional Science and Vitaminology*. 2019; 65:100-3. <https://doi.org/10.3177/jnsv.65.S100> PMID:31619605
80. Lee D, Go GW, Imm JY. Tricin, a methylated cereal flavone, suppresses fat accumulation by down-regulating AKT and mTOR in 3T3-L1 pre-adipocytes. *Journal of Functional Foods*. 2016; 26:548-56. <https://doi.org/10.1016/j.jff.2016.08.023>
81. Lee D, Imm JY. Tricin, a methylated cereal flavone suppresses fat accumulation through AKT-mTORC1-SREBP1 pathway in 3T3-L1 preadipocytes. *FASEB Journal*. 2016; 30. https://doi.org/10.1096/fasebj.30.1_supplement.lb281
82. Lee D, Imm JY. Anti-obesity effect of tricin, a methylated cereal flavone, in high-fat-diet-induced obese mice. *Journal of Agricultural and Food Chemistry*. 2018; 66(38):9989-94. <https://doi.org/10.1021/acs.jafc.8b03312> PMID:30173509
83. Cai H, Al-Fayez M, Tunstall RG, Platton S, Greaves P, Steward WP, *et al*. The rice bran constituent tricin potently inhibits cyclooxygenase enzymes and interferes with intestinal carcinogenesis in ApcMin mice. *Molecular Cancer Therapeutics*. 2005; 4(9):1287-92. <https://doi.org/10.1158/1535-7163.MCT-05-0165> PMID:16170019
84. Al-Fayez M, Cai H, Tunstall R, Steward WP, Gescher AJ. Differential modulation of cyclooxygenase-mediated prostaglandin production by the putative cancer chemopreventive flavonoids tricin, apigenin and quercetin. *Cancer Chemotherapy and Pharmacology*. 2006; 58(6):816-25. <https://doi.org/10.1007/s00280-006-0228-3> PMID:16552572
85. Seki N, Toh U, Kawaguchi K, Ninomiya M, Koketsu M, Watanabe K, *et al*. Tricin inhibits proliferation of human hepatic stellate cells *in vitro* by blocking tyrosine phosphorylation of PDGF receptor and its signalling pathways. *Journal of Cellular Biochemistry*. 2012; 113(7):2346-55. <https://doi.org/10.1002/jcb.24107> PMID:22359269
86. Mu Y, Li L, Hu SQ. Molecular inhibitory mechanism of tricin on tyrosinase. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*. 2013; 107:235-40. <https://doi.org/10.1016/j.saa.2013.01.058> PMID:23434549
87. Han JM, Kwon HJ, Jung HJ. Tricin, 4',5,7-trihydroxy-3',5'-dimethoxyflavone, exhibits potent antiangiogenic activity *in vitro*. *International Journal of Oncology*. 2016; 49(4):1497-504. <https://doi.org/10.3892/ijo.2016.3645> PMID:27498749
88. Kim S, Go GW, Imm JY. Promotion of glucose uptake in C2C12 myotubes by cereal flavone tricin and its underlying molecular mechanism. *Journal of Agricultural and Food Chemistry*. 2017; 65(19):3819-26. <https://doi.org/10.1021/acs.jafc.7b00578> PMID:28474889
89. Zhang H, Li H. Tricin enhances osteoblastogenesis through the regulation of Wnt/ β -catenin signalling in human mesenchymal stem cells. *Mechanisms of Development*. 2018; 152:38-43. <https://doi.org/10.1016/j.mod.2018.07.001> PMID:30056839
90. Liu Y, Qu X, Yan M, Li D, Zou R. Tricin attenuates cerebral ischemia/reperfusion injury through inhibiting nerve cell autophagy, apoptosis and inflammation by regulating the PI3K/Akt pathway. *Human and Experimental Toxicology*. 2022; 41:1-10. <https://doi.org/10.1177/09603271221125928> PMID:36113040
91. Yang R, Zhao G, Yan B. Discovery of novel c-jun N-terminal kinase 1 inhibitors from natural products: Integrating artificial intelligence with structure-based virtual screening and biological evaluation. *Molecules*. 2022; 27(19):6249-66 <https://doi.org/10.3390/molecules> PMID:19223822 PMCID:PMC6254026
92. Yang F, Liu W. Tricin attenuates the progression of LPS-induced severe pneumonia in bronchial epithelial cells by regulating AKT and MAPK signalling pathways. *Allergologia et Immunopathologia*. 2022; 50(3):113-8. <https://doi.org/10.15586/aei.v50i3.587> PMID:35527664